Review

Infections in the Etiology of Parkinson's Disease and Synucleinopathies: A Renewed Perspective, Mechanistic Insights, and Therapeutic Implications

Gabriela Mercado^a, Christopher Kaeufer^{b,c}, Franziska Richter^c and Wouter Peelaerts^{d,*} ^aDivision of Neurobiology, Department of Psychiatry, Johns Hopkins University School of Medicine, Baltimore, MD, USA ^bCenter for Systems Neuroscience, Hannover, Germany ^cDepartment of Pharmacology, Toxicology and Pharmacy, University of Veterinary Medicine Hannover, Hannover, Germany ^dLaboratory for Virology and Gene Therapy, Department of Pharmacy and Pharmaceutical Sciences, KU Leuven, Leuven, Belgium

Accepted 6 September 2024 Pre-press 25 September 2024 Published 15 October 2024

Abstract. Increasing evidence suggests a potential role for infectious pathogens in the etiology of synucleinopathies, a group of age-related neurodegenerative disorders including Parkinson's disease (PD), multiple system atrophy and dementia with Lewy bodies. In this review, we discuss the link between infections and synucleinopathies from a historical perspective, present emerging evidence that supports this link, and address current research challenges with a focus on neuroinflammation. Infectious pathogens can elicit a neuroinflammatory response and modulate genetic risk in PD and related synucleinopathies. The mechanisms of how infections might be linked with synucleinopathies as well as the overlap between the immune cellular pathways affected by virulent pathogens and disease-related genetic risk factors are discussed. Here, an important role for α -synuclein in the immune response against infections is emerging. Critical methodological and knowledge gaps are addressed, and we provide new future perspectives on how to address these gaps. Understanding how infections and neuroinflammation influence synucleinopathies will be essential for the development of early diagnostic tools and novel therapies.

Plain Language Summary

This review explores how infections might contribute to the development of Parkinson's disease and other synucleinopathies. It highlights evidence that microbial pathogens may trigger neurodegeneration by causing neuroinflammation. We emphasize

^{*}Correspondence to: Wouter Peelaerts, Laboratory for Virology and Gene Therapy, Department of Pharmacy and Pharmaceutical Sciences, KU Leuven, Leuven, Belgium. Tel.: +32 16 37 40 59; E-mail: wouter.peelaerts@kuleuven.be.

ISSN 1877-7171 © 2024 – The authors. Published by IOS Press. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (CC BY-NC 4.0).

the complex relationship between infections, genetics, and neurodegeneration, and discuss how understanding these connections could lead to earlier diagnosis and new treatments. In this review we also identify key knowledge gaps, and we suggest areas for future research.

Keywords: Parkinson's disease, multiple system atrophy, infections, α-synuclein

INTRODUCTION

Age-related neurodegenerative diseases such as synucleinopathies are likely caused by a complex interplay between host genetics and the environment. Within this environment, there is a lifelong accumulation of environmental exposure to toxicants and pathogens. The totality of environmental exposure (the exposome), lifestyle and demographics might be crucial determinants in the development of neurodegeneration.¹

Synucleinopathies are a category of age-related progressive neurodegenerative diseases, characterized by the intracellular accumulation of alphasynuclein (α Syn) containing aggregates.² This group includes Parkinson's disease (PD), dementia with Lewy Bodies (DLB), multiple system atrophy (MSA), and other atypical forms.^{3,4} Historical epidemiological studies have often suggested an infectious etiology of parkinsonism. Now, there is renewed attention for a possible infectious etiology as recent studies show that virulent agents can act as triggers of synucleinopathy, and that genetic risk modulates the pathogenic response to inflammatory or infectious agents.⁵

Several bacterial, viral, and fungal pathogens are associated with the onset of sporadic cases of synucleinopathy.⁵ However, whether infections causally contribute to the pathogenesis of synucleinopathies or help to exacerbate existing pathology remains to be established. This rapid evolving field of research has recently presented evidence for the involvement of α Syn and other PD risk factors in the innate and adaptive immune response. Specifically, it is shown that the host response against infections is mediated via α Syn. α Syn is a pre-synaptic protein with high expression in the brain, but also functions as a key protein in the response to pathogenic and commensal microorganisms.

Here, we summarize epidemiological and experimental observations linking infections with synucleinopathies. We propose that infections can trigger inflammatory processes that could contribute to α Syn pathological aggregation and associated neurodegeneration. We also explore the possibility that the pathological agent, the infection site, and the host genetic background could affect α Syn protein conformation and function, and as consequence, the neurodegenerative process.

EPIDEMIOLOGICAL EVIDENCE

An infectious etiology of parkinsonism

An infectious etiology for synucleinopathies has been discussed for decades. Early observations of viral epidemics followed by an outbreak of parkinsonism have led to a long-held hypothesis that parkinsonism could be of infectious origin, and it has long been discussed whether viruses could be the initiating etiology of primary PD or if viruses could be causative for secondary parkinsonism.

A notable example and a historic conception of a potential infectious etiology of PD is the postencephalitic parkinsonism pandemic in the years following the Spanish flu. The Spanish flu, caused by a severely virulent H1N1 influenza A virus, affected millions of people worldwide and resulted in a global pandemic. After the outbreak of the Spanish flu, in 1918–1919, many patients subsequently developed encephalitis lethargica, a condition that is characterized by high fever, delayed physical and mental response and lethargy.^{6,7} Although these symptoms from encephalitis lethargica only occurred sometime after infection it was clear that the onset pattern of encephalitis lethargica inherently followed the one of the influenza pandemics.

During this early phase of encephalitis lethargica, other neurological manifestations concurrently developed as patients experienced phases of delirium or episodes of psychotic manifestations.⁸ Encephalitis lethargica could almost be seen as a precursor of neurological disease as a bout of encephalitis was almost invariably followed by postencephalitic parkinsonism.⁸ Here, the classical signs of parkinsonism would include a mask-face like appearance,

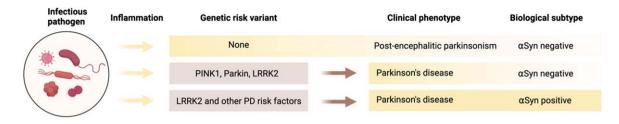


Fig. 1. A role for infections in the etiology of synucleinopathies. The environmental exposure to infectious pathogens can trigger inflammatory cellular pathways involved in disease pathogenesis. Parkinsonism following post-encephalitic infection with viruses arises from neuroinflammatory processes that cause degeneration of dopaminergic neurons in the substantia nigra. Genetic risk variants in mitochondrial or lysosomal genes (PINK1, Parkin, and LRRK2) may exacerbate or precipitate motor and non-motor symptoms by modulating immune responses to infections. This happens via a process that is independent of α Syn. For people with PD that are α Syn negative, PD is likely not a synucleinopathy. Here, the biology of disease would be fundamentally different from α Syn-positive PD. In contrast, PD-related genetic risk factors can lead to increased α Syn aggregation and exaggerated immune reactions, resulting in enhanced inflammatory and neuroinflammatory responses leading to neuronal damage and the onset of new symptoms. For people with PD that are α Syn positive, PD is a synucleinopathy. Interactions between genetic and inflammatory pathogens contribute to the onset and progression of different subtypes of PD, emphasizing the complex biology and the interplay between genetics, environmental insults, and neuroinflammatory pathways.

paralysis agitans, tremor, rigidity of all the muscles, and shuffling gait.⁷ These symptoms could develop in all infected individuals, including young adults.⁹ In the following decades, it was thus speculated that a link between viral respiratory infections and neurological diseases such as PD might exist.¹⁰ However, these links became muddled because of a profusion of clinical signs and the long time between infection and the onset of neurological symptoms in which the association between parkinsonism and influenza was simply forgotten.

Nevertheless, pathological examinations of patients with postencephalitic parkinsonism, some of which were done more recently, 1^{1-14} revealed that PD-related brain regions of patients infected with the Spanish flu were indeed severely affected. In infected cases, a remarkable hyaline degeneration with loss of blood vessels was observed in the striatum and more generally in the basal ganglia,^{6,15} Vascular subacute inflammation was sometimes seen, especially in the brainstem, but in the absence of signs of chronic or active neuroinflammation elsewhere in the brain¹⁶. From these historic cases, the most notably affected region was the substantia nigra, a region typically affected in synucleinopathies, showing loss of pigmented dopaminergic neurons and gliosis.^{11,12,17} Maybe most striking was that this neurodegeneration within the substantia nigra and the basal ganglia was accompanied by the presence of rounded or oval viral inclusions bodies but most often there were numerous cellular neurofibrillary tangles.^{11–14,18} Importantly, the presence of α Syn positive inclusions in these cases was investigated, with recent immunohistochemical techniques and

 α Syn-specific antibodies, but α Syn pathology could not be shown.¹¹⁻¹³ In cases of postencephalitic parkinsonism, and in the absence of synucleinopathy, it seems that based on these findings, there is more evidence for a tauopathy-related type of parkinsonism after influenza infection rather than predominantly synucleinopathy-related PD. It also suggests that based on its underlying biological mechanisms, postencephalitic parkinsonism might represent a unique entity in the biological classification of PD (Fig. 1) (we further discuss other implications of infections on the clinical classifications of PD below).

The development and onset of neurological symptoms after influenza infection varied. The onset of parkinsonism after influenza infection ranged from days to a more prolonged onset with cases that would show neurological symptoms not until decades after infection⁸. Symptoms also developed when the infected person was considered to have fully recovered from the infection. The establishment of infection-related encephalitis lethargica or postencephalitic parkinsonism as a clinical entity thus rests on the abundant epidemiological, clinical, and pathological descriptions during that time. Of note, the delayed onset of neurological symptoms is in line with the natural history of PD, where a long prodromal phase precedes the occurrence of the more typical cardinal symptoms.

Along those lines, case reports of infection with other viruses have also been reported to cause postencephalitic dopamine-responsive parkinsonism with alterations in the basal ganglia and neurodegeneration of the substantia nigra.^{19–21} Several of these case reports include infections with SARS-CoV-2,^{22–25} enterovirus,^{26–29} Japanese encephalitis virus,^{30–32} St. Louis encephalitis,³³ West Nile virus,^{34–37} HIV,^{38–40} and dengue viral infections.^{19,41} Importantly, this spectrum of viruses demonstrates that development of neurological symptoms and brain pathology after infection does not require the virus to enter brain tissue or to replicate in neurons as some of these viruses are not neurotropic.

An infectious etiology of idiopathic Parkinson's disease

Although viral post-encephalitic cases show many clinical similarities with PD, these cases of viral encephalitis with parkinsonism are etiologically distinct from idiopathic PD. The mechanisms leading to selective degeneration of the basal ganglia and the substantia nigra in the absence of overt synucleinopathy are in stark contrast to the natural history of PD and the progression of pathology in idiopathic cases.

Nevertheless, links between various microbial infections as causative or precipitating triggers of idiopathic PD have recently received renewed attention, especially in the wake of recent epidemiological studies. In a systematic and population-based case control study from Denmark, various types of infections were examined for their potential link with PD. An association was shown between the onset of PD and infection with influenza when the infection occurred more than 10 years before clinical diagnosis.42 Similar findings were observed in a second and independent systematic population-based study. Here, cases from Finland and the UK were studied.43 A significant association was again found between infection with influenza and the subsequent development of PD if the time between infection and diagnosis was 5 years.43

Another notable association from these studies is the link between PD and urinary tract infections.⁴² Aside from influenza infections, urinary tract infections were found to be the only other type of infection associated with an increased risk of PD more than 10 years after infection.⁴² Interestingly, similar findings were found for urinary tract infections and MSA.⁴⁴ Urinary tract infections are common in PD but also in MSA, especially in the years before diagnosis. Even though MSA has a shorter prodrome, urinary tract infections are associated with increased risk of MSA when the infection happens 8 years before the appearance of cardinal symptoms.⁴⁴ In the general population, urinary tract infections are prevalent, especially in women. However, in PD and in MSA, urinary tract infections affect men and women equally.^{44,45} The reasons for this are not fully understood, and it remains unclear how central or peripheral pathology of synucleinopathies might contribute to increased susceptibility to urinary tract infection, which in turn could exacerbate neurodegenerative processes, as further discussed below. This requires more research, as these infections are linked with the onset of synucleinopathies.

Other epidemiological results are less clear as to when they occur in relation to the prodrome and how they might contribute to synucleinopathies. Some epidemiological studies focused on the presence of pathogen-specific antigens present in the plasma or brain tissue of people with PD by using ELISA, PCR or immunohistochemical methods. Therefore, it is not known if these infections would have occurred before or after the onset of disease. Other studies also relied on self-reporting or used a questionnaire-based methodology to examine potential associations with PD,⁴⁶ which can potentially introduce a recall bias and an under- or overestimation of infection.

Nevertheless, from these studies it is apparent that multiple pathogens might increase the risk of developing synucleinopathy or unmask underlying pathogenesis during disease progression. As such, notable links have been established between infections with hepatitis C virus, herpes virus, *Helicobacter pylori* and other types of pathogens that are further summarized in Table 1.

Prevention and vaccination

Studies have reported that vaccination may decrease the association between influenza or herpes zoster infection and the development of PD after vaccination.^{46,60} However, in one prospective study in people older than 60 years there was no association found between influenza vaccination and risk of PD.⁶¹ Antivirals for the treatment of chronic hepatitis also result in a reduction of associated PD risk. 47,58,59 Vaccinations against these viruses could potentially reduce the risk of infection and, subsequently, lower the risk of developing PD. Individual factors such as age, overall health, and genetic risk factors could be taken into consideration when making decisions regarding vaccination and to determine the most appropriate vaccination strategy for individuals at risk of developing PD.

Vaccines are available for most of the viruses listed in Table 1; however, vaccination rates remain

Table 1

Infectious pathogens and their association with PD or MSA. This table summarizes the associations between various infectious pathogens and the risk of developing Parkinson's disease (PD) or multiple system atrophy (MSA) based on published epidemiological studies and reports. The table is organized by pathogen type, including viruses, bacteria, and fungi, and also highlights the impact of vaccination and antiviral treatments on PD risk. For some viruses, such as SARS-CoV-2, case reports and studies have suggested an increased risk of PD following infection, but no clear epidemiological association has been shown. Pathogens such as influenza A, hepatitis B and C viruses are associated with a higher likelihood of developing PD when the infection happens before disease onset. Some infections like the measles virus might reduce the risk of PD. Bacterial infections, such as those caused by Helicobacter pylori and Mycobacterium tuberculosis, are linked to an increased risk of PD, while fungal infections like Malassezia are also implicated. Vaccination, particularly for influenza, varicella zoster virus and antiviral treatment against hepatitis C, appear to reduce the risk of PD. Some research has also found no clear association between vaccination against influenza and PD risk. These findings underline the complex relationship between infections and PD, suggesting that both the presence of specific pathogens and the body's immune response to them could influence the development and progression of PD

Infectious pathogen	Association	Reference
Viruses		
SARS-CoV-2	Case Reports	22
Hepatitis B virus	Increased risk PD	47
Hepatitis C virus	Increased risk PD	47
Herpes virus	Reduced/increased risk PD	48-51
Influenza virus	Increased risk PD	42,43,46
Measles virus	Reduced risk PD	46,52
Bacteria		
Helicobacter pylori	Increased risk PD	47,53
Bordetella pertussis	Increased risk PD	54
Corynebacterium diphtheriae	Increased risk PD	55
Mycobacterium tuberculosis	Increased risk PD	56
Uropathogenic E. coli	Increased risk PD and MSA	42,44
Fungi		
Mallasezia	Increased risk PD	47,57
Vaccination or antiviral treatment		
Influenza vaccination	No association/Reduced risk of PD	42,43
Hepatitis C antivirals	Reduced risk of PD	47,58,59
Varicella-Zoster virus vaccination	Reduced risk of PD	60

relatively low, even for people that already have PD. It has been estimated that vaccination coverage for influenza is approximately only half of all PD patients⁶² whereas for SARS-CoV-2 vaccination rates in people with PD are even lower than that of the general population.⁶³ A reason for this could be restricted access for people with PD to primary healthcare⁶⁴ but efforts are needed to improve the number of people with PD or related disorders with healthcare access.

Before as well as after diagnosis of synucleinopathies, it could thus be important for people at risk to get vaccinated for several reasons. PD can weaken the immune system, making individuals more susceptible to infections.⁶⁵ Infections and illnesses can sometimes exacerbate disease symptoms or trigger temporary worsening. By avoiding illness through vaccination, individuals with PD may manage their symptoms more effectively. Particularly in MSA, or in patients with autonomic dysfunction, patients may be more vulnerable to developing complications from infections such as pneumonia or influenza. Vaccinations against these diseases can reduce the risk of serious complications and hospitalization.

It could thus be hypothesized that infectious pathogens might be associated with an elevated risk of developing a synucleinopathy. However, as we will also discuss in the next section, these associations do not necessarily imply causality, as these observations, although from multiple systematic and observational studies, cannot definitively establish a causal relationship. On the other hand, the association of these risk factors warrant their involvement in synucleinopathies, regardless of the directionality of these associations. Infections could precipitate a preclinical condition and thus accelerate or trigger the onset of synucleinopathy which needs to be further investigated.

A vicious cycle of infections and synucleinopathies?

These historical and epidemiological observations are a good example to illustrate the complexity of pathogen-host interactions: how do we study the relationship between a potential causative agent and a disease that is only diagnosed following a long asymptomatic period? The prodromal phase in synucleinopathies can range from approximately 5-10years in MSA to 15-20 years in PD.⁶⁶⁻⁶⁹ If infections would act as triggers of disease, they would therefore need to occur around the time when the asymptomatic period would start. An interaction would at best support the contention that an association might exist, but it cannot prove causality.

