# Protein kinase R and the metabolic syndrome

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**Abstract.** Metabolic syndrome greatly increases the risk for developing metabolic and cardiovascular disorders and has reached epidemic proportions globally. Despite recent advances in medical science, scientific understandings on the root mechanisms of metabolic syndrome are still not fully understood, and such insufficient knowledge contributes to the relative lack of effective treatments for such diseases. Protein Kinase R (PKR) is a serine threonine kinase activated during various stress conditions. Activation of PKR can increase reactive oxygen product generation, cause oxidative stress and inflammation. In this review we discuss the potential role of PKR in metabolic syndrome, pathways activated by it and the interrelationship between pathways activated, modes of propagation if one of the pathways is inhibited or activated. Specific and effective inhibitors of PKR are being developed and can become potential treatment for metabolic syndrome and prevent many diseases.

Keywords: PKR, metabolic syndrome, inflammation, insulin resistance

## 1. Introduction

Metabolic syndrome is a group of multiple disorders such as high blood pressure, high fasting plasma glucose (hyperglycemia), low HDL cholesterol, high tri-glyceride levels and obesity which ultimately increases the risk of developing cardiovascular disease and Type 2 diabetes. It is an energy utilization disorder and involves the simultaneous presence of 3 of the aforementioned 5 disorders [1]. These conditions invoke stress related response in the body, one of which is Protein Kinase R (PKR). RNA activated/dependent protein kinase (PKR) is a serine threonine kinase that can directly couple to the metabolic pathway due to its catalytic activity and has a role in pathogen recognition [2]. PKR is activated by a number of signals, such as high cholesterol diet [3], pathogens, irradiation, heme limitation [4–10] endoplasmic reticulum (ER) stress and mechanical stress [11]. PKR contains two dsRNA binding domains, one at its N-terminal and the other at its C-terminal [4–5, 12]. It is essential that both the terminals remain active in order to ensure protein Kinase activity. Mutating the lysine-64 end of the domain reduces the binding capacity of RNA domain [13, 14]. In the presence of a pathogen, the double stranded RNA binds to the N-terminal of the PKR enzyme and leads to the phosphorylation of eukaryotic initiation factor

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(eIF2A). It activates eIF2AK2 (eukaryotic translation initiation factor 2-alpha kinase 2) which is coded by the eIF2AK2 gene [2, 15]. This increases phosphorylation of eIF2 and provide the kinase enzyme better access to its substrate [13, 16]. Increase in eIF2 $\alpha$  leads to the inhibition of translation thereby impeding further replication of the virus. In normal state, eIF2 combines with methionyl t-RNA and GTP, which is followed by its combination with the 40 S ribosomal subunit. This complex recognizes the start codon during translation. When the larger subunit is additionally combining with this complex, the GTP-eIF2 complex is hydrolyzed to a GDP complex. During an infection, once eIF2 is phosphorylated to its eIF2 $\alpha$  form, the conversion of GTP-eIF2 complex to GDP is inhibited. This results in blocking translation due to low GDP levels and thus prevents viral replication in cells [17, 18]. Erythrocyte aggregation has also been reported to be associated with insulin resistance, obesity as well as hypertension. Oxidative stress and chronic inflammation are key features of metabolic syndrome. It has been reported earlier that prooxidants and adipocytokines generated in metabolic syndrome alter erythrocyte morphology, increase whole blood viscosity and decrease erythrocyte deformability. The combination of oxidative stress, inflammation and hematological factors have a detrimental effect in metabolic syndrome due to the disturbance in microcirculation [19, 20, 21].

Since the metabolic syndrome is made up of a group of different pathologies, there is no single treatment available as of now. However, therapies that address several of the risk factors concurrently are being researched and provide hope for the future. The starting event or pathology of the metabolic syndrome is not fully understood. In the following sections, we will discuss the involvement of PKR in pathogenesis of obesity, diabetes, insulin resistance and cardiovascular disorders. Since research on PKR and the metabolic syndrome is still in its nascent stage, it is not possible at this stage to tie the various reports into a cohesive mechanism of how PKR is involved in the pathogenesis of metabolic syndrome.

