

Comparing Tau PET Visual Interpretation with Tau PET Quantification, Cerebrospinal Fluid Biomarkers, and Longitudinal Clinical Assessment

Charles D. Chen^a, Maria Rosana Ponisio^a, Jordan A. Lang^a, Shaney Flores^a, Suzanne E. Schindler^b, Anne M. Fagan^b, John C. Morris^b and Tammie L.S. Benzinger^{a,*}

^a*Mallinckrodt Institute of Radiology, Washington University in St. Louis, St. Louis, MO, USA*

^b*Department of Neurology, Washington University in St. Louis, St. Louis, MO, USA*

Accepted 14 March 2023

Pre-press 17 April 2023

Abstract.

Background: ¹⁸F-flortaucipir PET received FDA approval to visualize aggregated neurofibrillary tangles (NFTs) in brains of adult patients with cognitive impairment being evaluated for Alzheimer's disease (AD). However, manufacturer's guidelines for visual interpretation of ¹⁸F-flortaucipir PET differ from how ¹⁸F-flortaucipir PET has been measured in research settings using standardized uptake value ratios (SUVRs). How visual interpretation relates to ¹⁸F-flortaucipir PET SUVR, cerebrospinal fluid (CSF) biomarkers, or longitudinal clinical assessment is not well understood.

Objective: We compare various diagnostic methods in participants enrolled in longitudinal observational studies of aging and memory ($n = 189$, 23 were cognitively impaired).

Methods: Participants had tau PET, A β PET, MRI, and clinical and cognitive evaluation within 18 months ($n = 189$); the majority ($n = 144$) also underwent lumbar puncture. Two radiologists followed manufacturer's guidelines for ¹⁸F-flortaucipir PET visual interpretation.

Results: Visual interpretation had high agreement with SUVR (98.4%) and moderate agreement with CSF p-tau181 (86.1%). Two participants demonstrated ¹⁸F-flortaucipir uptake from meningiomas. Visual interpretation could not predict follow-up clinical assessment in 9.52% of cases.

Conclusion: Visual interpretation was highly consistent with SUVR (discordant participants had hemorrhagic infarcts or occipital-predominant AD NFT deposition) and moderately consistent with CSF p-tau181 (discordant participants had AD pathophysiology not detectable on tau PET). However, close association between AD NFT deposition and clinical onset in group-level studies does not necessarily hold at the individual level, with discrepancies arising from atypical AD, vascular dementia, or frontotemporal dementia. A better understanding of relationships across imaging, CSF biomarkers, and clinical assessment is needed to provide appropriate diagnoses for these individuals.

Keywords: Alzheimer's disease, cerebrospinal fluid, positron emission tomography, tauopathies

INTRODUCTION

The pathological hallmarks of Alzheimer's disease (AD) are amyloid- β (A β) plaques and misfolded hyperphosphorylated tau neurofibrillary tangles (NFTs) [1, 2]. *In vivo* evaluation of aggregated

*Correspondence to: Tammie L.S. Benzinger, Washington University School of Medicine, 660 South Euclid, Campus Box 8225, St. Louis, MO 63110, USA. Tel.: +1 314 362 1558; Fax: +1 314 362 5297; E-mail: benzinger@wustl.edu.