The protracted prodrome in synucleinopathies now brings us to a second challenge: if an infection happens after the start of the prodrome, how can we be certain about the direction of causation? If an infection happens too close to the time of diagnosis, this would raise concerns about the causative role of the infection since some of the prodromal symptoms can be risk factors for infection. For these cases, the infection might then be a consequence of the disease rather than a trigger of the disease. However, this does not preclude infections from accelerating disease progression or aggravate symptoms.

One of these early cardinal symptoms of PD and MSA, which in some cases manifests during the prodrome, is autonomic dysfunction.^{68,70–72} The autonomic nervous system controls several crucial functions such as heart rate, blood pressure, swallowing, digestion, sweating, temperature, or urinary voiding—among others. In PD and especially in MSA, autonomic dysfunction can be severe. Autonomic dysfunction can occur with the loss of peripheral innervation,^{73,74} leading to desensitization.

Dysfunction of the autonomic nervous system can lead to a higher susceptibility to infections via several mechanisms. Swallowing difficulties or dysphagia can lead to aspiration pneumonia, whereas constipation can disrupt the composition of microbiota or lead to fecal retention and an increased risk of urinary tract infections (via translocation of bacteria). Similarly, loss of bladder control, can lead retention of volume in the urinary bladder, which again can increase the risk of urinary tract infections.

Another important function, that is impaired early in MSA, is thermoregulation.^{75,76} Upon infection, an increase in body temperature or sweating would normally indicate signs of infection, but in MSA these functions are impaired. Impaired sweating can create an environment conducive to the growth of bacteria and increase the risk of skin infections in PD and MSA.^{57,77} In the absence of these signs, this can create a dangerous situation where signs of an infection will be muted. Therefore, in some cases, it is possible that the normal signs of infection are missed in which a condition can worsen, causing neurological symptoms, such as confusion or delirium, which require urgent medical treatment.

There is also a more direct way via which the autonomic nervous system is important in the host response against infections. Neurotransmitters of the autonomous nervous system, such as acetylcholine or norepinephrine, can bind to immune cell receptors, which will lead to activation or proliferation of these cells.^{78,79} Alternatively, via the release of neurotransmitters immune cells that are stored in lymphoid organs can be recruited.⁸⁰ The loss of autonomic innervation can result in changes in neuroimmune interactions and potentially impair the ability of the immune response to fight off infections.⁸¹

Clearly, due to the protracted nature of synucleinopathies, and their long prodromal phase, there are significant challenges in studying how infections might influence the initiation or the progression of these diseases. Nevertheless, there is growing epidemiological evidence that infections play a role in both disease initiation and disease progression and that infections might have a causative and reciprocal role in the pathogenesis of these diseases.

INFECTIONS ARE POTENTIAL TRIGGERS OF α-SYNUCLEIN TRANSMISSION

Pathogens from the outside environment continuously threaten our body's defense systems through various routes. Inhalation exposes us to respiratory pathogens, while ingestion introduces harmful agents into our digestive system. Skin contact and compromised barriers allow entry for pathogens, and mucosal surfaces are susceptible to invasion through direct contact. Some pathogens can breach these barriers and enter the peripheral nervous system through direct infiltration or through blood circulation. Some viruses can invade peripheral nerves, or directly cross the blood-brain barrier (BBB) and enter the brain, causing neuropathies, encephalitis, or postencephalitic parkinsonism.^{16,30,33,82}

The Braak hypothesis postulates that one or two insults occur in the nose or the gut and that this can trigger a cascade of events leading to central pathology in idiopathic PD.^{83,84} In one of their original publications, Hawkes, Braak and Del Tredici mentioned that "*a neurotropic pathogen, probably*

viral, enters the brain via two routes: (i) nasal, with anterograde progression into the temporal lobe; and (ii) gastric, secondary to swallowing of nasal secretions in saliva".⁸⁴ More recently, Borghammer and colleagues further built on this hypothesis and proposed a framework in which the starting point and the anatomical distribution of α Syn pathology could lead to a categorization of PD into distinct subtypes; a brain-first and a body-first subtype, in which pathology specifically related to α Syn progresses via different but predictable routes throughout the body.^{68,85}

Although these ideas are focused on PD, there is experimental evidence that the underlying mechanisms related to α Syn transmission are shared with MSA and DLB.⁸⁶ Under pathological conditions, α Syn aggregates into small β -sheet-rich assemblies and further aggregate into large filamentous fibrils.⁸⁷ When fragmented, fibrillar fragments can transmit and be taken up by adjacent cells.⁸⁸ These aggregates can induce the misfolding and aggregation of soluble synuclein proteins in neighboring cells, leading to the progressive spread of α Syn pathology throughout the nervous system.⁸⁹

Thus, α Syn aggregates taken up by neighboring cells will serve as templates for the misfolding of soluble α Syn protein and facilitate the transmission of pathology between cells.⁸⁹ This has led to the suggestion of a prion-like propagation of α Syn pathology, characterized by adopting abnormal conformations that then propagate pathology through an infectiouslike mechanism.^{90,91} While synucleinopathies share similarities with classical prion diseases, it is important to note that the transmission of α Syn between individuals has not been established. The transmissible nature here thus refers to the transmission of α Syn between cells and not individuals.

Several studies have shown that microbial infections can directly trigger the accumulation of α Syn and its conversion from a soluble form to an aggregated state.^{44,92–94} This conversion appears to be highly dynamic and occurs within the first hours of infection.⁴⁴ The presence of pathogens or their byproducts can alter cellular metabolism and signaling pathways, leading to dysregulation of protein degradation mechanisms such as the ubiquitinproteasome system and autophagy.^{95,96} Impaired protein clearance can expedite the accumulation of misfolded synuclein proteins and facilitate their aggregation.⁹⁷ At any infected site, α Syn is thus prone to accumulate and misfold. It can build up within visceral barrier sites that are exposed to inflammatory pathogens. In this highly reactive and rapidly changing environment post-translational modifications such as phosphorylation, oxidation, acetylation, ubiquitination, glycation, glycosylation, nitration, and truncation can further modify the structural, biochemical, and cellular properties of α Syn.⁹⁸

The presence of aggregated α Syn in visceral sites is a observed in all individuals. In normal as well as affected individuals.44,99 From these visceral sites, aggregates are potentially prone to travel to the brain via axonal transport.¹⁰⁰ Injection of recombinant α Syn fibrils in peripheral sites leads to progressive symptoms that resemble PD and MSA in animal models.^{44,101–104} Successful and predictable experimental transmission of α Syn has now been shown for three common routes to the brain; the nose, the gastrointestinal system, and the urogenital system. ^{44,101,103–108} Repeated infections or inflammatory exposure within the gut or the urogenital system could thereby predispose to gut-to-brain pathology (bodyfirst type) whereas repeated infections in the nose might predispose to brain-to-gut pathology (brainfirst type).^{109,110} Infections in specific peripheral sites could thus contribute to heterogeneity in synucleinopathies as each site can provide a trigger with unique transmission routes and associated pathology.

Other transmission routes, such as systemic transmission have also been studied.¹¹¹⁻¹¹⁴ However, the pattern of spread here is less predictable compared to local injections. Nevertheless, given the native role of α Syn in hematopoietic or immune functions, the systemic route might have an important homeostatic role that influences or mitigates the overall burden and the turnover of α Syn protein. Intravenous delivery of a Syn fibrils for instance has been shown to result in pathology that mimics early prodromal stages of PD.¹¹⁴ It is also not inconceivable that peripheral organs, such as the liver or the kidneys might play a homeostatic role in the clearance of pathological aSyn.^{115,116} Along those lines, systemic transmission could be a relevant and possibly parallel peripheral route via which synucleinopathy could be influenced (Fig. 2).

Studying how virulent agents might interact with α Syn in peripheral barrier sites could help us understand how the initial steps of aggregate formation take place. A better understanding of this process might also provide new clues as to how distinct assemblies of are created. Since peripheral aggregates are common in people with and without clinical symptoms, it suggests that some of these aggregates might have only a little seeding capacity. Injection

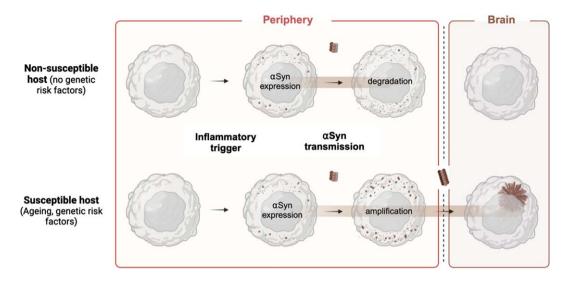


Fig. 2. **Body-to-brain transmission of \alphaSyn after infection.** Expression of α Syn can be trigger by inflammatory pathogens. Different assemblies of α Syn will be transiently expressed in response to inflammation. Inflammation can cause the misfolding and aggregation of the endogenous protein. Newly formed assemblies are likely to be degraded by the host protein degradation machinery. In non-susceptible individuals, there will be no body-to-brain transmission of α Syn. In susceptible individuals, ageing or gene variants impair proteosomal function or general protein homeostasis. Assemblies that escape protein degradation can structurally reorganize and form potent seeds for transmission. Aggregated assemblies of α Syn can further accumulate as they are resistant against degradation. Stable seeds can serially transmit to neighboring cells and to the brain via sympathetic and parasympathetic connections.

of aggregates isolated from peripheral sympathetic ganglia has indeed shown a lack of seeding capacity compared to aggregates isolated from the brain.¹¹⁷ Varying efficiencies of transmission could depend on the conformation, the stability, or the maturity of α Syn assemblies. Whether a specific virulent agent may induce the formation of α Syn assemblies with higher seeding capacities may also depend on age at exposure, genetic risk factors and further environmental impacts.

In that regard, it has been shown that distinct types of strains exist in PD, DLB and MSA.⁸⁶ Strains of α Syn are fibrils with different molecular structures that each have a specific biological or pathological role because of their unique conformation. While experimental models have provided tentative answers to where and how synucleinopathy might start, they do not yet explain how distinct strains of α Syn might arise. a Syn is known to form different fibril structures with distinct toxicities and seeding capacities.111 These strains of α Syn are notably distinct in Lewy Body disorders and MSA have as they have a different molecular structure^{118,119}. In experimental animal models, fibril strains derived from PD, DLB and MSA patients have unique pathological features, and it shows that a structural-pathological relationship exist between a given strain and the resulting phenotype.118-120

We and others recently postulated that during serial transmission of a Syn pathology the conformation of α Syn can change and that the host cellular environment is responsible for how α Syn assembles and forms strain-specific assemblies.^{88,121} Multiple studies have shown that the formation of strain-specific assemblies from heterogenous smaller assemblies requires multiple steps of structural reorganization, with the rearrangement of intramolecular interactions.^{86,122–124} This requires transient interactions with cell-specific chaperones or molecules, or conversely the failure of proteostatic systems, that facilitate the conversion and the maturation of unstable aggregates.^{125,126} This process continues until α Syn aggregates are thermodynamically stable and become the dominant structural conformer.¹²¹ As the transmission of α Syn assemblies from a peripheral trigger site, either from the nose, the gut or elsewhere, is determined by genetic and cellular risk factors,^{109,127-130} these risk factors will aid the formation of stable seeds within the cellular environment along the peripheral-central axis in which serial transmission takes place.

Even given the unique genetic makeup and the unique exposome of each individual, the progression of synucleinopathy could thus happen in a way that is unique but still predictable. The conformational landscape of α Syn will gradually and

progressively narrow while the disease progresses. The continued and repeated transmission of α Syn will eventually lead to a restricted number of disease-specific strains of α Syn within a larger group of α Syn assemblies.^{88,121} Infectious agents that rapidly trigger aggregation could thus be an important upstream event in the etiology of α Syn strains.

Infections are thus emerging as upstream triggers in α Syn-related idiopathic PD and other synucleinopathies. Insults in the periphery can initiate local pathology and potentially lead to distinct disease subtypes based on synucleinopathy progression routes. Infections rapidly catalyze the accumulation and conversion of α Syn into newly formed aggregates, of which most will be unstable and degraded. Instead, in some cases, these aggregates will persist so that they can transmit to other cells and cause progressive pathology. These peripheral aggregates may propagate to the brain via retrograde axonal transport, as they further mature into disease-relevant assemblies, highlighting the potential and potentially pivotal role of the exposome in the development of synucleinopathies.

MECHANISMS OF HOW INFECTIONS CAN AFFECT SYNUCLEINOPATHIES

Aside from the potential upstream role of infections in the etiology of synucleinopathies, there are several other mechanisms through which infections can influence disease development. This could happen via: 1) direct infection of the brain (in rare cases); 2) an immune response in peripheral organs, including the gut and the bladder, which transmits to the brain via the systemically or via the olfactory epithelium; 3) an alteration of peripheral microbiomes such as in the gut that transmits to the brain; 4) triggering of peripheral α Syn accumulation or aggregations that transmits via the peripheral nervous system; 5) invasion of microbial byproducts into the brain after peripheral infections. In all these scenarios the host immune response to the pathogen could play a central role in the pathology development.

Inflammation is part of the normal immune response to harmful stimuli, which can include pathogens or damaged cells, and is an important contributor to neurodegenerative diseases including synucleinopathies.^{131–133} Host cells use pattern recognition receptors (PRRs) and other receptors to monitor and identify a wide range of pathogen-associated molecular patterns (PAMPs) derived from

different microorganisms including viruses, bacteria, and fungi.¹³⁴ They also recognize damageor danger-associated molecular patterns (DAMPs) released from senescent and dying cells.¹³⁵ The activation of the innate immune system mediated by these receptors is emerging as an important pathological mechanism underlying neurodegenerative diseases.¹³⁶ There are several families of extracellular and cytosolic PRRs, that include the Toll-like receptors (TLRs), the NOD-like receptors (NLRs) and other cytosolic sensors. PRR activation is the first step of the immune response that triggers specific cellular signaling pathways inducing the production and release of pro-inflammatory mediators (cytokines and chemokines) and the removal of damaged cells (induction of autophagy or cell death).

Inflammatory insults can reach the brain and trigger a highly specialized immune response characteristic of the central nervous system (CNS) or neuroinflammation.^{137–140} This specialized immune response relies on unique immune cells and the lymphatic system that controls an intricate balance between the CNS and peripheral immunity.¹⁴¹ At the interface of the brain and the periphery microglia and astrocytes work with brain border immune cells in response to inflammatory stimuli. Using different strategies pathogens can trigger a neuroimmune response with long lasting effects on glial activation and cytokines production, with the potential to aggravate an ongoing pathological process.¹⁴⁰ The inflammatory process that occurs during an infection and its proper resolution further depends on the proteostatic cellular machinery. Chronic inflammation or recurrent exposure to inflammatory agents generates stress on these cellular pathways leading to toxic protein accumulation and eventually cell death. This pathological process can be aggravated by aging and the presence of genetic risk factors.

Synucleinopathies are characterized by specific signatures of central and peripheral inflammation. PD is characterized by chronic microglial activation and widespread neuroinflammation.^{142–145} There is more prominent microglial activation and T cell infiltration in typically affected regions including the substantia nigra.^{142,146} Similarly, MSA is characterized by widespread microglial activation in multiple affected areas.¹³² In the case of DLB, microglial activation is early and transient, while in later stages patients lack reactive microglia. Infiltration of natural killer cells and T cells is also observed in these affected areas^{131,147–149}.Next to central inflammation, signs of peripheral inflam-

mation are also typical of synucleinopathies.^{150,151} Some of these changes include increased proinflammatory cytokines, changes in immune cell numbers and circulating T cells that are reactivated to diseasespecific α Syn peptides.¹⁵⁰ In the case of PD, several studies have shown a decrease of circulating lymphocytes and an increase of neutrophils years before diagnosis.¹⁵² These inflammatory changes suggest an early involvement of the immune response in synucleinopathies pathogenesis.

Converging cellular signaling pathways during infection and synucleinopathy

Several cellular signaling pathways have been shown to be affected in synucleinopathies, including impaired proteostasis, vesicular transport and mitochondrial dysfunction.¹⁵³ Many of these affected cellular pathways are altered during neuroinflammatory states, where glial cells and neurons respond to a peripheral or central insult. Interestingly, such cellular pathways are also impaired by pathogens during infection. Some pathogens have the capacity to hijack the protein synthesis and autophagy machineries so that they can replicate and avoid the host immune response.^{82,154} Mitochondria are also central to the immune response as these organelles participate in several inflammatory signaling pathways. This includes an innate immune mechanism that detects the presence of cytoplasmic DNA within cells. This pathway plays a crucial role in the body's defense against infections, particularly viral infections, but it can also sense mitochondrial DNA. This pathway involves cyclic GMP-AMP synthase (cGAS) and the transmembrane protein stimulator of interferon genes (STING).¹⁵⁵ The cellular responses to inflammation can trigger deleterious events, including oxidative stress and cytokine-receptor-mediated apoptosis, which might eventually lead to neuronal loss.