#### 1.1. PKR and obesity

A number of stress and inflammatory responses are observed in metabolic tissues during obesity. As the disease progresses a number of inflammatory and stress responses are evoked in metabolic tissues

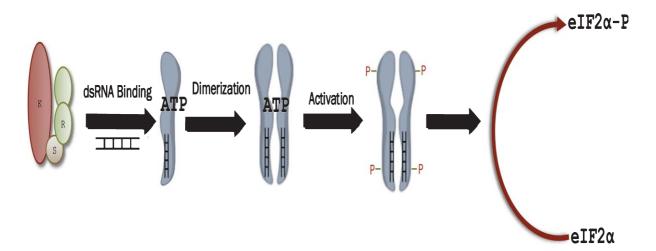


Fig. 1. Mechanism of PKR activation. The dsRNA binding domain (R), the spacer (S), and the protein kinase domain (K) are shown [22].

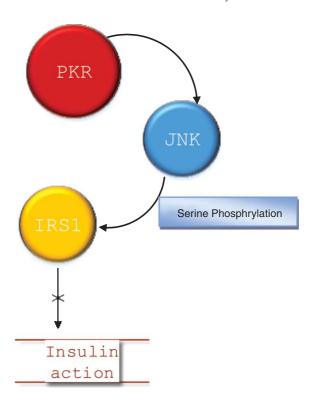


Fig. 2. PKR induced modification of insulin action [22].

leading to chronic inflammation which ultimately leads to the inhibition of insulin receptor signaling and disruption of systemic metabolic homeostasis. Nakamura et al. observed activation of many local low-grade inflammatory and chronic responses which are eventually responsible for the development of insulin resistance in obesity [3]. Both immune and non-immune responses are evoked during this process, referred to as metaflammation [23]. There are only a few receptors that are available which are capable of acting in response to not only metabolic disorders, but also pathogen infections. PKR is one such receptor, which was originally identified as a pathogen sensor but now has also been found to be capable of acting in combination with major inflammatory pathways [2]. PKR is activated by fatty acids, endoplasmic reticulum stress and controls major inflammatory signaling pathways. PKR can act in conjunction with major inflammatory signaling pathways that are involved in metabolic homeostasis, like c-Jun N-teminal kinase (JNK) and IkB kinase (IKK) [24–26]. A marked increase in dietary and genetic obesity is simultaneously accompanied by PKR activation, while its absence helps reduce metabolic deterioration, since excessive nutrients or energy that is available in the body gets utilized. Thus PKR is a critical component of inflammatory complex and it responds to both nutrient and organelle dysfunction [3]. Increased leptin receptor expression, increased PKR activity and increased PKR protein expression was observed in liver and white adipose tissue (WAT) of obese mice as compared to lean control mice [3]. Mice fed high fat diet (HFD) showed higher leptin and PKR activity than lean control mice. However in skeletal muscle of genetic obese (ob/ob) mice, PKR activity showed negligible regulation [3].

PKR is also implicated in regulating molecular integration of nutrient and pathogen sensing pathways in obese mice. Carvalho et al. reported increased PKR activation in liver, muscle, and adipose tissue