tau or associated pathophysiology in AD was first performed using immunoassays for cerebrospinal fluid (CSF) tau phosphorylated at position 181 (p-tau) [3]. Later, tau PET radiotracers were developed [4–6], along with methods for tau PET standardized uptake value ratio (SUVR) analyses [7, 8]. The first generation of tau PET radiotracers includes the arylquinoline derivatives ^{18}F -THK5317 and ^{18}F -THK5351, the pyrido-indole derivative ^{18}F -flortaucipir, and the phenyl/pyridinyl-butadienyl-benzothiazole/benzothiazolium derivative ^{11}C -PBB3. Among these, ^{18}F -flortaucipir (TauvidTM, Avid Radiopharmaceuticals) became the first to be approved by the United States Food and Drug Administration to estimate the density and distribution of aggregated tau NFTs in adult patients with cognitive impairment being evaluated for AD. Following the manufacturer's guidelines for performing a visual interpretation of ^{18}F -flortaucipir PET imaging involves identifying the presence or absence of contiguous radiotracer uptake greater than 1.65 times the cerebellar uptake in either the posterolateral temporal, occipital, or parietal/precuneus regions. This method differs greatly from most research procedures for automated quantification of tau PET imaging data, such as taking the volume-weighted mean standardized uptake value ratio (SUVR) in a temporal meta-region of interest (ROI) and comparing that to a cohort-defined threshold [7, 8]. These methodological differences may lead to disagreements between visual interpretation and SUVR quantification. In particular, the temporal meta-ROI often used in SUVR quantification does not contain any of the occipital or parietal/precuneus structures used in visual interpretation, and includes several medial temporal lobe structures ignored in visual interpretation. Additionally, in the clinic ^{18}F -flortaucipir PET imaging is only indicated for use in adult patients with cognitive impairment who are being evaluated for AD, whereas in a research setting ^{18}F -flortaucipir PET imaging is performed regardless of cognitive status, calling into question whether ^{18}F -flortaucipir PET imaging is a reliable measure of NFT deposition during preclinical stages of AD. Tau pathophysiology can also be evaluated by measuring phosphorylated tau concentrations in the CSF, and several studies have provided additional evidence that tau PET is more strongly coupled to cognitive decline, whereas CSF p-tau181 is more tightly linked to preclinical AD [9–12]. Understanding where these three methods—tau PET visual interpretation, tau PET SUVR quantification,

and CSF p-tau181 concentration—agree and differ may improve how we define AD NFT deposition and AD clinical diagnoses.

MATERIALS AND METHODS

Study participants

Participants selected for this study were enrolled in longitudinal observational studies of aging and memory at the Charles F. and Joanne Knight Alzheimer Disease Research Center (Knight ADRC, $n = 189$, of whom 23 were cognitively impaired, Table 1). All participants met the inclusion criteria of having a tau PET usable for visual reads, and an A β PET, MRI, and clinical and cognitive evaluation, all within 18 months; the majority of participants ($n = 144$) also underwent lumbar puncture within 18 months of their tau PET scan. The study was approved by the Washington University in St. Louis Human Research Protection Office and Institutional Review Board, and all participants or their designees signed an informed consent form.

Clinical and cognitive assessment

Participants were assessed clinically and cognitively using the neuropsychological test battery from the Uniform Data Set (UDS) [13], which includes the Clinical Dementia Rating (CDR[®]) [14] and the Mini-Mental State Examination (MMSE) [15]. The CDR assesses three domains of cognition (memory, orientation, judgment, and problem solving) and three domains of function (community affairs, home and hobbies, personal care): scores from the six domains can either be summed to yield the CDR Sum of Boxes score, or passed to a lookup table to yield the CDR Global score.

Tau PET acquisition

Participants were scanned on a Siemens Biograph 40 TruePoint (Siemens Healthineers). Participants received a single intravenous bolus injection (341 ± 29.8 MBq) of ^{18}F -flortaucipir (TauvidTM, Avid Radiopharmaceuticals). Emission data were collected 80–100 min post injection. List-mode data were reconstructed using ordered subset expectation maximization with three iterations and 21 subsets. A low-dose CT scan preceded PET acquisition for attenuation correction.