The ability to identify foreign DNA is essential for fending off bacterial and viral infections. Cytosolic DNA is a crucial DAMP, capable of activating the innate immune system through PRRs. Cyclic GMP-AMP synthase (cGAS), is a PRR that can detect foreign DNA from pathogens and self-DNA released during cellular damage, activating stimulator of interferon genes (STING), which will trigger a type I interferon (IFN-I) immune response. In recent years, the cGAS-STING pathway has emerged as a critical player in neurological disorders characterized by chronic neuroinflammation. Viral encephalitis after HSV-1 infection activates the cGAS-STING pathway leading to microglial-mediated IFN-I response.¹⁵⁶ STING-deficient mice are more susceptible to HSV-1 encephalitis after peripheral infection.¹⁵⁶ Moreover, the activation of IFN-I signaling has been observed in postmortem PD human samples and in several mouse models.^{157,158} Impaired mitophagy in PD animal models triggers the activation of the cGAS-STING pathway with functional effects on neuroinflammation, dopaminergic neuron loss and motor function.^{158,159} Inhibition of this signaling pathway alleviates the functional effects and even pathological α Syn accumulation in an α Syn preformed fibrils mouse model.¹⁶⁰

Another innate immune response signaling pathway that is activated upon infection is the inflammasome. Inflammasomes are a family of protein complexes that can sense environmental and cellular danger signals.¹⁶¹ One of the most studied inflammasome proteins is the NLR family pyrin domain containing 3 (NLRP3)-inflammasome, the apoptosis-associated speck-like protein containing a caspase activating recruitment domain (ASC) and caspase-1.¹⁶¹ Upon activation, inflammasomes produce and release pro-inflammatory cytokines IL-1B and IL-18 through pyroptotic pores.¹⁶¹ Markers of activation of this pathway are elevated in the brain and in blood samples from PD.162-164 Activation of the NLRP3-inflammsome pathway is also present in genetic and toxicological mouse models of PD.^{162,165–167} Recent studies suggest that downregulation of NLRP3-inflammasome can ameliorate dopaminergic neuron loss and motor deficits in different mouse models of PD.¹⁶⁸

Affected signaling pathways in synucleinopathies resemble those seen as a response to pathogen invasion or inflammatory processes. These changes often involve cellular stress and a combination of upregulation of defense mechanisms and downregulation of pathways essential for cell homeostasis. These similarities suggest that both synucleinopathies and infections may involve common mechanisms of neuronal damage and dysfunction (Fig. 3).

Infections as triggers of synucleinopathy

Viral triggers

As mentioned earlier, there are well-documented cases of viral parkinsonism caused by influenza, coxsackie virus, Japanese encephalitis B, St. Louis, West-Nile and HIV. More recently, there are case reports that infection with SARS-CoV-2 causes L-

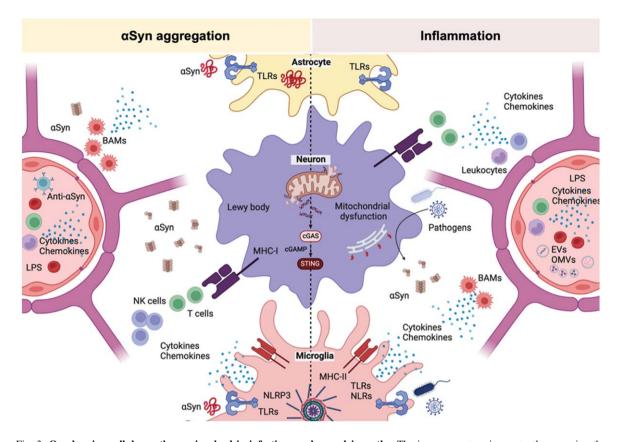


Fig. 3. **Overlapping cellular pathways involved in infections and synucleinopathy.** The immune system is constantly surveying the presence of pathogens and other inflammatory agents. In the brain, immune and non-immune cells use extracellular and intracellular pattern recognition receptors (PRRs) to identify pathogen-associated (PAMPs) and damage-associated molecular patterns (DAMPs) from microbes and dying cells, respectively. In astrocytes, toll-like receptors (TLRs) that recognize microbial-associated molecular patterns are activated by α Syn. TLRs also mediate the internalization of α Syn into astrocytes. In both cases, TLRs activation triggers a pro-inflammatory phenotype in these cells. In neurons, both infections and neurodegeneration are characterized by endoplasmic reticulum stress, autophagy impairment and mitochondrial dysfunction. Mitochondrial dysfunction can contribute to the activation of an interferon immune response via cGAS-STING. During neurodegeneration and infections, α Syn and other pro-inflammatory mediators are released. Some pathogens and pro-inflammatory mediators can damage the neurovascular unit and cause blood brain barrier (BBB) permeability. At the brain border, α Syn can interact with border associated macrophages to facilitate T cell infiltration. These inflammatory conditions are also characterized by the infiltration of peripheral immune cells including T cells and natural killer (NK) cells into the brain parenchyma. Nigral dopaminergic neurons express MHC-1 receptors that can present antigens to cytotoxic T cells. Brain macrophages, but in particular microglia, can recognize PAMPs and DAMPs including α Syn using different PRRs like TLRs and NOD-like receptors (NLRs). Upon recognition by PRRs, microglia establish a pro-inflammatory phenotype with cytokine production that can have persistent effects on neuroinflammation.

DOPA responsive parkinsonism.^{41,169} The reason why these viruses cause parkinsonism is not well understood and whether viral invasion of the CNS is necessary for the development of neurological symptoms is still under investigation.

However, many of these viruses have been shown to preferentially target the midbrain and the dopaminergic system.^{41,170,171} These viruses have the potential to access the CNS via different mechanisms.¹⁷² Some of these pathogens can access the CNS from the olfactory epithelium or alternatively, via innervation from visceral organs. Others access the brain by infecting immune cells that transport the virus over the BBB into the CNS. Another mechanism to enter the brain is by extravasation from brain capillaries. Viral infection can decrease the expression of tight junction proteins and increase vascular permeability to facilitate this process.¹⁷³ After entering the CNS, infectious pathogens can trigger a neuroimmune response, which includes glial activation, cytokine and chemokines release, oxidative stress, peripheral immune cells infiltration and potentially α Syn aggregation.¹⁶⁸

Recent evidence has shown that several viruses can cause α Syn pathology. Brain infections with influenza virus, West-Nile virus, picornavirus, cox-

sackievirus or SARS-CoV-2 have been shown to trigger α Syn pathology.^{157,174–178} Infection with H1N1 via the olfactory route inhibits proteostasis in the substantia nigra of immunocompromised *Rag* KO mice and leads to α Syn aggregation *in vivo*.¹⁷⁷ Coxsackievirus can directly cause aggregation of α Syn in cell and animal models and again cause Lewy-like pathology *in vivo*.¹⁷⁴

Similarly, in experimental models of SARS-CoV-2 infection, intracellular α Syn aggregates in the substantia nigra were found in SARS-CoV-2 infected macaques.¹⁷⁶ SARS-CoV-2 is generally not a neurotropic virus, but we and others demonstrated that intranasal infection in hamsters can cause persistent inflammation in the olfactory and cortical regions,^{179,180} followed by tau and α Syn pathology,¹⁸⁰ in the absence of overt CNS invasion.

Microglial activation and cytokine production in the brain can persist long after SARS-CoV-2 infection has resolved, suggesting that CNS damage after infection may be due to a cytokine storm and thus the neuroimmune response rather than direct infection with SARS-CoV-2.^{181,182} Further studies on the time course in intranasally infected hamsters revealed that microglia cell density and a Syn immunoreactivity decreased at 6 days post SARS-CoV-2 infection, then rebounded to overt accumulation at 21 days post infection. This biphasic response was most pronounced in the amygdala and striatum, brain regions affected in PD, and in female hamsters. Importantly, female sex appears to predispose to long term complications after SARS-CoV-2 termed post-COVID19.178

Thus, SARS-CoV-2 profoundly disrupts brain homeostasis without neuroinvasion, via neuroinflammatory and protein regulation mechanisms that persist beyond viral clearance, providing a potential link between viral infection and the risk of neurodegenerative diseases.

In vitro experiments have shown that SARS-CoV-2 peptides derived from the SARS-CoV-2 spike protein have amyloidogenic properties that seed α Syn fibrillization.^{183–186} Furthermore, the spike protein primed and activated the NLRP3 inflammasome in human microglia, and enhanced α Syn-mediated NLRP3 activation in microglia cells.¹⁸⁷ There is evidence that these viral proteins still cross the BBB and reach the brain parynchema.^{188,189} Of note, the neurovascular unit is altered upon SARS-CoV-2 infection. After infection there is brain damage to cerebral small vessels evidenced by increased string vessels and microvascular pathology,¹⁹⁰ as well as pericyte-mediated capillary constriction.¹⁹⁰ Under viral infection-induced BBB alterations and peripheral inflammation, viral proteins may cross into brain parenchyma causing neuroinflammation and concomitant neuronal damage.

However, such mechanisms are thus not restricted to the CNS or the nose-brain axis. Indeed, as mentioned earlier, α Syn aggregation is also observed in peripheral tissues, including the gut, the skin, and the urinary bladder. Infections can trigger α Syn aggregation and its release from neutrophils.⁴⁴ Molecular dynamics simulations suggest that a SARS-COV-2 spike protein fragment, cleaved by neutrophil elastase under inflammatory conditions, favors α Syn fibrillar seeding conformations, suggesting again that peripheral immune cells could potentially contribute to α Syn aggregation during infections.^{184,191}

Gut triggers

The involvement of the gut in synucleinopathies has been a field of intensive research. Gut symptoms such as bloating and constipation appear early in some people with PD, indicating that the gut might be a starting site of pathology for these cases. This is in conjunction with early observations of α Syn pathology of the enteric nervous system as well as autonomous nervous system.^{68,110,192} There is a bidirectional connection between the gastrointestinal tract and the brain, called the gut-brain axis, and it can be influenced by gut infections, dysbiosis, and inflammatory responses.

Changes in the gut microbiota happen early in PD. Multiple studies have shown that intestinal microbiota dysbiosis can be a prominent feature.¹⁹²⁻¹⁹⁵ Several factors can influence the gut microbiome, such as geographic factors and ethnicity. However, despite these factors, PD related dysbiosis is broadly characterized by a lower relative abundance of anti-inflammatory bacterial genera and an increase of pro-inflammatory and opportunistic pathogens.^{195,196} These can include *Escherichia*, Klebsiella, and Porphyromonas. 195, 196 Even though the causes for dysbiosis in PD are not fully understood, the gut microbiota can be affected by exposure to pollutants, pesticides, and infections.¹⁹⁷ Research in preclinical PD models has shown that gut microbiota manipulation can influence symptoms and pathology including α Syn aggregation.^{198–200} More recently, it was shown that a fecal microbiota transplantation can potentially alleviate motor symptoms in early-stage PD patients.²⁰¹

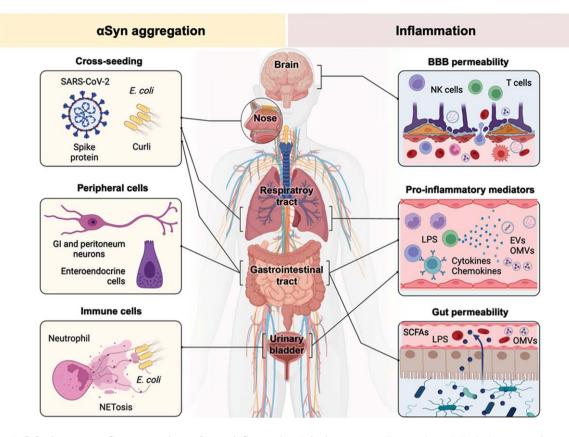


Fig. 4. Infections cause α Syn aggregation and neuroinflammation. Infections can contribute to the pathological prossess of synucleinopathies via several routes. In the left panel, the cellular mechanisms that directly increase α Syn expression or influence its aggregation are described. Different bacterial and viral proteins can seed the aggregation of α Syn via direct or indirect molecular interactions. Various cells of the peripheral nervous system can express α Syn in response to bacterial and viral infections. Different cell types in the gut (peritoneal neurons and enteroendocrine cells) respond to inflammation by expressing α Syn. These cells further connect to the parasympathetic vagal nerve via which α Syn can transmit. α Syn can be released by the immune system itself, via neutrophils, during the response to bacterial infections. In the right panel, it is described how inflammatory triggers contribute to synucleinopathy. Pathogens and pro-inflammatory mediators can induce dysfunction of the neurovascular unit and cause blood-brain barrier (BBB) permeability. This increased permeability can facilitate the infiltration of pathogens and peripheral immune cells. Pro-inflammatory mediators or other microbial-derived molecules can be released as a reaction to infections in various peripheral organs. Gut dysbiosis and/or infections increase epithelial permeability and as a consequence pro-inflammatory mediators are released into the circulation; including cis-activating microbial byproducts such as SCFAs and LPS; in addition to trans-activating elements including bacterial outer membrane vesicles (OMVs).

Gut dysbiosis and infections can cause pathological changes in the immunoregulatory function of the gut and increase intestinal permeability. Increased intestinal permeability is a characteristic of PD.²⁰² It is accompanied by changes in gut microbial metabolites, such as bacterial lipopolysaccharides (LPS) and short-chain fatty acids (SCFAs). Inflammatory mediators can be released from commensal and pathogenic gut microbes and trigger unwanted systemic and central effects. It has been proposed that the disruption of the gut epithelium could trigger a feedback loop that alters the microbiome to a more pro-inflammatory profile resulting in an increased permeability, oxidative stress, and aggregation of α Syn in the gut.²⁰³ Experimental and clinical studies have implicated the enteroendocrine cells of the gut epithelium.^{204,205} Enteroendocrine cells can sense the luminal content in the gastrointestinal epithelium, and they play crucial roles in regulating gastrointestinal functions, including digestion, nutrient absorption, and gut motility by producing and releasing hormones. LPS and SCFAs are potent activators of enteroendocrine cells and interestingly, enteroendocrine cells also produce α Syn. In response to inflammatory stimuli, these cells can transiently express and release α Syn.^{204,206} This thus directly links infectious agents and microbial metabolites (LPS or SCFAs) with the production of α Syn in the gut (Fig. 4). Microbiota-derived SCFAs are common bacterial fermentation products. These gut microbial metabolites, such as for instance butyrate, propionate, or acetate, have many physiological functions including reducing inflammation and preserving epithelial integrity.²⁰⁷ SCFAs-producing bacteria are less abundant, and these metabolites are decreased in the feces of PD patients.^{208–210} On the other hand, the release of SCFAs from the gut can influence microglial homeostasis.²¹¹ Animal studies have demonstrated that SCFAs have functional consequences by increasing α Syn aggregation and the development of motor dysfunction.²¹²

LPS is a component of the outer membrane of Gram-negative bacteria and a potent activator of the innate immune response. LPS from the Enterobacteriaceae family is one of the most immunogenic forms and is found abundantly in the gut of PD patients.^{196,213} It has been recently shown that PD patients have elevated levels of LPS in the blood and high levels of LPS binding protein (LBP) during the prodromal phase are associated with an increased risk of PD.^{214–216} LPS activates the innate Toll-like receptors 2 and 4 and triggers an immune response that promotes the production of pro-inflammatory cytokines, chemokines, and reactive oxygen species. TLRs activation in microglia leads to the production of pro-inflammatory cytokines, the initiation of the adaptive immune response and recruitment of T and B cells. In PD models, LPS induces a neuroinflammatory response that ultimately leads to neurodegeneration.²¹⁷⁻²²⁰ This is likely caused by the selective vulnerability of dopaminergic neurons of the substantia nigra to LPS insults.²¹⁷⁻²²⁰ Additionally, LPS administration can trigger α Syn expression and aggregation.²²¹⁻²²⁴ This has been observed in both the gut as well as in the brain, and more specifically in the substantia nigra, of animal models. 221-224

Aside from the direct or cis-acting pathogenic effects of bacteria on α Syn and PD pathogenesis, bacteria can also exert trans-acting effects. This happens via the release of outer membrane vesicles (OMVs) or alternatively, via the release of amyloidogenic proteins. Gram-negative bacteria can release OMVs from the stomach or the gut which can cross the BBB.²²⁵ *E coli* or *Helicobacter pylori* OMVs are only a size of 20–200 nm and thus can readily cross the BBB to infect neurons and glial cells.²²⁶ When taken up by astrocytes, this will lead to the expression of complement proteins and cause an inflammatory reaction.²²⁷ OMVs loaded with LPS can also

interact with microglia and other PRRs-expressing cells.²²⁵ Some of these bacterial vesicles also transfer different bacterial molecules such as RNAs, endotoxins and toxins.²²⁵ Because of the direct transfer of pathogen metabolites to the cytosol, OMVs can trigger the activation of the intracellular NLRP3 inflammasome pathway resulting in the release of pro-inflammatory cytokines and mitochondrial dysfunction.²²⁸

Next to these potentially pathogenic carrier vesicles, bacteria can release amyloidogenic proteins. One such well-characterized bacterial protein is the amyloid protein curli. Curli is produced by *E. coli* and the protein can aggregate into fibrils to facilitate biofilm formation and help with cellular adhesion. Curli fibrils play dual roles in promoting bacterial colonization and infection while also triggering the host immune response, contributing to both bacterial persistence and host defense mechanisms. Curli can cross-seed the aggregation of α Syn *in vitro* and injection of curli fibrils can lead to α Syn pathology in the gut.²²⁹ Pathological transmission of α Syn to the CNS has been observed in mice after transplantation of curli-expressing *E. coli.*²³⁰

Aside from gut bacteria, viral gut pathogens have also been associated with synucleinopathy.²³¹ Viral infection with norovirus can induce a persistent increase in α Syn expression in the enteric nervous system.⁹² Here, the degree and the duration of viral infection in infected cases correlates with the expression of α Syn in gut biopsies.⁹²

Bacterial and viral gut infections can thus have prominent effects on the gut and the brain. They influence gut permeability resulting in systemic inflammation and neuroinflammation. Via the gutbrain axis gut infections can influence brain health and act as a trigger of synucleinopathy.