of obese humans and after bariatric surgery, reduction in PKR activation accompanied by a decrease in protein kinases like endoplasmic reticulum kinase, c-Jun N-terminal kinase, inhibitor of kappa B kinase, and insulin receptor substrate-1 serine 312 phosphorylation in subcutaneous adipose tissue from these patients [27]. In PKR knockout  $(N-Pkr^{-/-})$  mice, there is an improvement in insulin sensitivity as well as in glucose tolerance and a reduction in fasting blood glucose related to decrease in PP2A phosphatase activity and a parallel increase in insulin-induced Akt phosphorylation, and decrease in glucagon secretion as compared to wild type  $(Pkr^{+/+})$  control mice. In diet induced obese mice, the absence of PKR protects the mice from obesity and insulin resistance by preventing the activation of JNK and IKKβ, thus indicating that PKR is an important modulator of insulin signaling under normal physiological conditions and in obesity [28]. Dietary and genetic obesity causes marked activation of PKR in adipose and liver tissues and absence of PKR alleviates metabolic deterioration due to nutrient or energy excess in mice [3]. Nakamura et al. treated PKR knockout ( $Pkr^{-/-}$ ) and wild type mice ( $Pkr^{+/+}$ ) with *in vivo* lipid infusion and found that upon exposure, only the  $Pkr^{+/+}$  mice showed increased PKR activity, due to PKR activation by metabolic stress and excessive nutrients [3]. Mouse Embryonic Fibroblasts (MEF) on exposure to free fatty acids (FFA) demonstrated the similar results (increased PKR activity) [3]. Besides PKR another enzyme, Toll like receptor 4 (TLR4) (29) is known to show activity in the presence of excess nutrients. In order to determine if TLR4 is involved in PKR activation, TLR4 knockout (TLR4<sup>-/-</sup>) mice were exposed to high fat diet, however no significant difference in PKR activity was observed between (TLR4<sup>-/-</sup>) and control (TLR4+<sup>/+</sup>) mice thus indicating TLR4 is not involved in PKR activation [3].

In obese men and women adipose tissue and liver showed an increased activation of c-jun N-terminal kinase (JNK), the inhibitor of k kinase (IKK), and PKR in comparison to lean control non obese group. The inflammasome and the Toll-like receptors (TLRs) of the innate immune system are also activated. Inflammatory signals and excess nutrients may activate the TLRs pathways and ultimately JNK, IKK, and PKR. These kinases will further regulate downstream transcriptional programs through the transcription factors activator protein-1 (AP-1), NF-κB, and interferon regulatory factor (IRF), inducing upregulation of inflammatory mediator gene expression. The increase in cytokines aggravates receptor activation by establishing a positive feedback loop of inflammation and the inhibitory signaling of metabolic pathways [29].

## 1.2. PKR and diabetes

Metabolic syndrome is associated with elevated blood glucose levels, which in turn will affect plasma insulin levels. It has been reported earlier that in PKR knockout ( $Pkr^{-/-}$ ) mice, fasting plasma glucose is reduced while insulin action and insulin-induced Akt phosphorylation is improved as compared to wild type control ( $Pkr^{+/+}$ ) mice. PKR is known to phosphorylate the regulatory subunit of PP2A, which then activates the catalytic subunit of PP2A inducing its phosphatase activity [25]. Mice islet  $\beta$ -cells and insulinoma cell lines exposed to high glucose and proinflammatory cytokines showed significantly increased PKR activity associated with significantly inhibited cell proliferation by arresting cell cycle at G1 phase. PKR activation abolished the pro-proliferative effects of IGF-I by activating JNK and disrupting IRS1/PI3K/Akt signaling pathway [30].

Inhibition of PKR reduces stress-induced JNK activation and IRS1 serine phosphorylation *in vitro* and *in vivo* [31]. PKR is known to directly target and modify the insulin receptor and thus inhibiting insulin action. It has been reported earlier that PKR induces the inhibitory phosphorylation of IRS at site Ser312 and activates the transcription factor, Foxo1, which in turn up-regulates the protein expression level of

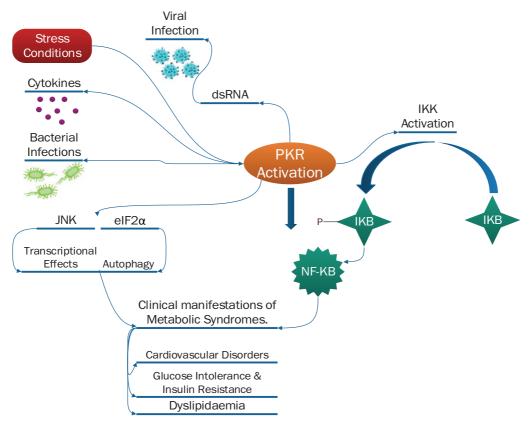


Fig. 3. Different pathways of PKR activation [3].