Table 1
Participant characteristics

		Cognitively normal	Cognitively impaired	Total
Number		166	23	189
Mean age in years (SD)		68.9 ± 8.34	75.7 ± 7.36	69.8 ± 8.51
Female (%)		93 (56.0)	12 (52.2)	105 (55.6)
Race	White	147	23	170
	Black or African American	18	0	18
	Asian	1	0	1
Mean MMSE (SD)		29.2 (1.12)	26.0 (3.66)	28.8 (1.94)
CDR [®]	0	166	0	166
	0.5	0	16	16
	1	0	6	6
	2	0	1	1
Clinical diagnosis	Cognitively normal	166	0	166
	Uncertain dementia	0	9	9
	0.5 in memory only	0	1	1
	AD dementia	0	13	13
APOE genotype	2/2	1	0	1
	2/3	27	1	28
	2/4	6	1	7
	3/3	83	8	91
	3/4	42	11	53
	4/4	6	2	8
	Unknown	1	0	1
Tau PET temporal meta-ROI SUVR	Mean ± SD	1.15 ± 0.106	1.44 ± 0.364	1.18 ± 0.185
	[min, max]	[0.924, 1.882]	[1.024, 2.43]	[0.924, 2.43]
	Positive (%)	4 (2.41)	13 (56.5)	17 (8.99)
Tau PET visual interpretation	Positive (%)	6 (3.61)	14 (60.9)	20 (10.6)
Aβ PET (Centiloid)	Mean ± SD	19.9 ± 34.4	74.3 ± 45.6	26.5 ± 40.0
	Positive (%)	45 (27.1)	19 (82.6)	64 (33.9)

CDR[®], Clinical Dementia Rating[®]; MMSE, Mini-Mental State Exam; SD, standard deviation.

Tau PET SUVR

Reconstructed PET images were processed using the PET Unified Pipeline (<https://github.com/ysu001/PUP>) and coregistered to corresponding MR images [16, 17]. After segmenting MR images into ROIs using FreeSurfer version 5.3 [18], regional SUVRs were defined from the reconstructed PET images using a cerebellar gray reference region. The temporal meta-ROI SUVR was defined as the volume-weighted mean SUVR of the amygdala, entorhinal cortex, fusiform, parahippocampal, inferior temporal, and middle temporal ROIs [7, 8].

Tau PET visual interpretation

Two radiologists with training in nuclear medicine (J.A.L. and M.R.P.) followed the manufacturer's guidelines for ¹⁸F-flortaucipir PET visual interpretation of participant scans using MIM Encore (MIM Software). Reconstructed PET images were coregistered with corresponding MR images. A ROI was drawn around the whole cerebellum in the axial plane

that maximizes its cross-sectional area. A color scale with a rapid transition at 1.65 times the mean cerebellar counts was defined. The temporal lobe was divided into the anterolateral, anterior mesial, posterolateral, and posterior mesial temporal quadrants by placing the horizontal crosshair posterior to the brainstem nuclei, and the vertical crosshair at the widest portion of the temporal pole. An image was considered positive if it showed contiguous activity above the rapid transition/cutoff in the cortical gray matter of the posterolateral temporal, occipital, or parietal/precuneus regions. An image was considered negative if it showed no activity above the cutoff in the cortical gray matter of the posterolateral temporal, occipital, or parietal/precuneus regions, or if it showed activity above the cutoff in the cortical gray matter restricted to the medial temporal, anterolateral temporal, and frontal regions. Off-target binding, which may be seen in the choroid plexus, striatum, and brainstem nuclei, and small foci of noncontiguous activity, which may be seen throughout the cortical gray matter, were not used when determining tau positivity. Radiologists were blinded

to all other information about each participant. In addition to following the manufacturer's guidelines for ^{18}F -flortaucipir PET visual interpretation, in this study, radiologists also reported whether radiotracer activity was symmetric across left and right hemispheres and whether there was off-target binding in the choroid plexus, striatum, brainstem nuclei, or bone/meninges. Notable findings (such as incidental meningiomas) were also reported. Both radiologists determined off-target binding and incidental findings using MR imaging. Additionally, incidental findings were confirmed with a neuroradiologist (T.L.S.B.).