THE ROLE OF α-SYNUCLEIN DURING THE HOST RESPONSE AGAINST INFECTIONS

Point missense mutations in the *SNCA* locus but also gene duplication and triplication cause autosomal dominant PD, indicating that α Syn plays a central role in familial PD.²³² *SNCA* is also a risk factor for idiopathic PD as non-coding variants are associated with an increased risk of PD or DLB.²³³ α Syn is mostly studied for its role as a synaptic protein in neurons but new and compelling evidence is emerging that α Syn is also involved in immune function.

Several studies have now shown that α Syn has an important role in the innate and adaptive immune response.^{5,234} α Syn expression is carefully controlled during infections. $^{92-94}$ The way this happens is not fully understood, but the presence of inflammation-related regulatory regions in the promoter of α Syn could regulate its expression during inflammation. The human SNCA promoter includes a 34 kb upstream region that contains several additional regulatory regions including the interferon- or other cytokine responsive elements of STAT1 and NF1.²³⁵ These regulatory elements in the SNCA promoter could thus be transcriptionally activated by pro-inflammatory cytokines such as interferon- γ , IL-6 or growth factors that can cross the BBB.^{236,237}

The role of α Syn in the innate immune response

Innate immunity involves the recognition of specific ligands by PRRs and the production of inflammatory mediators to limit infections and initiate tissue repair. TLRs are expressed by immune and non-immune cells, including microglia, neurons, astrocytes, and oligodendrocytes.²³⁸ TLRs recognize a wide range of PAMPs¹⁵⁴ and DAMPs, including α Syn.¹³⁵

Increasing evidence thus suggests the involvement of TLRs in the pathogenesis of synucleinopathies.²³⁹ It has been reported that in postmortem PD brain tissue there is an increased expression of the microbial-associated molecular pattern (MAMPs) receptors, TLR4 and TLR2.^{240,241} In neuronal cell cultures, TLR2 activation increases a Syn protein levels. Since α Syn is recognized by TLRs, it could serve as a chemoattractant for the recruitment of T and B cells.⁹² In PD patients, neurons with a Syn pathology have increased expression of TLR2²⁴¹, which could make them particularly susceptible to the proinflammatory effects of pathogen derived MAMPs. Moreover, α Syn can increase the expression of TLRs in microglia to promote neuroinflammation,²⁴² Different high molecular weight assemblies of α Syn have varying affinities for TLRs and also activate the immune response with varying efficiencies.243-245

Besides the effects of α Syn on microglia, α Syn also modulates the astroglia immune response. The intracellular accumulation of α Syn in astrocytes is necessary for the induction of a proinflammatory response. While astrocytes internalize α Syn mediated exclusively by TLR2, proinflammatory cytokine expression can be mediated by both TLR2 and TLR4.^{245–247} These observations have led to the hypothesis that astroglial TLR2 could play a dual role in synucleinopathies. On one hand by protecting neurons by the internalization of potentially toxic extracellular α Syn, and on the other hand by triggering neuroinflammation.²³⁸

In the periphery, α Syn has similar effects on leukocytes via stimulation of TLRs. α Syn stimulates the maturation and activation of dendritic cells through TLR4.⁹² In mice, α Syn is required for both a normal pro-inflammatory response to bacterial peptidoglycan and the T cell mediated response to immunization in the peritoneal cavity.²⁴⁸ Interestingly, neurons that innervate the peritoneum seem to be the source of α Syn as a ligand for TLRs, supporting a mechanistic link between neuronal pathology and the immune response in the periphery.

The role of α Syn in the adaptive immune response

Adaptive immunity requires the activation of antigen-sensitized cytotoxic T cells and release of cytokines and chemokines in response to foreign molecules or microorganisms. While T cells help to eliminate pathogen infected cells, B cells produce antibodies to protect against extracellular microbes and toxic molecules.²⁴⁹ T cells can only recognize antigenic peptides displayed by major histocompatibility complex (MHC)-I or MHC-II molecules on the cell surface²⁴⁹

MHC-I molecules are expressed in all cells to present antigen peptides to CD8+T cells, which allows them to identify and eliminate infected cells.²⁴⁹ Under normal conditions CD8 + T cells are tolerant to autologous or self-proteins. However, during infections cells can express microbial genes and present these 'non-self' antigenic peptides allowing CD8+T cells to eliminate these cells. Nigral dopaminergic neurons express MHC-I suggesting that these neurons are potentially susceptible to T cell mediated cytotoxicity²⁵⁰. T cell infiltration in areas of the brain with high α Syn burden is seen in PD and DLB patients as well as in the brain of synucleinopathy animal models.147,251-253 This reinforces the idea that the adaptive immune response is a potential driver of pathogenesis. Furthermore, the infiltration of cytotoxic CD8 + T cells has been seen as an early pathological event that precedes α Syn aggregation and neurodegeneration in the substantia nigra.²⁵⁴

While MHC-I molecules are expressed ubiquitously, MHC-II molecules are expressed on immune antigen presenting cells. MHC-II molecules present peptides from endocytosed molecules to CD4+T cells to activate them.²⁴⁹ Genome-wide association studies (GWAS) have found an association between PD risk and the human leukocyte antigen isotype DR (HLA-DR) alleles. HLA-DR is a cell receptor heterodimer expressed on the surface of antigen presenting cells including microglia and macrophages. Reactive microglia and border-associated macrophages positive for HLA-DR are abundant in the substantia nigra of PD cases.^{143,255,256} HLA-DR are part of the MHC-II family, and the genetic link between HLA-DR variants and an increased risk of developing PD supports a role of the adaptive immune response in PD pathology.^{257–259}

Within these lines, circulating T cells from PD patients recognize specific aSyn peptides.²⁶⁰ Pathological α Syn can trigger an immune response and inflammation, producing autoreactive T cells.²⁶¹ This loss of self-tolerance could be associated with environmental factors including infections.²⁶² Here, repeated exposure to α Syn in the context of infection could thereby cause cellular damage. Furthermore, the HLA-DR allele HLA-DRB1*15:01 has been linked to an increased risk of PD, and this allele may also be involved in the presentation of a Syn peptides to T cells.²⁶³ Peripherally circulating CD4 + and CD8+T cells derived from PD patients produce cytokines in response to α Syn, suggesting there may be a chronic memory T cell response in PD.²⁶⁴ MHC-II expression and T cell infiltration are essential for neuroinflammation and loss of nigral dopaminergic neurons in a mouse model of α Syn overexpression.265

It is known that infections that activate the immune system may exacerbate existing conditions or trigger new disease activity in autoimmune diseases. For example, urinary tract infections or respiratory tract infections, which are common, have been associated with increased risk of relapses in multiple sclerosis.^{266,267} The immune response to these infections might lead to increased activity of autoreactive T cells that attack the central nervous system. In the case of PD, and given the potential role of infections in synucleinopathies, peripheral infections could trigger autoimmune activation and cause a Syn autoreactive immune cells to attack α Syn presenting cells and contribute to neurodegeneration via a similar mechanism although it needs to be investigated if such links might indeed exist.

An emerging role for the immune response at the brain border

CNS or border-associated macrophages comprise perivascular, meningeal, and choroid plexus macrophages with phagocytic capacity, which support the BBB by scavenging harmful molecules and promoting efficient immune surveillance, including antigen presentation and cytokine production.^{268,269} Leptomeningeal macrophages migrate postnatally into the perivascular space and differentiate into perivascular macrophages.²⁷⁰ As described above, under disease conditions with alterations of the BBB, such as in synucleinopathies or neuroinflammation²⁷¹ peripheral immune cells potentially invade the CNS and encounter the response of border-associated macrophages.^{272,273} Furthermore, studies in PD models indicate that peripheral monocyte infiltration plays a critical role in α Syn-induced neuroinflammation and neurodegeneration.274

There are only few studies which address the role of border-associated macrophages as potential gatekeepers in such CNS immune cell invasion. A recent study in a model of synucleinopathy has shown that border-associated macrophages, not microglia, are responsible for CD4+T cell antigen presentation and recruitment necessary for tissue infiltration and cytokine production upon α Syn pathology.²⁵⁵ Interestingly, α Syn oligomers were detected in perivascular macrophages in synucleinopathy brain tissue.²⁷⁵ Depletion of borderassociated macrophages prevented aSyn-induced neuroinflammation, including microglial reactivity, T cell infiltration, and monocyte recruitment.²⁵⁵ Whether this has therapeutic implications for synucleinopathies requires further research.²⁷⁶

If T-cell and CNS macrophage responses are associated with α Syn pathology in synucleinopathies, then infections that cause neuroinflammation could further exacerbate these processes and thereby neurodegeneration. Proliferation of perivascular macrophages was shown to contribute to the development of encephalitic lesions during simian immunodeficiency virus (SIV) infection of adult macaques.²⁷⁷ Interestingly, border-associated macrophages but not disease-associated microglia show overt long-term transcriptional alterations which persist after resolution of trypanosoma brucei infection in mice.²⁷⁸ Thus, single infections may alter the brain's resident immune response at the border. Border-associated macrophages also play a critical role in protecting the brain parenchyma from viruses. In mice infected with lymphocytic choriomeningitis virus (LCMV), depletion of border-associated macrophages prior to infection or conditional deletion of interferonreceptor signaling pathways in myeloid cells resulted in extensive viral spread into the CNS. This appeared to be mediated by MHC-II-positive meningeal macrophages, and low numbers of these cells, as seen upon LPS challenge, correlated with higher viral load.²⁷⁹

Altogether, available evidence supports the involvement of brain resident and peripheral immune cells driving pathological events that can lead to neurodegeneration in synucleinopathies. Many of these cellular responses could potentially be triggered by infections and further aggravated by genetic risk factors.

INFECTIONS INTERACT WITH GENETIC RISK FACTORS OF SYNUCLEINOPATHIES

Within the hypothesis that infections might be involved in the etiology of sporadic forms of synucleinopathy, an important role has been proposed for genetic risk factors as 'facilitators' of disease.²⁸⁰ Certain pathogens possibly initiate or exacerbate the neurodegenerative process but in most individuals that are exposed to pathogens, infection will not lead to the onset of disease. It is likely that in most cases, infections will not cause neurological disease as research suggests that infections and sustained pathological inflammation needs to interact with genetic vulnerabilities for synucleinopathy to develop.^{5,280} (Fig. 1). The low penetrance and heritability of most PD or DLB-linked variants, suggest that cumulative exposure to environmental factors such as inflammatory toxins, pesticides or infections may be necessary to reach a level of cellular stress where these genetic factors become physiological relevant and lead to neurodegeneration.

Infections can trigger the immune system to release inflammatory molecules that can infiltrate the central nervous system, leading to neuroinflammation. Overlapping affected signaling pathways between neuroinflammation and neurodegenerative diseases reinforce the notion of a potentially positive feedback loop leading to cellular stress and dysfunction. Multiple PD-associated genes encode proteins with roles in the regulation of cellular transport and vesicle function, critical for the development of the immune response.^{281,282} PD-linked variants in mitochondrial genes also affect mitochondrial function important for the elimination of pathogens and danger signaling during infections. It is possible that genetic risk factors could gain physiological relevance under this cellular stress condition, further amplifying the pathological process. Environmental factors such as bacterial or viral infections but also gut dysbiosis or exposure to toxins may interact with genetic susceptibility in predisposed individuals to influence disease risk.

The relationship between specific PD risk genes and susceptibility to infections is an area of ongoing research. While only a small fraction of genes are implicated in familial PD, there is a myriad of gene variants that potentially increase the susceptibility to develop idiopathic PD. These risk factors are likely upstream in the disease process and many of these genes are implicated in immune function and neuroinflammation. Variants in several of these genes (*SNCA*, *LRRK2*, *GBA*, *PRKN*, *PINK1*, etc.) could potentially affect our response to infections and modulate inflammatory processes, which may in turn influence the risk of developing PD.

On the other hand, some genetic variants associated with PD may confer protection against infections or modulate the immune response in beneficial ways. Variants in genes involved in innate immunity or pathogen recognition may enhance the body's ability to combat infections.

During infections, LRRK2 PD variants can modulate the susceptibility to different infections.²⁸³⁻²⁸⁵ For example, LRRK2 G2019S, one of the most common risk variants of PD, is known to potentially protect against respiratory infections by modulating bacterial lung infection.²⁸⁶ It therefore seems that bacterial infections in the lung (where LRRK2 is highly expressed) could interact with LRRK2 risk, but whether other types of infections might as well be associated with this particular risk factor is unknown. It will be important for future studies to examine whether such unique host-pathogens interactions might exist. Genetic risk factors may modulate the onset or the course of PD in a manner that depends on the type of pathogen.²⁸⁰ Knowing which risk variants might interact with which type of pathogen could aid in the prevention and diagnosis of different subtypes of PD or other synucleinopathies.

In the case of α Syn, several experimental studies have shown that *SNCA* deletion results in an increased susceptibility to West Nile virus, Venezuelan equine encephalitis, reovirus and *Salmonella* *typhimurium* infections.^{93,94} It has been proposed that this enhanced susceptibility could be due to the role of α Syn in modulating proteostasis and interferon signaling and that α Syn has an important role on the host immune response during infection.^{94,157}

Based on the biological classification of PD,^{287,288} the development of some subtypes of PD could rely on a unique set of virulent interactions. On the one hand, pathogens can cause the clinical manifestations of PD, but in the absence of α Syn pathology (Fig. 1). These sporadic cases are distinct from the post-encephalitic cases of parkinsonism, but they do trigger dopamine-specific neurodegeneration after exposure to a specific pathogen.²⁸⁰ Experimental models have shown that in the presence of PD risk factors, gut infections can directly lead to dopamine-ergic degeneration, even in the absence of α Syn pathology.^{289,290}

On the other hand, infections can directly trigger α Syn aggregation and lead to the deposition of α Syn in peripheral sites.^{44,248,291,292} α Syn could then be prone to spread from these sites and trigger pathology in the CNS via connected regions. However, despite the presence of Lewy bodies, these types of PD might be fundamentally different and therefore require a different preventive or therapeutic approach due to distinct underlying biological mechanisms.

For instance, for the same risk factors PD patients can be either negative or positive for α Syn pathology. Mutations in LRRK2 are some of the most common PD-linked genetic risks and contribute to both genetic and sporadic forms of the disease.^{293–295} Some LRRK2 carriers with idiopathic PD have no detectable α Syn pathology, whereas other LRRK2 carriers are positive for α Syn pathology (Fig. 1). Accumulating evidence suggests that LRRK2 is involved in inflammation and the immune response.²⁹⁶ LRRK2 is expressed in high levels in immune cells^{297–300} and can modulate proinflammatory cytokine production.³⁰¹ The cellular function of LRRK2 is important for many cellular processes including vesicular trafficking, autophagy, and lysosomal function³⁰⁰.

The appearance of PD symptoms in the absence of α Syn pathology has important implications, as it has been debated if α Syn is required for the clinical diagnosis of PD. α Syn is a cardinal feature in PD and other synucleinopathies, but in some cases pathological α Syn aggregates are undetectable via diagnostic assays or even in the brain. For those patients, PD is likely not a synucleinopathy and the mechanism of disease and the biology of disease would be fundamentally different.

Such examples can also be found in cases with mutations in the mitochondrial genes PINK1 and Parkin¹⁰⁶. The phenotype in these patients could as well be linked to infections as these genes have been associated with increased susceptibility to intracellular pathogens.^{302,303} Parkin plays a key role in the innate immune response to intracellular infections by targeting them for ubiquitin-mediated autophagy in a similar role that this protein plays in mitophagy.³⁰⁴ PINK1 is functionally implicated in the response to gut bacterial infection in PD models, where Pink1 deficient mice develop motor deficit after infection²⁹⁰. It has been proposed that in the absence of Pink1 mitochondria specific CD8 + T cells can contribute to the degeneration of dopaminergic neurons²⁹⁰, adding to the evidence for a potential autoimmune mechanism in Pink1/Parkin-related PD.

Only little is known about how the underlying biology of PD and an individual's clinical presentation are potentially linked with the exposome. It is possible that exposure to different pathogens can modulate differential disease outcomes in people with the same genetic risk. Future research will need to investigate how these pathological interactions are mediated and can be prevented. In addition, research will need to carefully assess the patient's exposome in the context of their genetic risk.

Based on the available epidemiological studies we suggest that microbial pathogens can be a precipitating factor in the etiology of PD. Except maybe for very rare cases, virulent pathogens do not directly cause PD, but they are a potential upstream determinant in the disease pathway. Certain types of pathogens might uniquely interact with genetic risk in predisposed individuals. In the case of DLB and MSA only little is known about the types of pathogens that could trigger disease, as epidemiological studies about infectious pathogens in DLB and MSA are scarce. Nevertheless, infections can have lasting effects on the brain, via direct or systemic routes, and chronic effects can outlast the acute effects of the initial infection.

FUTURE DIRECTIONS

Implications for treatment, diagnosis, and prevention

PD is the second most common and the fastest growing neurodegenerative disease affecting more

than 4 million people worldwide and its prevalence has doubled in the last 25 years.⁶⁷ This increase persists even after correcting for aging suggesting that changing environmental factors could be important players in the pathophysiology of PD. Here, attention has been given to the role of pesticides, but less attention has been given to the role of other inflammatory agents, such as microbial pathogens.