IRS2 [31]. Knockout of PKR ( $Pkr^{-/-}$ ) in mice showed protection against insulin resistance and diabetes [3]. Thus pharmacologically targeting PKR may be an effective therapeutic strategy for the treatment of type 2 diabetes. Under stress condition, JNK negatively controls insulin signaling through serine phosphorylation of IRS1 instead of the normal tyrosine phosphorylation [32]. JNK activation by PKR may also lead to serine phosphorylation of IRS1. Nakamura et al. tested this theory by taking Pkr knockout ( $Pkr^{-/-}$ ) and wild type ( $Pkr^{+/+}$ ) MEFs and exposing them to palmitic acid and thapsigargin. In Wild type ( $Pkr^{+/+}$ ) MEFs phosphorylation of IRS1 was observed whereas in PKR knockout ( $Pkr^{-/-}$ ) MEF's, no IRS1 phosphorylation was observed. This proves that PKR is involved in the eventual phosphorylation of IRS1 [3]. Garcia et al. reported that treating wild type ( $Pkr^{+/+}$ ) and PKR knockout ( $Pkr^{-/-}$ ) MEFs with polyinosinic-polycytidylic (PolyI.C), a direct activator of PKR, IRS1 phosphorylation was only observed in in wild type ( $Pkr^{+/+}$ ) MEFs [33].

Einarson et al. confirmed the interaction between PKR and IRS1 by Pull down assays, when a direct interaction was observed between IRS1 and PKR [34]. Nakamura et al. reported that PKR causes the direct phosphorylation of the serine 307 residue of IRS1, using TNF- $\alpha$  (known to activate PKR) and TG on both wild type (Pkr<sup>+/+</sup>) and PKR knockout (Pkr<sup>-/-</sup>) MEFs and demonstrated the extent of IRS1 phosphorylation using a phospho specific antibody. It was found that the WT MEFs were able to show the excessive phosphorylation, unlike their counterparts [3].

PKR plays an important role in insulin resistance as well. Nakamura et al. exposed wild type  $(Pkr^{+/+})$  and PKR knockout  $(Pkr^{-/-})$  mice to high fat diet and observe an increase in insulin induced Akt phos-

| Factor                 | Pkr <sup>+/+</sup> (HFD) | Pkr <sup>-/-</sup> (HFD) |  |
|------------------------|--------------------------|--------------------------|--|
| Insulin resistance     | Present                  | Absent                   |  |
| Obesity                | Present                  | Absent                   |  |
| Leptin levels          | High                     | Low                      |  |
| Adinopectin levels     | No significant change    | No significant change    |  |
| Glucose levels         | High                     | Low                      |  |
| Adipose tissue content | High                     | Low                      |  |

Table 1 Effect of PKR knockout on different metabolic parameters [3]

phorylation (Serine 473) in liver and adipose tissue of PKR knockout ( $Pkr^{-/-}$ ) mice as compared to wild type ( $Pkr^{+/+}$ ) control mice (3).