A β PET

Participants were scanned on either a Siemens Biograph 40 TruePoint, Biograph mMR, or Biograph Vision 600 (Siemens Healthineers). Participants received either a single intravenous bolus injection (539 ± 159 MBq) of ^{11}C -Pittsburgh compound B (PiB) or (369 ± 22.4 MBq) of ^{18}F -florbetapir (AmyvidTM, Avid Radiopharmaceuticals). Emission data were either collected 30–60 min post injection (^{11}C -PiB) or 50–70 min post injection (^{18}F -florbetapir). Reconstructed PET images were formed and pre-processed in the same manner as tau PET. An A β PET SUVR was defined for each radiotracer [16, 17] and standardized to the Centiloid scale [19, 20].

MR acquisition

Participants were scanned on either a Siemens Biograph mMR or Magnetom Vida (Siemens Healthineers). Across all scanners, T1-weighted head MR images were acquired using a magnetization prepared rapid gradient echo (MPRAGE) generalized auto-calibrating partial parallel acquisition (GRAPPA) sequence using a repetition time = 2300 ms, echo time = 2.95 ms, flip angle = 9° , at $1.1 \times 1.1 \times 1.2$ mm³ voxel resolution.

CSF

CSF was collected under standardized operating procedures. Participants underwent lumbar puncture in the morning following overnight fasting and 20–30 ml of CSF was collected in a 50 ml polypropylene tube via gravity drip using an atraumatic Sprotte 22-gauge spinal needle. CSF samples were kept on ice and centrifuged at low speed within 2 h of collection, then transferred to another 50 ml tube to remove cells. CSF was aliquoted at 500 μl into polypropylene tubes

and stored at -80°C . Concentrations of CSF p-tau181, A β_{42} , and A β_{40} were measured by chemiluminescent enzyme immunoassay using a fully automated platform (LUMIPULSE G1200, Fujirebio) according to the manufacturer's specifications.

Statistical analyses

Cutoffs for binarizing tau PET, A β PET, CSF p-tau181, and CSF A β_{42} /A β_{40} values were determined by fitting a two-component univariate Manly mixture model [21] in R software [22] to all relevant baseline PET SUVR or CSF measurements available in the Knight ADRC Data Freeze 17 (Supplementary Table 1) and finding the decision boundary. Manly mixture modeling was used to account for possible severe skewness in the data that would be difficult to model using Gaussian mixture modeling, and to account for skewness that can vary from component to component, which would be impossible to model using log or Box-Cox transformations [21]. Cohen's kappa (κ) was used to measure inter-rater reliability between the two radiologists' tau PET visual interpretations, as well as between tau PET visual interpretation and tau PET SUVR quantification, and between tau PET visual interpretation and CSF p-tau181 concentration.

RESULTS

Study participants

Overall, participants were on average (\pm standard deviation) 69.8 ± 8.51 years old, most were cognitively normal with a global Clinical Dementia Rating (CDR[®]) [14] of 0 ($n = 166/189$, 87.8%) and most did not carry the APOE $\epsilon 4$ allele ($n = 120/188$, 63.8%) (Table 1). Cognitively normal participants had a mean tau PET temporal meta-ROI SUVR of 1.15 ± 0.106 and a mean Centiloid of 19.9 ± 34.4 . Cognitively impaired participants ($n = 23/189$, 12.2%) had a clinical diagnosis of either uncertain dementia ($n = 9$), a CDR = 0.5 in memory only ($n = 1$), or AD dementia ($n = 13$). They also had a mean tau PET temporal meta-ROI SUVR of 1.44 ± 0.364 and a mean Centiloid of 74.3 ± 45.6 .