More recently, the neurological symptoms observed after SARS-CoV-2 infection have brought new attention to the long-term neuroinflammatory effects on brain health after infection. The evidence reviewed here suggests that infections could have a role in the etiology of synucleinopathies (Fig. 1). Infections are part of the exposome, and this includes the total environmental exposures that individuals encounter throughout their lives, including factors such as pesticides, heavy metals, air pollution, solvents, and dietary habits. In PD and related synucleinopathies, these exposures may play a role in disease onset or disease development. ³⁰⁵ Studies have linked pesticides, herbicides like paraquat and rotenone, heavy metals such as lead and manganese, air pollution, and certain industrial chemicals (trichloroethylene, perchloroethylene or polychlorinated biphenyls) to an increased risk of PD.^{305,306} When this is further accompanied by aging and immunosenescence (the inability of the immune system to appropriately respond to infection), exposure to environmental factors will lead to increased susceptibility and cause mild or chronic neuroinflammation.³⁰⁷ Understanding the contribution of different environmental factors is thus crucial for developing preventive strategies and identifying modifiable risk factors for PD.

Because of the clinical heterogeneity in synucleinopathies, subcategorization or stratification of at-risk patients will be key to achieve better therapeutic results.³⁰⁸ Even though in recent years great progress has been made in the development of molecular diagnosis for synucleinopathies, including protein seeding amplification assays, skin biopsies and new imaging modalities, there are no early diagnostic tests or disease progression markers that could aid in diagnosis or taking preventive measure in the case of modifiable genetic risk factors. Identifying individual risk factors, genetic or environmental, is imperative to perform personalized risk assessment and enable early intervention. It would allow preventive measures to be taken such as lifestyle intervention, diet, exercise or even medication to mitigate risk factors. By understanding an individual's risk,

preventive therapies could have the potential to delay disease onset or improve disease outcomes for those who are at risk of developing PD.

It will also be important to study and consider stratification to evaluate the efficacy of preventive measures. For example, despite the abundant evidence of the link between influenza infections and PD, a recent study based in the UK and Finland found no association with reduced risk of PD and influenza vaccination after 12 years, although a significant association was found between influenza infection and other types of dementia.⁴³ This suggests that specific host-pathogen interactions might be important in neurological disorders where co-pathologies are present. Recognizing familial or genetic risk factors is vital for making informed decisions about vaccination.

Understanding these risks helps tailor recommendations and emphasizes the importance of vaccination in protecting against potential health threats, especially for those who are predisposed. A downstream consequence of peripheral or systemic infection is neuroinflammation.³⁰⁹ Neuroinflammatory changes can remain present in the brain long after resolution of infection or even in cases where the pathogen cannot be detected in the brain.^{181,182,310} It has been suggested that the neurodegenerative process is driven by a long lasting low-grade neuroimmune response rather than by direct viral neuroinvasion.^{181,182,310} Persistent neuroinflammation could help the brain to protect itself from subsequent insults via a primed innate immune system, but at the same time it will aggravate the age-related pathological process.

Much effort has been put into developing therapies that efficiently target neuroinflammation. Several of these approaches include anti-inflammatory medication or the targeted modulation of microglial activity. Another approach is to inhibit the activation of the inflammasome.³¹¹ Here, several compounds are being studied for their potential to inhibit inflammasome activation directly.³¹² These include small molecule inhibitors targeting specific inflammasome components.^{311,312} Immunomodulatory therapies, including monoclonal antibodies aim to target specific immune cells or cytokines, which are also being investigated to regulate the immune response³¹³. Lifestyle factors, such as diet, exercise, and stress reduction techniques, may help reduce inflammation and oxidative stress since they have been shown to ameliorate motor- and non-motor symptoms in PD.³¹⁴⁻³¹⁹

The recent and increasing popularity of GLP-1 receptor agonists, which are currently used for the treatment of type-2 diabetes, could also help to reduce inflammation in synucleinopathies. GLP-1 receptor agonists are being studied for the treatment of PD as they are potent inhibitors of neuroinflammation via inhibiting NF-kB activation in microglia.^{320,321} In turn, this reduces the conversion of astrocytes into neurotoxic astrocytes.³²⁰ Recent clinical studies have shown that the progression of motor and non-motor symptoms is slowed down or can even be stabilized when the GLP-1 receptor agonists are taken early in the disease course.^{320–322} While larger trials are ongoing, much research is needed to further evaluate the effects of GLP-1 receptor agonists in the treatment of early PD.

These anti-inflammatory therapeutic approaches could be combined with existing therapies and it is likely that anti-inflammatory therapy might be more successful during early stages. Precision medicine in PD or other synucleinopathies aims to tailor medical treatment to the individual needs of each patient and it should consider factors such as genetic makeup, environmental exposure to risk factors, and lifestyle.^{308,323} We should therefore consider any prior but also future exposure to infectious or inflammatory pathogens as it is an integral part of the patients' environment and lifestyle. By identifying individuals who may benefit from anti-inflammatory treatments based on their genetic predispositions other tailored anti-inflammatory therapies can then be advised to address their specific needs.

Implications for research

If infections are indeed involved in the etiology of synucleinopathies, it will necessitate research into the pathogens or type of infections that can contribute to the development of pathology. This will involve studying potential mechanisms by which infections can trigger or exacerbate neurodegeneration. Some of these mechanisms might overlap with the mechanisms involved in toxins, pesticides or other inflammatory molecules that could trigger pathology via environmental exposure.

We would also need to address methodological challenges. The lack of molecular diagnosis tools, the long prodromal phases and the clinical heterogeneity make it especially challenging to evaluate the effect of infections on the disease process and to establish a causal effect in the etiology of synucleinopathies. Not only the type of infection but also the timing of infection might be crucial for when pathology might or might not develop. New animal models that investigate how acute or chronic inflammation caused by infections could shed new light on how these diseases might arise.

It has been proposed that some of PD-linked pathogenic mutations with high prevalence and ancient origin (e.g., LRRK2, PINK, GBA and SNCA) might have provided a survival advantage when life expectancy was shorter and when infections were more common.^{296,300,305} Impaired lysosomal function might influence susceptibility to neurodegenerative processes but could potentially have modified immune responses favorably against certain pathogens. Some studies have also suggested that overexpression of a Syn can restrict viral replication or viral assembly.³²⁴ Now, with improved hygiene, and better anti-microbial treatments, these effects are less beneficial and exposure to virulent pathogens could instead lead to age-related neurodegenerative diseases as the same genetic risk factors will act as a disease facilitator at a later age.²⁸⁰

The notion that these genes provided advantages in the face of infectious diseases is thus supported by their roles in the immune system. This is also seen in other neuroinflammatory diseases, like multiple sclerosis, where age-related and disease associated genetic variants were positively selected because they offered an evolutionarily advantage during times when infections were more common.³²⁵ However, more detailed epidemiological and genetic studies are needed to substantiate these hypotheses and theories. This includes studying how these genetic variants might have been selected in populations when faced with high burdens of infectious diseases. Understanding these mechanisms could provide insights into why these genes are preserved in the human genome despite their association with disease in today's context. Understanding these risk genes will help us to understand their physiological role and the molecular mechanisms in which they are involved during infection or inflammation. It could identify new potential therapeutic targets or help to devise new strategies for the prevention or treatment of high-risk populations. Importantly, it also warrants the use of novel therapies aimed at these pathways, as inhibiting the cellular pathways important for the immune response could potentially increase the susceptibility towards certain infectious pathogens in a population that is already more immunosenescent.

CONCLUSION

The hypothesis that infections might play a role in the etiology of synucleinopathies provides a framework for future research. The involvement of genetic risk factors in synucleinopathies, particularly those that regulate the immune response, underscores the potential interplay between genetic predispositions and environmental inflammatory triggers. Advancements in epidemiological methods have begun to shed light on specific pathogens that may be implicated in the development of these neurodegenerative diseases. By integrating genetic, immunological, and epidemiological insights, a multidisciplinary approach is required to further explore the origins and progression of synucleinopathies. Acknowledging the potential role of infections not only enriches our understanding of how synucleinopathies might arise but it also paves the way for the development of novel diagnostic tools and targeted treatments. By combining these approaches, preventive or precision medicine might hold potential to improve early diagnosis and treatment, to potentially delay disease onset and offer effective personalized care.

ACKNOWLEDGMENTS

The authors have no acknowledgments to report.

FUNDING

The authors have no funding to report.

CONFLICT OF INTEREST

W.P. has received commercial support for research from Myrtelle Gene Therapeutics. W.P. has received support as a consultant from Roche.

REFERENCES

- Sakowski SA, Koubek EJ, Chen KS, et al. Role of the exposome in neurodegenerative disease: recent insights and future directions. *Annals of Neurology* 2024; 95: 635–652.
- Calabresi P, Mechelli A, Natale G, et al. Alpha-synuclein in Parkinson's disease and other synucleinopathies: from overt neurodegeneration back to early synaptic dysfunction. *Cell Death Dis* 2023; 14: 176.
- 3. Goedert M, Jakes R and Spillantini MG. The synucleinopathies: twenty years on. *J Parkinsons Dis* 2017; 7: S51.
- 4. Spillantini MG, Crowther RA, Jakes R, et al. Filamentous alpha-synuclein inclusions link multiple system atrophy

with Parkinson's disease and dementia with Lewy bodies. *Neurosci Lett* 1998; 251: 205–208.

- Tulisiak CT, Mercado G, Peelaerts W, et al. Can infections trigger alpha-synucleinopathies?. *Prog Mol Biol Transl Sci* 2019; 168: 299–322.
- Di Vito A, Donato A, Bria J, et al. Encephalitis lethargica. What is still wrong? *Int J Immunopathol Pharmacol* 2023; 37: 1–7.
- Dourmashkin RR. What caused the 1918–30 epidemic of encephalitis lethargica? J R Soc Med 1997; 90: 515–520.
- Ravenholt RT and Foege WH. 1918 Influenza, encephalitis lethargica, parkinsonism. *Lancet* 1982; 320: 860–864.
- 9. Dimsdale H. Changes in the Parkinsonian syndrome in the Twentieth Century. *Q J Med* 1946; 15: 155–170.
- 10. Encephalitis lethargica. Lancet 1981; 318: 1396–1397.
- Jellinger KA. Absence of α-synuclein pathology in postencephalitic parkinsonism. *Acta Neuropathol* 2009; 118: 371–379.
- Mccall S, Henry JM, Reid AH, et al. Influenza RNA not detected in archival brain tissues from acute encephalitis lethargica cases or in postencephalitic Parkinson cases. J Neuropathol Exp Neurol 2001; 60: 696–704.
- Josephs KA, Parisi JE and Dickson DW. Alpha-synuclein studies are negative in postencephalic Parkinsonism of von economo. *Neurology* 2002; 59: 645–646.
- Geddes JF, Hughes AJ, Lees AJ, et al. Pathological overlap in cases of parkinsonism associated with neurofibrillary tangles: A study of recent cases of postencephalitic parkinsonism and comparison with progressive supranuclear palsy and Guamanian parkinsonism-dementia complex. *Brain* 1993; 116: 281–302.
- 15. Haynes GS. Encephalitis Lethargica. *Br Med J* 1921; 1: 849.
- Foley PB. Encephalitis lethargica and the influenza virus. III. The influenza pandemic of 1918/19 and encephalitis lethargica: neuropathology and discussion. *J Neural Transm (Vienna)* 2009; 116: 1309.
- Love S, Louis D and Ellison DW. Greenfield's Neuropathology. 8th ed. London: CRC Press, 2008.
- Ishii T and Nakamura Y. Distribution and ultrastructure of Alzheimer's neurofibrillary tangles in postencephalitic parkinsonism of Economo type. *Acta Neuropathol* 1981; 55: 59–62.
- Bopeththa BVKM and Ralapanawa U. Post encephalitic parkinsonism following dengue viral infection. *BMC Res Notes* 2017; 10: 655.
- He S, Yuan LP, Zhang JW, et al. Postencephalitic parkinsonism and selective involvement of substantia nigra. *Neurol Sci* 2015; 36: 653–655.
- Smeyne RJ, Noyce AJ, Byrne M, et al. Infection and risk of Parkinson's disease. J Parkinsons Dis 2021; 11: 31–43.
- Cohen ME, Eichel R, Steiner-Birmanns B, et al. A case of probable Parkinson's disease after SARS-CoV-2 infection. *Lancet Neurol* 2020; 19: 804–805.
- Fearon C, Mikulis DJ and Lang AE. Parkinsonism as a sequela of SARS-CoV-2 infection: pure hypoxic injury or additional COVID-19-related response? *Mov Disord* 2021; 36: 1483–1484.
- Faber I, Brandão PRP, Menegatti F, et al. Coronavirus disease 2019 and parkinsonism: a non-post-encephalitic case. *Mov Disord* 2020; 35: 1721–1722.
- Méndez-Guerrero A, Laespada-García MI, Gómez-Grande A, et al. Acute hypokinetic-rigid syndrome following SARS-CoV-2 infection. *Neurology* 2020; 95: E2109–E2118.

- Vlad B, Jelcic I and Balint B. Parkinsonism due to coxsackie B virus infection—case report and literature review. *Mov Disord Clin Pract* 2023; 10: S24–S28.
- Toledano M and Davies NWS. Enteroviral postencephalitic parkinsonism with evidence of impaired presynaptic dopaminergic function. *JAMA Neurol* 2016; 73: 1023–1025.
- Walters JH. Postencephalitic Parkinson syndrome after meningoencephalitis due to coxsackie virus group B, type
 N Engl J Med 1960; 263: 744–747.
- Dourmashkin RR, Mccall SA, Dourmashkin N, et al. Virus-like particles and enterovirus antigen found in the brainstem neurons of Parkinson's disease. *F1000Research* 2018; 7: 302.
- Pradhan S, Pandey N, Shashank S, et al. Parkinsonism due to predominant involvement of substantia nigra in Japanese encephalitis. *Neurology* 1999; 53: 1781–1786.
- Murgod UA, Muthane UB, Ravi V, et al. Persistent movement disorders following Japanese encephalitis. *Neurology* 2001; 57: 2313–2315.
- Misra UK and Kalita J. Movement disorders in Japanese encephalitis. J Neurol 1997; 244: 299–303.
- Cerna F, Mehrad B, Luby JP, et al. St. Louis encephalitis and the substantia nigra: MR imaging evaluation. *AJNR Am J Neuroradiol* 1999; 20: 1281.
- Robinson RL, Shahida S, Madan N, et al. Transient parkinsonism in West Nile Virus encephalitis. *Am J Med* 2003; 115: 252–253.
- Lenka A, Kamat A and Mittal SO. Spectrum of movement disorders in patients with neuroinvasive West Nile Virus infection. *Mov Disord Clin Pract* 2019; 6: 426.
- Burton JM, Kern RZ, Halliday W, et al. Neurological manifestations of West Nile Virus infection. *Can J Neurol Sci* 2004; 31: 185–193.
- Maramattom BV and Philips G. Acute parkinsonism with West Nile Virus infection. *Ann Indian Acad Neurol* 2023; 26: 801–803.
- Bhidayasiri R and Tarsy D. HIV-induced parkinsonism. *Curr Clin Neurol* 2012; 36: 44–45.
- Amod F, Holla VV, Ojha R, et al. A review of movement disorders in persons living with HIV. *Parkinsonism Relat Disord* 2023; 114: 105774.
- Mirsattari SM, Power C and Nath A. Parkinsonism with HIV infection. *Mov Disord* 1998; 13: 684–689.
- 41. Jang H, Boltz DA, Webster RG, et al. Viral parkinsonism. *Biochim Biophys Acta* 2009; 1792: 714–721.
- Cocoros NM, Svensson E, Szépligeti SK, et al. Long-term risk of Parkinson disease following influenza and other infections. *JAMA Neurol* 2021; 78: 1461–1470.
- Levine KS, Leonard HL, Blauwendraat C, et al. Virus exposure and neurodegenerative disease risk across national biobanks. *Neuron* 2023; 111: 1086–1093.e2.
- Peelaerts W, Mercado G, George S, et al. Urinary tract infections trigger synucleinopathy via the innate immune response. *Acta Neuropathol* 2023; 145: 541–559.
- 45. Hogg E, Frank S, Oft J, et al. Urinary tract infection in Parkinson's disease. *J Parkinsons Dis* 2022; 12: 743.
- Harris MA, Tsui JK, Marion SA, et al. Association of Parkinson's disease with infections and occupational exposure to possible vectors. *Mov Disord* 2012; 27: 1111–1117.
- Wang H, Liu X, Tan C, et al. Bacterial, viral, and fungal infection-related risk of Parkinson's disease: Metaanalysis of cohort and case–control studies. *Brain Behav* 2020; 10: e01549.