# 1.3. PKR and cardiovascular disorders

Young et al. reported the significance of endoplasmic reticulum (ER) stress as signaling event for angiotensin II-induced hypertension in cells of central nervous system [35]. PKR is a  $2\alpha$  kinase initiation factor and is known to inhibit translation of mRNA under stress condition [19]. It also initiates signalling of apoptosis and inflammation, independent of translational regulation. Congestive heart failure (CHF) is associated with inflammation, cardiomyocyte hypertrophy, and apoptosis [36]. Various factors have been reported to contribute to development of CHF like oxidative stress [36], chronic inflammation, and Toll receptor activation [37]. The chronic inflammations also play role in defence against viral myocarditis [38]. From reported literature, it is evident that PKR is not only an anti-viral factor activated by interferons but also induced or activated in various forms of stress [39–43] and PKR activation due to viral infection may be beneficial in inhibiting viral replication and infections by repression of inflammatory signalling and translation. However, activated PKR induces cellular stress in the heart leading to significant increase in inflammation and apoptosis ultimately leading to chronic pathological conditions such as congestive heart failure (CHF) [33].

Wang et al. and group reported PKR expression in human suffering from CHF. There was significant increase in myocardial expression and translocation of PKR in human patients and mice suffering from CHF [33]. They utilized left ventricular (LV) samples from a human CHF patient, and PKR knockout mice to investigate role of PKR. PKR has significant role in development of CHF by intensifying apoptosis and inflammation of cardiomyocytes by inducing chronic transverse aortic constriction (TAC). On the basis of their research it is evident that PKR inhibition can be utilized as a therapeutic target to treat CHF, as deletion or blocking PKR protects heart from systolic-overload-induced congestive heart failure [33]. B. Tian et al. studied myotonic dystrophy CTG repeat in 3' untranslated region of protein kinase (DMPK). They hypothesized the mechanism for myotonic dystrophy to be due to increased affinity of CUG repeats towards PKR, which was proved by activation of PKR *in vitro*. It was concluded that dsRNA binding is responsible for nuclear retention or toxicity due expanded CUG repeats. This repeats occur much more extended almost 30 times in heart and brain [44]. Early myotonic dystrophy leads to heart block, muscle wasting, and neuropsychiatric impairment [45]. It is evident blocking of this interaction CUG repeats and PKR in heart muscle or skeletal muscle can prevent myotonic dystrophy.

Bleiblo et al. has shown that natural RNA derived from bacteria binds to and activates PKR and this bacterial RNA induces human cardiac myocyte apoptosis in a PKR-dependent manner [46].

## 1.4. Therapeutic potential of PKR inhibitors in components of the metabolic syndrome

The main limitation we have in this area is that the role of PKR in various aspects of the metabolic syndrome is still in the initial stages of investigation and more and more reports are coming out. Even then, these reports deal with the conditions of hypertension, obesity and diabetes as single entities and describe limited findings on the effects of PKR change in cultured cells and animal models. Apparently, it will take some time before an integrated picture of the role of PKR in the metabolic syndrome starts emerging. As such, the initiating events in the pathogenesis of the metabolic syndrome are also still far from clear.

## 2. Conclusion and future prospects

The present review indicates that PKR plays a crucial role in the many complications of metabolic disorders. Many important questions still remain to be addressed. Current understanding on the inflammatory mechanisms of metabolic syndrome and related disorders are still in its primitive stage. We anticipate in near future eventually these findings will be translated into novel and effective treatments/preventions against metabolic and related diseases.