The following quantitative cutoffs were identified through statistical modeling and are used to determine biomarker positivity in the remainder of the analyses: tau PET temporal meta-ROI SUVR cutoff = 1.32, A β PET (Centiloid) cutoff = 21.6, CSF

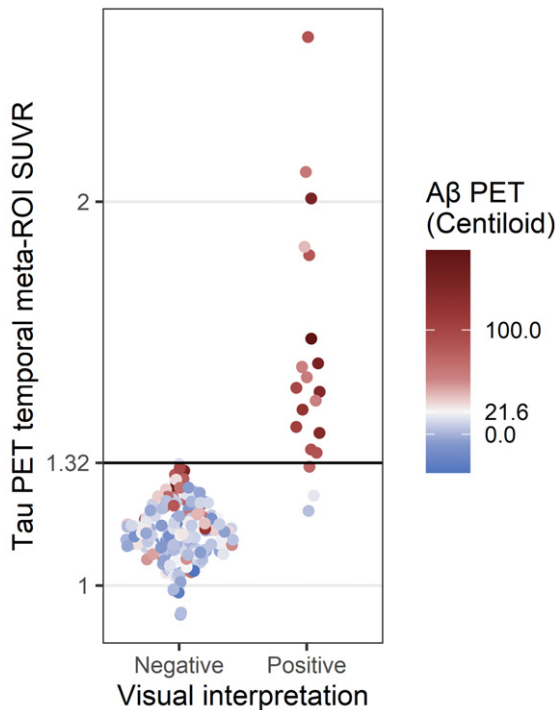


Fig. 1. Comparison of tau PET visual interpretation with tau PET SUVR. Each PET study was assessed by visual interpretation using the manufacturer's guidelines to determine positivity (x-axis) and by temporal meta-ROI SUVR analysis using a cutoff of SUVR = 1.32 to determine positivity (y-axis). The color indicates the A β PET status for each case (positive A β PET, red; negative A β PET, blue; cutoff = 21.6 Centiloids).

p-tau181 cutoff = 58.1 pg/ml, and CSF A β_{42} /A β_{40} cutoff = 0.0737.

Tau PET visual interpretation and tau PET SUVR

Of the 189 ^{18}F -florataucipir PET images, 20 (10.6%) were read as positive by both radiologists. Both radiologists also read 169 images as negative and thus agreed on the overall visual interpretation of each image in the current study ($n = 189/189$, 100%, $\kappa = 1$). Agreement between visual interpretation and SUVR quantification was high ($n = 186/189$, 98.4%, $\kappa = 0.910$) (Fig. 1).

The three participants who had discordant results between visual interpretation and SUVR quantification all had tau-positive visual interpretations and tau-negative SUVRs. One participant (Fig. 2a) demonstrated elevated ^{18}F -florataucipir uptake in the right precuneus and was A β PET, CSF A β_{42} /A β_{40} , and CSF p-tau181 negative (Table 2). Additional MR imaging revealed a hypointensity on T2*-weighted MRI that colocalized with the elevated right pre-

cuneus radiotracer uptake on ^{18}F -florataucipir PET, suggesting a hemorrhagic infarct to be the cause of elevated radiotracer uptake instead of AD NFT deposition (Fig. 2b). Upon review of the additional T2*-weighted MR imaging, the readers also revised their interpretation of the image to be tau negative.

The other two participants (Fig. 2c, d) demonstrated lateralized occipital uptake, with greater uptake in either the left (Fig. 2c) or right (Fig. 2d), and were A β PET, CSF A β_{42} /A β_{40} , and CSF p-tau181 positive (Table 2). The participant with greater left occipital uptake than right, likely has an occipital-predominant form of AD tau pathology [23].

The participant with greater right occipital uptake than left also had posterolateral temporal and parietal/precuneus uptake. The temporal meta-ROI SUVR was borderline negative, suggesting that, perhaps due to the lateralized uptake, the SUVR was artificially low for this case.

Incidental findings

In terms of incidental findings, frontal meningiomas were identified in two participants. One participant had a meningioma in their left posterior frontal lobe (Fig. 3a, b); the other participant had it in their left frontal lobe (Fig. 3c, d). Both meningiomas had elevated levels of radiotracer uptake. The first participant also had elevated right posterolateral temporal uptake and tau-positive visual interpretation and SUVR and was A β PET positive (Table 3). The other participant had tau-negative visual interpretation and SUVR and was A β PET negative.