- Lai SW, Lin CH, Lin HF, et al. Herpes zoster correlates with increased risk of Parkinson's disease in older people A population-based cohort study in Taiwan. *Medicine* (*Baltimore*) 2017; 96: e6075.
- Ragozzino MW, Kurland LT and Rajput AH. Investigation of the association between herpes zoster and Parkinson's disease. *Neuroepidemiology* 1983; 2: 89–92.
- Camacho-Soto A, Faust I, Racette BA, et al. Herpesvirus infections and risk of Parkinson's disease. *Neurodegener Dis* 2021; 20: 97–103.
- Tunnicliffe L, Weil RS, Breuer J, et al. Herpes zoster and risk of incident Parkinson's disease in US Veterans: a matched cohort study. *Mov Disord* 2024; 39: 438–444.
- 52. Sasco AJ and Paffenbarger RS. Measles infection and Parkinson's disease. *Am J Epidemiol* 1985; 122: 1017–1031.
- Shen X, Yang H, Wu Y, et al. Meta-analysis: Association of Helicobacter pylori infection with Parkinson's diseases. *Helicobacter* 2017; 22: e12398.
- De Pedro-Cuesta J, Gudmundsson G, Abraira V, et al. Whooping cough and Parkinson's disease. *Int J Epidemiol* 1996; 25: 1301–1311.
- Martyn CN and Osmond C. Parkinson's disease and the environment in early life. J Neurol Sci 1995; 132: 201–206.
- Shen CH, Chou CH, Liu FC, et al. Association between tuberculosis and Parkinson disease: a nationwide, population-based cohort study. *Medicine (Baltimore)* 2016; 95: e2883.
- Tanner C, Albers K, Goldman S, et al. Seborrheic dermatitis and risk of future Parkinson's disease. *Neurology* 2012; 78: S42.001.
- Su TH, Yang HC, Tseng TC, et al. Antiviral therapy in patients with chronic hepatitis C is associated with a reduced risk of parkinsonism. *Mov Disord* 2019; 34: 1882–1890.
- Lin WY, Lin MS, Weng YH, et al. Association of antiviral therapy with risk of Parkinson disease in patients with chronic hepatitis C virus infection. *JAMA Neurol* 2019; 76: 1019–1027.
- Lehrer S and Rheinstein PH. Vaccination reduces risk of Alzheimer's disease, Parkinson's disease, and other neurodegenerative disorders. *Discov Med* 2022; 34: 97.
- Zhao H, Zhou X, Fu K, et al. Prospective cohort study evaluating the association between influenza vaccination and neurodegenerative diseases. *NPJ Vaccines* 2024; 9: 51.
- 62. Fink A, Hermann A, Günster C, et al. Influenza vaccination in patients with dementia and Parkinson's disease: an analysis of routine health insurance data for the years 2014–2019. Dtsch Arztebl Int 2022; 119: 66.
- Han C, Zhao ZZ, Chan P, et al. A large survey on COVID-19 vaccination in patients with Parkinson's disease and healthy population. *Vaccine* 2023; 41: 6483–6494.
- Phanhdone T, Drummond P, Meisel T, et al. Barriers to vaccination among people with Parkinson's disease and implications for COVID-19. *J Parkinsons Dis* 2021; 11: 1057.
- Tansey MG, Wallings RL, Houser MC, et al. Inflammation and immune dysfunction in Parkinson disease. *Nat Rev Immunol* 2022; 22: 657–673.
- Postuma RB and Berg D. Advances in markers of prodromal Parkinson disease. *Nat Rev Neurol* 2016; 12: 622–634.
- Kalia LV and Lang AE. Parkinson's disease. Lancet 2015; 386: 896–912.

- Berg D, Borghammer P, Fereshtehnejad SM, et al. Prodromal Parkinson disease subtypes – key to understanding heterogeneity. *Nat Rev Neurol* 2021; 17: 349–361.
- Fanciulli A and Wenning GK. Multiple-system atrophy. N Engl J Med 2015; 372: 249–263.
- Tada M, Onodera O, Tada M, et al. Early development of autonomic dysfunction may predict poor prognosis in patients with multiple system atrophy. *Arch Neurol* 2007; 64: 256–260.
- Palma JA, Norcliffe-Kaufmann L and Kaufmann H. Diagnosis of multiple system atrophy. *Auton Neurosci* 2018; 211: 15–25.
- McKay JH and Cheshire WP. First symptoms in multiple system atrophy. *Clin Auton Res* 2018; 28: 215–221.
- Nahimi A, Kinnerup MB, Sommerauer M, et al. Molecular imaging of the noradrenergic system in idiopathic Parkinson's disease. *Int Rev Neurobiol* 2018; 141: 251–274.
- Bohnen NI, Yarnall AJ, Weil RS, et al. Cholinergic system changes in Parkinson's disease: emerging therapeutic approaches. *Lancet Neurol* 2022; 21: 381–392.
- Augustis S, Saferis V and Jost WH. Autonomic disturbances including impaired hand thermoregulation in multiple system atrophy and Parkinson's disease. *J Neural Transm (Vienna)* 2017; 124: 965–972.
- Coon EA, Fealey RD, Sletten DM, et al. Anhidrosis in multiple system atrophy involves pre- and postganglionic sudomotor dysfunction. *Mov Disord* 2017; 32: 397.
- Egeberg A, Hansen PR, Gislason GH, et al. Exploring the association between rosacea and Parkinson disease: A Danish nationwide cohort study. *JAMA Neurol* 2016; 73: 529–534.
- Bellinger DL and Lorton D. Autonomic regulation of cellular immune function. *Autonomic Neuroscience* 2014; 182: 15–41.
- 79. Jänig W. Sympathetic nervous system and inflammation: A conceptual view. *Auton Neurosci* 2014; 182: 4–14.
- Fischer L, Barop H, Ludin SM, et al. Regulation of acute reflectory hyperinflammation in viral and other diseases by means of stellate ganglion block. A conceptual view with a focus on Covid-19. *Auton Neurosci* 2022; 237: 102903.
- Koyuncu OO, Hogue IB and Enquist LW. Virus infections in the nervous system. *Cell Host Microbe* 2013; 13: 379–393.
- Zhao YJ, Xu KF, Shu FX, et al. Neurotropic virus infection and neurodegenerative diseases: Potential roles of autophagy pathway. CNS Neurosci Ther 2024; 30: e14548.
- Braak H, Del Tredici K, Rüb U, et al. Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiol Aging* 2003; 24: 197–211.
- Hawkes CH, Del Tredici K and Braak H. Parkinson's disease: a dual-hit hypothesis. *Neuropathol Appl Neurobiol* 2007; 33: 599–614.
- Horsager J, Andersen KB, Knudsen K, et al. Brain-first versus body-first Parkinson's disease: a multimodal imaging case-control study. *Brain* 2020; 143: 3077–3088.
- 86. Peelaerts W, Bousset L, Baekelandt V, et al. α-Synuclein strains and seeding in Parkinson's disease, incidental Lewy body disease, dementia with Lewy bodies and multiple system atrophy: similarities and differences. *Cell Tissue Res* 2018; 373: 195–212.
- Fink AL. The aggregation and fibrillation of α-synuclein. Acc Chem Res 2006; 39: 628–634.
- Peelaerts W and Baekelandt V. α-Synuclein structural diversity and the cellular environment in α-synuclein

transmission models and humans. *Neurotherapeutics* 2023; 20: 67–82.

- Hijaz BA and Volpicelli-Daley LA. Initiation and propagation of α-synuclein aggregation in the nervous system. *Mol Neurodegener* 2020; 15: 19.
- Luk KC, Kehm V, Carroll J, et al. Pathological α-synuclein transmission in nontransgenic mice. *Science* 2012; 338: 949–953.
- Watts JC, Giles K, Oehler A, et al. Transmission of multiple system atrophy prions to transgenic mice. *Proc Natl Acad Sci U S A* 2013; 110: 19555–19560.
- Stolzenberg E, Berry D, Yang D, et al. A role for neuronal alpha-synuclein in gastrointestinal immunity. J Innate Immun 2017; 9: 456–463.
- 93. Tomlinson JJ, Shutinoski B, Dong L, et al. Holocranohistochemistry enables the visualization of α -synuclein expression in the murine olfactory system and discovery of its systemic anti-microbial effects. *J Neural Transm* 2017; 124: 721–738.
- Beatman EL, Massey A, Shives KD, et al. Alpha-synuclein expression restricts RNA viral infections in the brain. J Virol 2016; 90: 2767–2782.
- Sanchez EL and Lagunoff M. Viral activation of cellular metabolism. *Virology* 2015; 479–480: 609–618.
- 96. Eisenreich W, Heesemann J, Rudel T, et al. Metabolic host responses to infection by intracellular bacterial pathogens. *Front Cell Infect Microbiol* 2013; 3: 24.
- da Fonseca TL, Villar-Piqué A and Outeiro TF. The interplay between alpha-synuclein clearance and spreading. *Biomolecules* 2015; 5: 435.
- Burré J, Sharma M and Südhof TC. Cell biology and pathophysiology of α-synuclein. Cold Spring Harb Perspect Med 2018; 8: a024091.
- Killinger BA, Madaj Z, Sikora JW, et al. The vermiform appendix impacts the risk of developing Parkinson's disease. *Sci Transl Med* 2018; 10: eaar5280.
- Peng C, Trojanowski JQ and Lee VMY. Protein transmission in neurodegenerative disease. *Nat Rev Neurol* 2020; 16: 199–212.
- Rey NL, Wesson DW and Brundin P. The olfactory bulb as the entry site for prion-like propagation in neurodegenerative diseases. *Neurobiol Dis* 2018; 109: 226–248.
- Mercado G and Brundin P. Lots of movement in gut and Parkinson's research. *Trends Endocrinol Metabol* 2019; 30: 687–689.
- 103. Ding X, Zhou L, Jiang X, et al. Propagation of pathological α -synuclein from the urogenital tract to the brain initiates MSA-like syndrome. *iScience* 2020; 23: 101166.
- 104. Kim S, Kwon S-H, Kam T-I, et al. Transneuronal propagation of pathologic α-synuclein from the gut to the brain models Parkinson's disease. *Neuron* 2019; 103: 627–641.e7.
- Van Den Berge N, Ferreira N, Gram H, et al. Evidence for bidirectional and trans-synaptic parasympathetic and sympathetic propagation of alpha-synuclein in rats. *Acta Neuropathol* 2019; 138: 535–550.
- 106. Rey NL, George S, Steiner JA, et al. Spread of aggregates after olfactory bulb injection of α -synuclein fibrils is associated with early neuronal loss and is reduced long term. *Acta Neuropathol* 2018; 135: 65–83.
- 107. Rey NL, Steiner JA, Maroof N, et al. Widespread transneuronal propagation of α-synucleinopathy triggered in olfactory bulb mimics prodromal Parkinson's disease. *J Exp Med* 2016; 213: 1759–1778.

- Mason DM, Nouraei N, Pant DB, et al. Transmission of α-synucleinopathy from olfactory structures deep into the temporal lobe. *Mol Neurodegener* 2016; 11: 412–459.
- Borghammer P. The α-synuclein origin and connectome model (SOC model) of Parkinson's disease: explaining motor asymmetry, non-motor phenotypes, and cognitive decline. *J Parkinsons Dis* 2021; 11: 455.
- Borghammer P and Van Den Berge N. Brain-first versus gut-first Parkinson's disease: a hypothesis. J Parkinsons Dis 2019; 9: S281–S295.
- Peelaerts W, Bousset L, Van Der Perren A, et al. α-Synuclein strains cause distinct synucleinopathies after local and systemic administration. *Nature* 2015; 522: 340–344.
- Lohmann S, Bernis ME, Tachu BJ, et al. Oral and intravenous transmission of α-synuclein fibrils to mice. *Acta Neuropathol* 2019; 138: 515–533.
- Ayers JI, Brooks MM, Rutherford NJ, et al. Robust central nervous system pathology in transgenic mice following peripheral injection of α-synuclein fibrils. *J Virol* 2017; 91: e02095-16.
- 114. Kuan WL, Stott K, He X, et al. Systemic α -synuclein injection triggers selective neuronal pathology as seen in patients with Parkinson's disease. *Mol Psychiatry* 2021; 26: 556.
- 115. Reyes JF, Ekmark-Léwen S, Perdiki M, et al. Accumulation of alpha-synuclein within the liver, potential role in the clearance of brain pathology associated with Parkinson's disease. *Acta Neuropathol Commun* 2021; 9: 46.
- Braun F, Abed A, Sellung D, et al. Accumulation of αsynuclein mediates podocyte injury in Fabry nephropathy. *J Clin Invest* 2023; 133: e157782.
- 117. Recasens A, Carballo-Carbajal I, Parent A, et al. Lack of pathogenic potential of peripheral α-synuclein aggregates from Parkinson's disease patients. *Acta Neuropathol Commun* 2018; 6: 8.
- Schweighauser M, Shi Y, Tarutani A, et al. Structures of αsynuclein filaments from multiple system atrophy. *Nature* 2020; 585: 464–469.
- 119. Yang Y, Shi Y, Schweighauser M, et al. Structures of α -synuclein filaments from human brains with Lewy pathology. *Nature* 2022; 610: 791–795.
- 120. Van der Perren A, Gelders G, Fenyi A, et al. The structural differences between patient-derived α-synuclein strains dictate characteristics of Parkinson's disease, multiple system atrophy and dementia with Lewy bodies. *Acta Neuropathol* 2020; 139: 977–1000.
- Just MK, Gram H, Theologidis V, et al. Alpha-synuclein strain variability in body-first and brain-first synucleinopathies. *Front Aging Neurosci* 2022; 14: 907293.
- 122. Melki R. Role of different alpha-synuclein strains in synucleinopathies, similarities with other neurodegenerative diseases. *J Parkinsons Dis* 2015; 5: 217–227.
- 123. Gracia P, Camino JD, Volpicelli-Daley L, et al. Multiplicity of α-synuclein aggregated species and their possible roles in disease. *Int J Mol Sci* 2020; 21: 8043.
- Bhopatkar AA and Kayed R. Flanking regions, amyloid cores, and polymorphism: the potential interplay underlying structural diversity. *J Biol Chem* 2023; 299: 105122.
- Peelaerts W and Baekelandt V. α-Synuclein strains and the variable pathologies of synucleinopathies. *J Neurochem* 2016; 256–274.
- Candelise N, Schmitz M, Thüne K, et al. Effect of the micro-environment on α-synuclein conversion and impli-

cation in seeded conversion assays. *Transl Neurodegener* 2020; 9: 5.

- Goralski TM, Meyerdirk L, Breton L, et al. Spatial transcriptomics reveals molecular dysfunction associated with cortical Lewy pathology. *Nat Commun* 2024; 15: 2642.
- 128. Henderson MX, Cornblath EJ, Darwich A, et al. Spread of α-synuclein pathology through the brain connectome is modulated by selective vulnerability and predicted by network analysis. *Nat Neurosci* 2019; 22: 1248–1257.
- 129. Henderson MX, Sedor S, McGeary I, et al. Glucocerebrosidase activity modulates neuronal susceptibility to pathological α-synuclein insult. *Neuron* 2020; 105: 822–836.e7.
- 130. Henrich MT, Geibl FF, Lakshminarasimhan H, et al. Determinants of seeding and spreading of α -synuclein pathology in the brain. *Sci Adv* 2020; 6: eabc2487.
- 131. Surendranathan A, Su L, Mak E, et al. Early microglial activation and peripheral inflammation in dementia with Lewy bodies. *Brain* 2018; 141: 3415–3427.
- Kübler D, Wächter T, Cabanel N, et al. Widespread microglial activation in multiple system atrophy. *Mov Disord* 2019; 34: 564–568.
- 133. Gao C, Jiang J, Tan Y, et al. Microglia in neurodegenerative diseases: mechanism and potential therapeutic targets. *Signal Transduct Target Ther* 2023; 8: 359.
- Kawai T and Akira S. Toll-like receptors and their crosstalk with other innate receptors in infection and immunity. *Immunity* 2011; 34: 637–650.
- 135. El-Zayat SR, Sibaii H and Mannaa FA. Toll-like receptors activation, signaling, and targeting: an overview. *Bull Natl Res Cent* 2019; 43: 1–12.
- Castro-Gomez S and Heneka MT. Innate immune activation in neurodegenerative diseases. *Immunity* 2024; 57: 790–814.
- 137. Rocha NP, De Miranda AS and Teixeira AL. Insights into neuroinflammation in Parkinson's disease: from biomarkers to anti-inflammatory based therapies. *Biomed Res Int* 2015; 2015: 628192.
- Chen L, Mo M, Li G, et al. The biomarkers of immune dysregulation and inflammation response in Parkinson disease. *Transl Neurodegener* 2016; 5: 16.
- Stephenson J, Nutma E, van der Valk P, et al. Inflammation in CNS neurodegenerative diseases. *Immunology* 2018; 154: 204.
- Kline EM, Houser MC, Herrick MK, et al. Genetic and environmental factors in Parkinson's disease converge on immune function and inflammation. *Mov Disord* 2021; 36: 25–36.
- Kisler K and Zlokovic BV. How the brain regulates its own immune system. *Nat Neurosci* 2022; 25: 532–534.
- 142. Imamura K, Hishikawa N, Sawada M, et al. Distribution of major histocompatibility complex class II-positive microglia and cytokine profile of Parkinson's disease brains. *Acta Neuropathol* 2003; 106: 518–526.
- 143. McGeer PL, Itagaki S, Boyes BE, et al. Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains. *Neurology* 1988; 38: 1285–1291.
- Ouchi Y, Yoshikawa E, Sekine Y, et al. Microglial activation and dopamine terminal loss in early Parkinson's disease. *Ann Neurol* 2005; 57: 168–175.
- 145. Gerhard A, Pavese N, Hotton G, et al. In vivo imaging of microglial activation with [11C](R)-PK11195 PET in idiopathic Parkinson's disease. *Neurobiol Dis* 2006; 21: 404–412.