## References

- [1] Kaur J. A comprehensive review on metabolic syndrome, Cardiol Res Pract 2014;2014:943162.
- [2] Samuel C. The eIF-2 alpha protein kinases, regulators of translation in eukaryotes from yeasts to humans. J Biol Chem 1993;268:7603-6.
- [3] Nakamura et al. Double-stranded RNA-dependent protein kinase links pathogen sensing with stress and metabolic homeostasis. Cell 2010;140:338-48.
- [4] Clemens MJ. 5 Protein kinases that phosphorylate eIF2 and eIF2B, and their role in eukaryotic cell translational control. Cold Spring Harbor Monograph Archive 1996;30:139-72.
- [5] Proud CG. PKR: A new name and new roles. Trends Biochem Sci 1995;20:241-6.
- [6] Kaufman RJ. Stress signaling from the lumen of the endoplasmic reticulum: Coordination of gene transcriptional and translational controls. Genes Dev 1999;13:1211-33.
- [7] Brostrom CO, Brostrom MA. Regulation of translational initiation during cellular responses to stress. Prog Nucleic Acid Res Mol Biol 1998:58:79-125.
- [8] Gale M, Katze MG. Molecular mechanisms of interferon resistance mediated by viral-directed inhibition of PKR, the interferon-induced protein kinase. Pharmacol Ther 1998;78:29-46.
- [9] Wek RC. elF-2 kinases: Regulators of general and gene-specific translation initiation. Trends Biochem Sci 1994;19:491-6.
- [10] Hinnebusch AG. Translational regulation of yeast GCN4 A window on factors that control initiator-tRNA binding to the ribosome. J Biol Chem 1997;272:21661-4.
- [11] Ron D, Walter P. Signal integration in the endoplasmic reticulum unfolded protein response. Nat Rev Mol Cell Biol 2007;8:519-29.
- [12] Williams B. PKR; a sentinel kinase for cellular stress. Oncogene 1991;18:6112-20.
- [13] WuS, Kumar KU, Kaufman RJ. Identification and requirement of three ribosome binding domains in dsRNA-dependent protein kinase (PKR). Biochemistry 1998;37:13816-26.
- [14] McCormack SJ, Ortega LG, Doohan JP, Samuel CE. Mechanism of interferon action motif I of the interferon-induced, RNA-dependent protein kinase (PKR) is sufficient to mediate RNA-binding activity. Virology 1994;198:92-9.

- [15] Feng G, Chong K, Kumar A, Williams B. Identification of double-stranded RNA-binding domains in the interferon-induced double-stranded RNA-activated p68 kinase. Proc Natl Acad Sci 1992;89: 5447-51.
- [16] Zhu S, Romano PR, Wek RC. Ribosome targeting of PKR is mediated by two double-stranded RNA-binding domains and facilitates *in vivo* phosphorylation of eukaryotic initiation factor-2. J Biol Chem 1997;272:14434-41.
- [17] Hershey JW, Merrick WC. Pathway and mechanism of initiation of protein synthesis, cold spring harbor monograph series. 2000;39:33-88.
- [18] Kaufman RJ. Double-stranded RNA-activated protein kinase mediates virus-induced apoptosis: A new role for an old actor. Proc Natl Acad Sci 1999;96:11693-95.
- [19] Gyawali P, Richards RS, Hughes DL, Tinley P. Erythrocyte aggregation and metabolic syndrome. Clin Hemorheol Microcirc 2014;57(1):73-83.
- [20] Wiewiora et al. Association between hemorheological alteration and clinical diagnosis of metabolic syndrome among patients qualified for bariatric surgery. Clin Hemorheol Microcirc 2014;56(2):101-9.
- [21] Ucak S, Basat O, Cetinkaya E. Plasma viscosity in patients with metabolic syndrome. Clin Hemorheol Microcirc 2013;54(1):15-22.
- [22] Tavantzis SM. dsRNA genetic elements: Concepts and applications in agriculture, forestry, and medicine, CRC Press, 2001.
- [23] Hotamisligil GS. Inflammation and metabolic disorders. Nature 2006;444:860-67.
- [24] Bonnet MC, Weil R, Dam E, Hovanessian AG, Meurs EF. PKR stimulates NF-kappaB irrespective of its kinase function by interacting with the IkappaB kinase complex. Mol Cell Biol 2000;20:4532-42.
- [25] Goh KC, DeVeer MJ, Williams BR. The protein kinase PKR is required for p38 MAPK activation and the innate immune response to bacteria endotoxin. EMBO J 2000;19:4292-97.
- [26] Takada Y, Ichikawa H, Pataer A, Swisher S, Aggarwal BB. Genetic deletion of PKR abrogates TNF-induced activation of IkappaBalpha kinase, JNK, Akt and cell proliferation but potentiates p44/p42 MAPK and p38 MAPK activation. Oncogene 2007;26:1201-12.
- [27] Carvalho et al. Modulation of double stranded RNA activated protein kinase in insulin sensitive tissues of obese humans. Obesity 2013;21:2452-7.
- [28] Carvalho et al. Double-stranded RNA-activated protein kinase is a key modulator of insulin sensitivity in physiological conditions and in obesity in mice. Endocrinology 2012;153:5261-74.
- [29] Emanuela et al. Inflammation as a link between obesity and metabolic syndrome. J Nutr Metab 2012;2012:476380.
- [30] Chen SS. Activation of double-stranded RNA-dependent protein kinase inhibits proliferation of pancreatic β-cells. Biochem Biophys Res Commun 2014 17;443(3):814-20.
- [31] Yang X, Nath A, Opperman MJ, Chan C. The Double-stranded RNA–dependent Protein Kinase Differentially Regulates Insulin Receptor Substrates 1 and 2 in HepG2 Cells. Mol Biol Cell 2010;21(19):3449-58.
- [32] Dearth et al. Mammary tumorigenesis and metastasis caused by overexpression of insulin receptor substrate 1 (IRS-1) or IRS-2. Mol Cell Biol 2006;26:9302-14.
- [33] Garcia et al. Impact of protein kinase PKR in cell biology: From antiviral to antiproliferative action. Microbiol Mol Biol Rev 2006;70:1032-60.
- [34] Einarson MB, Pugacheva EN, Orlinick JR. Identification of protein-protein interactions with glutathione-S-transferase (GST) fusion proteins. Cold Spring Harbor Protocols. 2007.
- [35] Young et al. Davisson, ER stress in the brain subfornical organ mediates angiotensin-dependent hypertension. J Clin Invest 2012; 122:3960-64.
- [36] Wang et al. Double stranded RNA-dependent protein kinase deficiency protects the heart from systolic overload-induced congestive heart failure, Circulation 2014;129(13):1397-406.
- [37] Kadokami et al. Anti–Tumor necrosis factor-α antibody limits heart failure in a transgenic model, Circulation 2001:104:1094-97.
- [38] Stewart MJ, Blum MA, Sherry B. PKR's protective role in viral myocarditis. Virology 2003;314:92-100.
- [39] Stark GR, Kerr IM, Williams BR, Silverman RH, Schreiber RD. How cells respond to interferons. Annu Rev Biochem 1998;67:227-64.
- [40] Pitini V, Arrigo C, Altavilla G. How cells respond to interferons. J clin Oncol 2010;28:e439-e439.
- [41] Li G, Scull C, Ozcan LTabas I. NADPH oxidase links endoplasmic reticulum stress, oxidative stress, and PKR activation to induce apoptosis. J cell Biol 2010;191:1113-25.
- [42] Wang et al. Mechanical force activates eIF- $2\alpha$  phospho-kinases in fibroblast. Biochem Biophys Res Commun 2005;330:123-30.

- [43] Asakura Y, Fujiwara Y, Kato N, Sato Y, Komatsu T. Serine/threonine kinase PKR: A sentinel kinase that discriminates a signaling pathway mediated by TLR4 from those mediated by TLR3 and TLR9. Am J Hemato 2007;82:640-42.
- [44] Tian et al. Expanded CUG repeat RNAs form hairpins that activate the double-stranded RNA-dependent protein kinase PKR. RNA 2000;6:79-87.
- [45] Harper P. Myotonic dystrophy. Oxford University Press, 2009.
- [46] Bleiblo et al. Bacterial RNA induces myocyte cellular dysfunction through the activation of PKR. J Thorac Dis 2012;4(2):114-25.