- 146. Doorn KJ, Moors T, Drukarch B, et al. Microglial phenotypes and toll-like receptor 2 in the substantia nigra and hippocampus of incidental Lewy body disease cases and Parkinson's disease patients. *Acta Neuropathol Commun* 2014; 2: 90.
- 147. Amin J, Holmes C, Dorey RB, et al. Neuroinflammation in dementia with Lewy bodies: a human post-mortem study. *Translational Psychiatry* 2020; 10: 267.
- Streit WJ and Xue QS. Microglia in dementia with Lewy bodies. Brain Behav Immun 2016; 55: 191–201.
- Gate D, Tapp E, Leventhal O, et al. CD4+T cells contribute to neurodegeneration in Lewy body dementia. *Science* 2021; 374: 868.
- Terkelsen MH, Klaestrup IH, Hvingelby V, et al. Neuroinflammation and immune changes in prodromal Parkinson's disease and other synucleinopathies. *J Parkinsons Dis* 2022; 12: S149–S163.
- Lauritsen J and Romero-Ramos M. The systemic immune response in Parkinson's disease: focus on the peripheral immune component. *Trends Neurosci* 2023; 46: 863–878.
- 152. Craig DW, Hutchins E, Violich I, et al. RNA sequencing of whole blood reveals early alterations in immune cells and gene expression in Parkinson's disease. *Nature Aging* 2021; 1: 734–747.
- Borsche M, Pereira SL, Klein C, et al. Mitochondria and Parkinson's disease: clinical, molecular, and translational aspects. *J Parkinsons Dis* 2021; 11: 45–60.
- 154. Aviner R and Frydman J. Proteostasis in viral infection: Unfolding the complex virus–chaperone interplay. *Cold Spring Harb Perspect Biol* 2020; 12: a034090.
- Ferecskó AS, Smallwood MJ, Moore A, et al. STINGtriggered CNS inflammation in human neurodegenerative diseases. *Biomedicines* 2023; 11: 1375.
- 156. Reinert LS, Lopušná K, Winther H, et al. Sensing of HSV-1 by the cGAS–STING pathway in microglia orchestrates antiviral defence in the CNS. *Nat Commun* 2016; 7: 13348.
- Monogue B, Chen Y, Sparks H, et al. Alpha-synuclein supports type 1 interferon signalling in neurons and brain tissue. *Brain* 2022; 145: 3622–3636.
- Main BS, Zhang M, Brody KM, et al. Type-1 interferons contribute to the neuroinflammatory response and disease progression of the MPTP mouse model of Parkinson's disease. *Glia* 2016; 64: 1590–1604.
- Sliter DA, Martinez J, Hao L, et al. Parkin and PINK1 mitigate STING-induced inflammation. *Nature* 2018; 561: 258–262.
- Hinkle JT, Patel J, Panicker N, et al. STING mediates neurodegeneration and neuroinflammation in nigrostriatal α-synucleinopathy. *Proc Natl Acad Sci U S A* 2022; 119: e2118819119.
- Fu J and Wu H. Structural mechanisms of nlrp3 inflammasome assembly and activation. *Annu Rev Immunol* 2023; 41: 301–316.
- Cheng J, Liao Y, Dong Y, et al. Microglial autophagy defect causes Parkinson disease-like symptoms by accelerating inflammasome activation in mice. *Autophagy* 2020; 16: 2193–2205.
- 163. Zhou Y, Lu M, Du RH, et al. MicroRNA-7 targets Nod-like receptor protein 3 inflammasome to modulate neuroinflammation in the pathogenesis of Parkinson's disease. *Mol Neurodegener* 2016; 11: 28.
- Gordon R, Albornoz EA, Christie DC, et al. Inflammasome inhibition prevents-synuclein pathology and dopaminergic neurodegeneration in mice. *Sci Transl Med* 2018; 10: eaah4066.

- 165. Sarkar S, Malovic E, Harishchandra DS, et al. Mitochondrial impairment in microglia amplifies NLRP3 inflammasome proinflammatory signaling in cell culture and animal models of Parkinson's disease. NPJ Parkinsons Dis 2017; 3: 30.
- Zhang C, Zhao M, Wang B, et al. The Nrf2-NLRP3caspase-1 axis mediates the neuroprotective effects of Celastrol in Parkinson's disease. *Redox Biol* 2021; 47: 102134.
- 167. Han X, Sun S, Sun Y, et al. Small molecule-driven NLRP3 inflammation inhibition via interplay between ubiquitination and autophagy: implications for Parkinson disease. *Autophagy* 2019; 15: 1860–1881.
- Ravichandran KA and Heneka MT. Inflammasomes in neurological disorders – mechanisms and therapeutic potential. *Nat Rev Neurol* 2024; 20: 67–83.
- Polverino P, Cocco A and Albanese A. Post-COVID parkinsonism: A scoping review. *Parkinsonism Relat Dis*ord 2024; 123: 106066.
- Lotz SK, Blackhurst BM, Reagin KL, et al. microbial infections are a risk factor for neurodegenerative diseases. *Front Cell Neurosci* 2021; 15: 691136.
- Olsen LK, Dowd E and McKernan DP. A role for viral infections in Parkinson's etiology? *Neuronal Signal* 2018; 2: 314–317.
- McGavern DB and Kang SS. Illuminating viral infections in the nervous system. *Nat Rev Immunol* 2011; 11: 318–329.
- 173. Galea I. The blood-brain barrier in systemic infection and inflammation. *Cell Mol Immunol* 2021; 18: 2489–2501.
- 174. Park SJ, Jin U and Park SM. Interaction between coxsackievirus B3 infection and α-synuclein in models of Parkinson's disease. *PLoS Pathog* 2021; 17: e1010018.
- 175. Niklasson B, Lindquist L, Klitz W, et al. Picornavirus may be linked to Parkinson's disease through viral antigen in dopamine-containing neurons of substantia nigra. *Microorganisms* 2022; 10: 599.
- 176. Philippens IHCHM, Böszörményi KP, Wubben JA, et al. SARS-CoV-2 causes brain inflammation and induces Lewy body formation in macaques. *bioRxiv* 2021; doi: https://doi.org/10.1101/2021.02.23.432474 [Preprint]. Posted May 05, 2021.
- 177. Marreiros R, Müller-Schiffmann A, Trossbach SV, et al. Disruption of cellular proteostasis by H1N1 influenza A virus causes α-synuclein aggregation. *Proc Natl Acad Sci* U S A 2020; 117: 6741–6751.
- Schreiber CS, Wiesweg I, Stanelle-Bertram S, et al. Sex-specific biphasic alpha-synuclein response and alterations of interneurons in a COVID-19 hamster model. *EBioMedicine* 2024; 105: 105191.
- 179. Li C, Song W, Chan JFW, et al. Intranasal infection by SARS-CoV-2 Omicron variants can induce inflammatory brain damage in newly weaned hamsters. *Emerg Microbes Infect* 2023; 12: 2207678
- Käufer C, Schreiber CS, Hartke AS, et al. Microgliosis and neuronal proteinopathy in brain persist beyond viral clearance in SARS-CoV-2 hamster model. *EBioMedicine* 2022; 79: 103999.
- Matschke J, Lütgehetmann M, Hagel C, et al. Neuropathology of patients with COVID-19 in Germany: a post-mortem case series. *Lancet Neurol* 2020; 19: 919–929.
- Dhib-Jalbut S, Gogate N, Jiang H, et al. Human microglia activate lymphoproliferative responses to recall viral antigens. *J Neuroimmunol* 1996; 65: 67–73.

- 183. Semerdzhiev SA, Fakhree MAA, Segers-Nolten I, et al. Interactions between SARS-CoV-2 N-protein and α-synuclein accelerate amyloid formation. ACS Chem Neurosci 2022; 13: 143–150.
- Nyström S and Hammarström P. Amyloidogenesis of SARS-CoV-2 spike protein. J Am Chem Soc 2022; 144: 8945–8950.
- 185. Zilio G, Masato A, Sandre M, et al. SARS-CoV-2mimicking pseudoviral particles accelerate α-synuclein aggregation *in vitro*. ACS Chem Neurosci 2024; 15: 215–221.
- 186. Jana AK, Lander CW, Chesney AD, et al. Effect of an amyloidogenic SARS-COV-2 protein fragment on αsynuclein monomers and fibrils. *J Phys Chem B* 2022; 126: 3648–3658.
- Albornoz EA, Amarilla AA, Modhiran N, et al. SARS-CoV-2 drives NLRP3 inflammasome activation in human microglia through spike protein. *Mol Psychiatry* 2023; 28: 2878–2893.
- Rhea EM, Logsdon AF, Hansen KM, et al. The S1 protein of SARS-CoV-2 crosses the blood–brain barrier in mice. *Nat Neurosci* 2020; 24: 368–378.
- Petrovszki D, Walter FR, Vigh JP, et al. Penetration of the SARS-CoV-2 spike protein across the blood-brain barrier, as revealed by a combination of a human cell culture model system and optical biosensing. *Biomedicines* 2022; 10: 188.
- Wenzel J, Lampe J, Müller-Fielitz H, et al. The SARS-CoV-2 main protease Mpro causes microvascular brain pathology by cleaving NEMO in brain endothelial cells. *Nat Neurosci* 2021; 24: 1522–1533.
- Chesney AD, Maiti B and Hansmann UHE. SARS-COV-2 spike protein fragment eases amyloidogenesis of α-synuclein. J Chem Phys 2023; 159: 015103.
- Shannon K and Vanden Berghe P. The enteric nervous system in PD: gateway, bystander victim, or source of solutions. *Cell Tissue Res* 2018; 373: 313–326.
- 193. Bedarf JR, Hildebrand F, Coelho LP, et al. Functional implications of microbial and viral gut metagenome changes in early stage L-DOPA-naïve Parkinson's disease patients. *Genome Med* 2017; 9: 39.
- 194. Shen T, Yue Y, He T, et al. The association between the gut microbiota and Parkinson's disease, a meta-analysis. *Front Aging Neurosci* 2021; 13: 636545.
- Li P, Killinger BA, Ensink E, et al. Gut microbiota dysbiosis is associated with elevated bile acids in Parkinson's disease. *Metabolites* 2021; 11: 29.
- Palacios N, Wilkinson J, Bjornevik K, et al. Metagenomics of the gut microbiome in Parkinson's disease: prodromal changes. *Ann Neurol* 2023; 94: 486–501.
- 197. Ramirez J, Guarner F, Bustos Fernandez L, et al. Antibiotics as major disruptors of gut microbiota. *Front Cell Infect Microbiol* 2020; 10: 572912.
- 198. Sun MF, Zhu YL, Zhou ZL, et al. Neuroprotective effects of fecal microbiota transplantation on MPTP-induced Parkinson's disease mice: Gut microbiota, glial reaction and TLR4/TNF-α signaling pathway. *Brain Behav Immun* 2018; 70: 48–60.
- Yan Y, Ren S, Duan Y, et al. Gut microbiota and metabolites of α-synuclein transgenic monkey models with early stage of Parkinson's disease. *NPJ Biofilms Microbiomes* 2021; 7: 69.
- Singh Y, Trautwein C, Romani J, et al. Overexpression of human alpha-Synuclein leads to dysregulated microbiome/metabolites with ageing in a rat

model of Parkinson disease. Mol Neurodegener 2023; 18: 44.

- 201. Bruggeman A, Vandendriessche C, Hamerlinck H, et al. Safety and efficacy of faecal microbiota transplantation in patients with mild to moderate Parkinson's disease (GUT-PARFECT): a double-blind, placebo-controlled, randomised, phase 2 trial. *EClinicalMedicine* 2024; 71: 102563.
- 202. Forsyth CB, Shannon KM, Kordower JH, et al. increased intestinal permeability correlates with sigmoid mucosa alpha-synuclein staining and endotoxin exposure markers in early Parkinson's disease. *PLoS One* 2011; 6: e28032.
- 203. Mahbub NU, Islam MM, Hong ST, et al. Dysbiosis of the gut microbiota and its effect on α-synuclein and prion protein misfolding: consequences for neurodegeneration. *Front Cell Infect Microbiol* 2024; 14: 1348279.
- Chandra R, Hiniker A, Kuo Y-M, et al. α-Synuclein in gut endocrine cells and its implications for Parkinson's disease. JCI Insight 2017; 2: 1–13.
- 205. Rodrigues PV, de Godoy JVP, Bosque BP, et al. Transcellular propagation of fibrillar α-synuclein from enteroendocrine to neuronal cells requires cell-to-cell contact and is Rab35-dependent. *Sci Rep* 2022; 12: 4168.
- 206. Hurley MJ, Menozzi E, Koletsi S, et al. α-Synuclein expression in response to bacterial ligands and metabolites in gut enteroendocrine cells: an *in vitro* proof of concept study. *Brain Commun* 2023; 5: fcad285.
- 207. Romano S, Savva GM, Bedarf JR, et al. Meta-analysis of the Parkinson's disease gut microbiome suggests alterations linked to intestinal inflammation. NPJ Parkinsons Dis 2021; 7: 27.
- Toh TS, Chong CW, Lim SY, et al. Gut microbiome in Parkinson's disease: New insights from meta-analysis. *Parkinsonism Relat Disord* 2022; 94: 1–9.
- 209. Unger MM, Spiegel J, Dillmann KU, et al. Short chain fatty acids and gut microbiota differ between patients with Parkinson's disease and age-matched controls. *Parkinsonism Relat Disord* 2016; 32: 66–72.
- Nishiwaki H, Ito M, Ishida T, et al. Meta-analysis of gut dysbiosis in Parkinson's disease. *Mov Disord* 2020; 35: 1626–1635.
- Erny D, De Angelis ALH, Jaitin D, et al. Host microbiota constantly control maturation and function of microglia in the CNS. *Nat Neurosci* 2015; 18: 965–977.
- Sampson TR, Debelius JW, Thron T, et al. Gut microbiota regulate motor deficits and neuroinflammation in a model of Parkinson's disease. *Cell* 2016; 167: 1469–1480.e12.
- Di Lorenzo F, De Castro C, Silipo A, et al. Lipopolysaccharide structures of Gram-negative populations in the gut microbiota and effects on host interactions. *FEMS Microbiol Rev* 2019; 43: 257–272.
- Zhao Y, Walker DI, Lill CM, et al. Lipopolysaccharidebinding protein and future Parkinson's disease risk: a European prospective cohort. *J Neuroinflammation* 2023; 20: 170.
- Wijeyekoon RS, Kronenberg-Versteeg D, Scott KM, et al. Peripheral innate immune and bacterial signals relate to clinical heterogeneity in Parkinson's disease. *Brain Behav Immun* 2020; 87: 473–488.
- Loffredo L, Ettorre E, Zicari AM, et al. Oxidative stress and gut-derived lipopolysaccharides in neurodegenerative disease: role of NOX2. Oxid Med Cell Longev 2020; 2020: 8630275.
- 217. Kim WG, Mohney RP, Wilson B, et al. Regional difference in susceptibility to lipopolysaccharide-induced neurotox-

icity in the rat brain: role of microglia. *J Neurosci* 2000; 20: 6309–6316.

- Herrera AJ, Castaño A, Venero JL, et al. The single intranigral injection of LPS as a new model for studying the selective effects of inflammatory reactions on dopaminergic system. *Neurobiol Dis* 2000; 7: 429–447.
- Castaño A, Herrera AJ, Cano J, et al. Lipopolysaccharide intranigral injection induces inflammatory reaction and damage in nigrostriatal dopaminergic system. *J Neurochem* 1998; 70: 1584–1592.
- Gao HM, Jiang J, Wilson B, et al. Microglial activationmediated delayed and progressive degeneration of rat nigral dopaminergic neurons: relevance to Parkinson's disease. J Neurochem 2002; 81: 1285–1297.
- 221. He Q, Yu W, Wu J, et al. Intranasal LPS-mediated Parkinson's model challenges the pathogenesis of nasal cavity and environmental toxins. *PLoS One* 2013; 8: e78418.
- 222. Kelly LP, Carvey PM, Keshavarzian A, et al. Progression of intestinal permeability changes and alpha-synuclein expression in a mouse model of Parkinson's disease. *Mov Disord* 2014; 29: 999–1009.
- 223. Gao HM, Zhang F, Zhou H, et al. Neuroinflammation and α-synuclein dysfunction potentiate each other, driving chronic progression of neurodegeneration in a mouse model of Parkinson's disease. *Environ Health Perspect* 2011; 119: 807–814.
- 224. Tanaka S, Ishii A, Ohtaki H, et al. Activation of microglia induces symptoms of Parkinson's disease in wild-type, but not in IL-1 knockout mice. *J Neuroinflammation* 2013: 10: 143.
- 225. Koukoulis TF, Beauchamp LC, Kaparakis-Liaskos M, et al. Do bacterial outer membrane vesicles contribute to chronic inflammation in Parkinson's disease? *J Parkinsons Dis* 2024; 14: 227–244.
- Bittel M, Reichert P, Sarfati I, et al. Visualizing transfer of microbial biomolecules by outer membrane vesicles in microbe-host-communication *in vivo. J Extracell Vesicles* 2021; 10: e12159.
- 227. Xie J, Cools L, Van Imschoot G, et al. Helicobacter pylori-derived outer membrane vesicles contribute to Alzheimer's disease pathogenesis via C3-C3aR signalling. *J Extracell Vesicles* 2023; 12: 12306.
- Vanaja SK, Russo AJ, Behl B, et al. Bacterial outer membrane vesicles mediate cytosolic localization of LPS and caspase-11 activation. *Cell* 2016; 165: 1106–1119.
- Schwartz K and Boles BR. Microbial amyloids–functions and interactions within the host. *Curr Opin Microbiol* 2013; 16: 93–99.
- 230. Chen SG, Stribinskis V, Rane MJ, et al. Exposure to the functional bacterial amyloid protein curli enhances alpha-synuclein aggregation in aged Fischer 344 rats and Caenorhabditis elegans. *Sci Rep* 2016; 6: 34477.
- Nerius M, Doblhammer G and Tamgüney G. GI infections are associated with an increased risk of Parkinson's disease. *Gut* 2020; 69: 1154–1156.
- 232. Deng H, Wang P and Jankovic J. The genetics of Parkinson disease. *Ageing Res Rev* 2018; 42: 72–85.
- 233. Maraganore DM, De Andrade M, Elbaz A, et al. Collaborative analysis of α -synuclein gene promoter variability and Parkinson disease. *JAMA* 2006; 296: 661–670.
- 234. Kasen A, Houck C, Burmeister AR, et al. Upregulation of α -synuclein following immune activation: Possible trigger of Parkinson's disease. *Neurobiol Dis* 2022; 166: 105654.

- Samarajiwa SA, Forster S, Auchettl K, et al. INTERFER-OME: the database of interferon regulated genes. *Nucleic Acids Res* 2009; 37: D852.
- Galic MA, Riazi K and Pittman QJ. Cytokines and brain excitability. *Front Neuroendocrinol* 2012; 33: 116–125.
- Banks WA, Kastin AJ and Broadwell RD. Passage of cytokines across the blood-brain barrier. *Neuroimmunomodulation* 1995; 2: 241–248.
- Li L, Acioglu C, Heary RF, et al. Role of astroglial toll-like receptors (TLRs) in central nervous system infections, injury and neurodegenerative diseases. *Brain Behav Immun* 2021; 91: 740–755.
- Mazzotta GM, Ceccato N and Conte C. Synucleinopathies take their toll: are TLRs a way to go? *Cells* 2023; 12: 1231.
- Drouin-Ouellet J, St-Amour I, Saint-Pierre M, et al. Tolllike receptor expression in the blood and brain of patients and a mouse model of Parkinson's disease. *Int J Neuropsychopharmacol* 2014; 18: pyu10.
- Dzamko N, Gysbers A, Perera G, et al. Toll-like receptor 2 is increased in neurons in Parkinson's disease brain and may contribute to alpha-synuclein pathology. *Acta Neuropathol* 2017; 133: 303–319.
- Béraud D, Twomey M, Bloom B, et al. α-synuclein alters toll-like receptor expression. *Front Neurosci* 2011; 5: 80.
- Codolo G, Plotegher N, Pozzobon T, et al. Triggering of inflammasome by aggregated α–synuclein, an inflammatory response in synucleinopathies. *PLoS One* 2013; 8: e55375.
- Daniele SG, Béraud D, Davenport C, et al. Activation of MyD88-dependent TLR1/2 signaling by misfolded αsynuclein, a protein linked to neurodegenerative disorders. *Sci Signal* 2015; 8: ra45.
- 245. Kim C, Ho DH, Suk JE, et al. Neuron-released oligomeric α-synuclein is an endogenous agonist of TLR2 for paracrine activation of microglia. *Nat Commun* 2013; 4: 1562.
- Rannikko EH, Weber SS and Kahle PJ. Exogenous α-synuclein induces toll-like receptor 4 dependent inflammatory responses in astrocytes. *BMC Neurosci* 2015; 16: 57.
- 247. Fellner L, Irschick R, Schanda K, et al. Toll-like receptor 4 is required for α-synuclein dependent activation of microglia and astroglia. *Glia* 2013; 61: 349–360.
- Alam MM, Yang D, Li XQ, et al. Alpha synuclein, the culprit in Parkinson disease, is required for normal immune function. *Cell Rep* 2022; 38: 110090.
- Rock KL, Reits E, Neefjes J. Present yourself! By MHC class I and MHC class II molecules. *Trends Immunol* 2016; 37: 724–737.
- Cebrián C, Zucca FA, Mauri P, et al. MHC-I expression renders catecholaminergic neurons susceptible to T-cellmediated degeneration. *Nat Commun* 2014; 5: 3633.
- Brochard V, Combadière B, Prigent A, et al. Infiltration of CD4+ lymphocytes into the brain contributes to neurodegeneration in a mouse model of Parkinson disease. *J Clin Invest* 2009; 119: 182–192.
- Iba M, Kim C, Sallin M, et al. Neuroinflammation is associated with infiltration of T cells in Lewy body disease and α-synuclein transgenic models. *J Neuroinflammation* 2020; 17: 214.
- Richter F, Stanojlovic M, Käufer C, et al. A mouse model to test novel therapeutics for Parkinson's disease: an update on the Thy1-aSyn ("line 61") mice. *Neurotherapeutics* 2023; 20: 97–116.

- 254. Galiano-Landeira J, Torra A, Vila M, et al. CD8 T cell nigral infiltration precedes synucleinopathy in early stages of Parkinson's disease. *Brain* 2020; 143: 3717–3733.
- Schonhoff AM, Figge DA, Williams GP, et al. Borderassociated macrophages mediate the neuroinflammatory response in an alpha-synuclein model of Parkinson disease. *Nat Commun* 2023; 14: 3754.
- Benskey MJ, Sellnow RC, Sandoval IM, et al. Silencing alpha synuclein in mature nigral neurons results in rapid neuroinflammation and subsequent toxicity. *Front Mol Neurosci* 2018; 11: 36.
- Nalls MA, Pankratz N, Lill CM, et al. Large-scale metaanalysis of genome-wide association data identifies six new risk loci for Parkinson's disease. *Nat Genet* 2014; 46: 989–993.
- Ahmed I, Tamouza R, Delord M, et al. Association between Parkinson's disease and the HLA-DRB1 locus. *Mov Disord* 2012; 27: 1104–1110.
- Hamza TH, Zabetian CP, Tenesa A, et al. Common genetic variation in the HLA region is associated with lateonset sporadic Parkinson's disease. *Nat Genet* 2010; 42: 781–785.
- Sulzer D, Alcalay RN, Garretti F, et al. T cells from patients with Parkinson's disease recognize α-synuclein peptides. *Nature* 2017; 546: 656–661.
- 261. Garretti F, Agalliu D, Arlehamn CSL, et al. Autoimmmunity in parkinson's disease: The role of α:-synucleinspecific T cells. *Front Immunol* 2019; 10: 442306.
- Chervonsky AV. Influence of microbial environment on autoimmunity. *Nat Immunol* 2009; 11: 28–35.
- 263. Garretti F, Monahan C, Sloan N, et al. Interaction of an αsynuclein epitope with HLA-DRB1*15:01 triggers enteric features in mice reminiscent of prodromal Parkinson's disease. *Neuron* 2023; 111: 3397–3413.e5.
- 264. Lindestam Arlehamn CS, Dhanwani R, Pham J, et al. α-Synuclein-specific T cell reactivity is associated with preclinical and early Parkinson's disease. *Nat Commun* 2020; 11: 1875.
- Williams GP, Schonhoff AM, Jurkuvenaite A, et al. CD4 T cells mediate brain inflammation and neurodegeneration in a mouse model of Parkinson's disease. *Brain* 2021; 144: 2047–2059.
- Ascherio A and Munger KL. Environmental risk factors for multiple sclerosis. Part I: The role of infection. Ann Neurol 2007; 61: 288–299.
- 267. Phé V, Pakzad M, Curtis C, et al. Urinary tract infections in multiple sclerosis. *Mult Scler* 2016; 22: 855–861.
- Denes A, Hansen CE, Oezorhan U, et al. Endothelial cells and macrophages as allies in the healthy and diseased brain. *Acta Neuropathol* 2024; 147: 38.
- Dalmau Gasull A, Glavan M, Samawar SKR, et al. The niche matters: origin, function and fate of CNS-associated macrophages during health and disease. *Acta Neuropathol* 2024; 147: 37.
- Masuda T, Amann L, Monaco G, et al. Specification of CNS macrophage subsets occurs postnatally in defined niches. *Nature* 2022; 604: 740–748.
- 271. Wu YC, Bogale TA, Koistinaho J, et al. The contribution of β-amyloid, Tau and α-synuclein to blood–brain barrier damage in neurodegenerative disorders. *Acta Neuropathol* 2024; 147: 39.
- Wen W, Cheng J and Tang Y. Brain perivascular macrophages: current understanding and future prospects. *Brain* 2024; 147: 39–55.

- Prinz M and Priller J. The role of peripheral immune cells in the CNS in steady state and disease. *Nat Neurosci* 2017; 20: 136–144.
- Harms AS, Thome AD, Yan Z, et al. Peripheral monocyte entry is required for alpha-Synuclein induced inflammation and Neurodegeneration in a model of Parkinson disease. *Exp Neurol* 2018; 300: 179–187.
- Kovacs GG, Breydo L, Green R, et al. Intracellular processing of disease-associated α-synuclein in the human brain suggests prion-like cell-to-cell spread. *Neurobiol Dis* 2014; 69: 76–92.
- Frosch M, Amann L and Prinz M. CNS-associated macrophages shape the inflammatory response in a mouse model of Parkinson's disease. *Nat Commun* 2023; 14: 3753.
- Filipowicz AR, McGary CM, Holder GE, et al. Proliferation of perivascular macrophages contributes to the development of encephalitic lesions in HIV-infected humans and in SIV-infected macaques. *Sci Rep* 2016; 6: 32900.
- De Vlaminck K, Van Hove H, Kancheva D, et al. Differential plasticity and fate of brain-resident and recruited macrophages during the onset and resolution of neuroinflammation. *Immunity* 2022; 55: 2085–2102.e9.
- Rebejac J, Eme-Scolan E, Arnaud Paroutaud L, et al. Meningeal macrophages protect against viral neuroinfection. *Immunity* 2022; 55: 2103–2117.e10.
- Johnson ME, Stecher B, Labrie V, et al. Triggers, facilitators, and aggravators: redefining Parkinson's disease pathogenesis. *Trends Neurosci* 2019; 42: 4–13.
- Ortega MA, Fraile-Martinez O, Garcia-Montero C, et al. An updated view of the importance of vesicular trafficking and transport and their role in immune-mediated diseases: potential therapeutic interventions. *Membranes* 2022; 12: 552.
- Prashar A, Schnettger L, Bernard EM, et al. Rab GTPases in immunity and inflammation. *Front Cell Infect Microbiol* 2017; 7: 296153.
- Härtlova A, Herbst S, Peltier J, et al. LRRK2 is a negative regulator of Mycobacterium tuberculosis phagosome maturation in macrophages. *EMBO J* 2018; 37: e98694.
- Zhang Q, Pan Y, Yan R, et al. Commensal bacteria direct selective cargo sorting to promote symbiosis. *Nat Immunol* 2015; 16: 918–926.
- Liu W, Liu X, Li Y, et al. LRRK2 promotes the activation of NLRC4 inflammasome during Salmonella Typhimurium infection. *J Exp Med* 2017; 214: 3051–3066.
- Shutinoski B, Hakimi M, Harmsen IE, et al. Lrrk2 alleles modulate inflammation during microbial infection of mice in a sex-dependent manner. *Sci Transl Med* 2019; 11: eaas9292.
- Simuni T, Chahine LM, Poston K, et al. A biological definition of neuronal α-synuclein disease: towards an integrated staging system for research. *Lancet Neurol* 2024; 23: 178–190.
- Höglinger GU, Adler CH, Berg D, et al. A biological classification of Parkinson's disease: the SynNeurGe research diagnostic criteria. *Lancet Neurol* 2024; 23: 191–204.
- Cabezudo D, Tsafaras G, Van Acker E, et al. Mutant LRRK2 exacerbates immune response and neurodegeneration in a chronic model of experimental colitis. *Acta Neuropathol* 2023; 146: 245–261.
- Matheoud D, Cannon T, Voisin A, et al. Intestinal infection triggers Parkinson's disease-like symptoms in Pink1-/mice. 2019; 571: 565–569.

- 291. Alam MM, Yang D, Li X-Q, et al. Alpha-synuclein (αS) functions as an alarmin to promote inflammatory and immune responses by activating dendritic cells and macrophages. *J Immunol* 2020; 204 (Suppl 1): 160.7.
- 292. Labrie V and Brundin P. Alpha-synuclein to the rescue: immune cell recruitment by alpha-synuclein during gastrointestinal infection. *J Innate Immun* 2017; 9: 437–440.
- 293. Di Maio R, Hoffman EK, Rocha EM, et al. LRRK2 activation in idiopathic Parkinson's disease. *Sci Transl Med* 2018; 10: eaar5429.
- Cookson MR. The role of leucine-rich repeat kinase 2 (LRRK2) in Parkinson's disease. *Nat Rev Neurosci* 2010; 11: 791.
- 295. Heckman MG, Soto-Ortolaza AI, Aasly JO, et al. Population-specific frequencies for LRRK2 susceptibility variants in the Genetic Epidemiology Of Parkinson's Disease (GEO-PD) consortium. *Mov Disord* 2013; 28: 1740.
- Herbst S and Gutierrez MG. LRRK2 in infection: friend or foe? ACS Infect Dis 2019; 5: 809–815.
- Moehle MS, Webber PJ, Tse T, et al. LRRK2 inhibition attenuates microglial inflammatory responses. *J Neurosci* 2012; 32: 1602–1611.
- Miklossy J, Arai T, Guo JP, et al. LRRK2 expression in normal and pathologic human brain and in human cell lines. *J Neuropathol Exp Neurol* 2006; 65: 953–963.
- Gardet A, Benita Y, Li C, et al. LRRK2 is involved in the IFN-γ response and host response to pathogens. *J Immunol* 2010; 185: 5577–5585.
- Alessi DR and Sammler E. LRRK2 kinase in Parkinson's disease. Science 2018; 360: 36–37.
- Yan R and Liu Z. LRRK2 enhances Nod1/2-mediated inflammatory cytokine production by promoting Rip2 phosphorylation. *Protein Cell* 2017; 8: 55–66.
- Ali S, Vollaard AM, Widjaja S, et al. PARK2/PACRG polymorphisms and susceptibility to typhoid and paratyphoid fever. *Clin Exp Immunol* 2006; 144: 425–431.
- Mira MT, Alcaïs A, Van Thuc H, et al. Susceptibility to leprosy is associated with PARK2 and PACRG. *Nature* 2004; 427: 636–640.
- Manzanillo PS, Ayres JS, Watson RO, et al. The ubiquitin ligase parkin mediates resistance to intracellular pathogens. *Nature* 2013; 501: 512–516.
- Bogers JS, Bloem BR and Den Heijer JM. The etiology of Parkinson's disease: new perspectives from gene-environment interactions. *J Parkinsons Dis* 2023; 13: 1281–1288.
- Dorsey ER, De Miranda BR, Horsager J, et al. The body, the brain, the environment, and Parkinson's disease. J Parkinsons Dis 2024; 14: 363.
- 307. Li X, Li C, Zhang W, et al. Inflammation and aging: signaling pathways and intervention therapies. *Signal Transduct Target Ther* 2023; 8: 239.
- Frasier M, Fiske BK and Sherer TB. Precision medicine for Parkinson's disease: The subtyping challenge. *Front Aging Neurosci* 2022; 14: 1064057.

- 309. Sun Y, Koyama Y and Shimada S. Inflammation from peripheral organs to the brain: how does systemic inflammation cause neuroinflammation? *Front Aging Neurosci* 2022; 14: 903455.
- 310. Mori I, Imai Y, Kohsaka S, et al. Upregulated expression of Iba1 molecules in the central nervous system of mice in response to neurovirulent influenza A virus infection. *Microbiol Immunol* 2000; 44: 729–735.
- Li Y, Huang H, Liu B, et al. Inflammasomes as therapeutic targets in human diseases. *Signal Transduct Target Ther* 2021; 6: 247.
- 312. Blevins HM, Xu Y, Biby S, et al. The NLRP3 inflammasome pathway: a review of mechanisms and inhibitors for the treatment of inflammatory diseases. *Front Aging Neurosci* 2022; 14: 879021.
- Deckers J, Anbergen T, Hokke AM, et al. Engineering cytokine therapeutics. *Nat Rev Bioeng* 2023; 1: 286–303.
- Gollan R, Ernst M, Lieker E, et al. Effects of resistance training on motor- and non-motor symptoms in patients with Parkinson's disease: a systematic review and metaanalysis. *J Parkinsons Dis* 2022; 12: 1783–1806.
- Amara AW and Memon AA. Effects of exercise on nonmotor symptoms in Parkinson's disease. *Clin Ther* 2018; 40: 8–15.
- Lister T. Nutrition and lifestyle interventions for managing Parkinson's disease: a narrative review. *J Mov Disord* 2020; 13: 97.
- Knight E, Geetha T, Burnett D, et al. The role of diet and dietary patterns in Parkinson's disease. *Nutrients* 2022; 14: 4472.
- Choi HY, Cho KH, Jin C, et al. Exercise therapies for Parkinson's disease: a systematic review and metaanalysis. *Parkinsons Dis*; 2020; 2020: 2565320.
- Uhrbrand A, Stenager E, Pedersen MS, et al. Parkinson's disease and intensive exercise therapy – a systematic review and meta-analysis of randomized controlled trials. *J Neurol Sci* 2015; 353: 9–19.
- Yun SP, Kam TI, Panicker N, et al. Block of A1 astrocyte conversion by microglia is neuroprotective in models of Parkinson's disease. *Nat Med* 2018; 24: 931–938.
- Brundin L, Bergkvist L and Brundin P. Fire prevention in the Parkinson's disease brain. *Nat Med* 2018; 24: 900–902.
- 322. Athauda D, Maclagan K, Skene SS, et al. Exenatide once weekly versus placebo in Parkinson's disease: a randomised, double-blind, placebo-controlled trial. *Lancet* 2017; 390: 1664.
- Espay AJ, Brundin P and Lang AE. Precision medicine for disease modification in Parkinson disease. *Nat Rev Neurol* 2017; 13: 119–126.
- Massey AR and Beckham JD. Alpha-synuclein, a novel viral restriction factor hiding in plain sight. DNA Cell Biol 2016; 35: 643–645.
- Barrie W, Yang Y, Irving-Pease EK, et al. Elevated genetic risk for multiple sclerosis emerged in steppe pastoralist populations. *Nature* 2024; 625: 321–328.