CSF p-tau181

Agreement between visual interpretation and CSF p-tau181 was moderate ($n = 124/144$, 86.1%, $\kappa = 0.526$, Table 4). Two participants had tau-positive visual interpretations but were CSF p-tau181 negative (Fig. 4a, b). One participant was previously identified as having a tau-positive visual interpretation but tau-negative SUVR (the same case as in Fig. 2a, b). The other participant demonstrated posterolateral temporal uptake in both hemispheres and was A β PET and CSF A β_{42} /A β_{40} positive. In addition, 18 participants had tau-negative visual interpretations but were CSF p-tau181 positive (Fig. 4a, b). These cases were mostly A β PET positive ($n = 14/18$, 77.8%) and/or CSF A β_{42} /A β_{40} positive ($n = 17/18$, 94.4%).

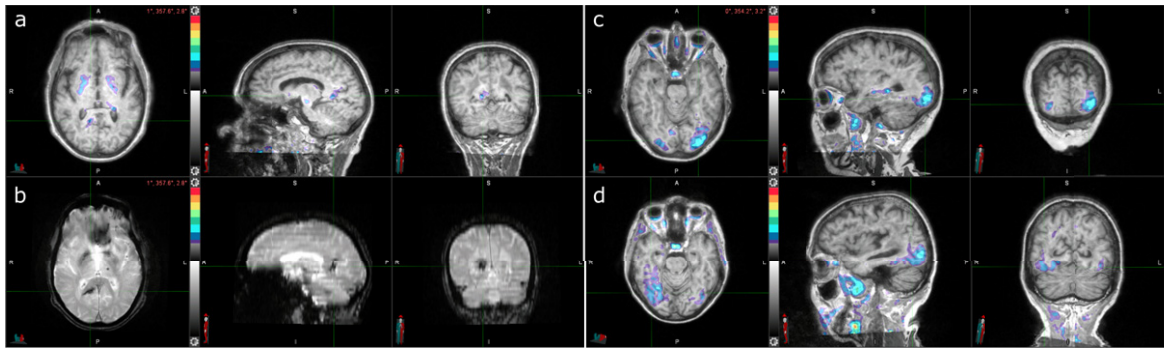


Fig. 2. Three cases with tau-positive visual interpretations, but tau-negative SUVRs. a) Tau PET coregistered with MRI of a male participant in his 80s with elevated right precuneus uptake. b) Corresponding T2*-weighted MRI showing a hypointensity (indicated by the crosshair) that colocalizes with the elevated right precuneus uptake from (a). c) Tau PET coregistered with MRI of a female participant in her 70s with elevated occipital lobe uptake, left greater than right. d) Tau PET coregistered with MRI of a female participant in her 70s with elevated posterolateral temporal, occipital, and parietal/precuneus lobe uptake, right greater than left.

Table 2
AD biomarker status for cases with positive tau PET visual interpretation but negative tau PET SUVR analysis

	Age	Sex	APOE	CDR [®]	A β PET (Centiloid)	Tau PET (SUVR)	CSF A β_{42} /A β_{40}	CSF p-tau181 (pg/ml)
Parietal/precuneus hemorrhagic infarct	80s	Male	3/4	0	3.87	1.19	0.0975	21.6
Left occipital	70s	Female	3/4	0	17.0 \rightarrow 50.0*	1.23	0.0523	69.2
Right occipital	70s	Female	3/3	0.5	72.1	1.31	0.0493	63.5

Numbers in bold denote positive biomarker status. *This participant had a Centiloid = 17.0 (below cutoff) approximately one year before their tau PET visit, and a Centiloid = 50.0 (above cutoff) approximately two years after their tau PET visit.

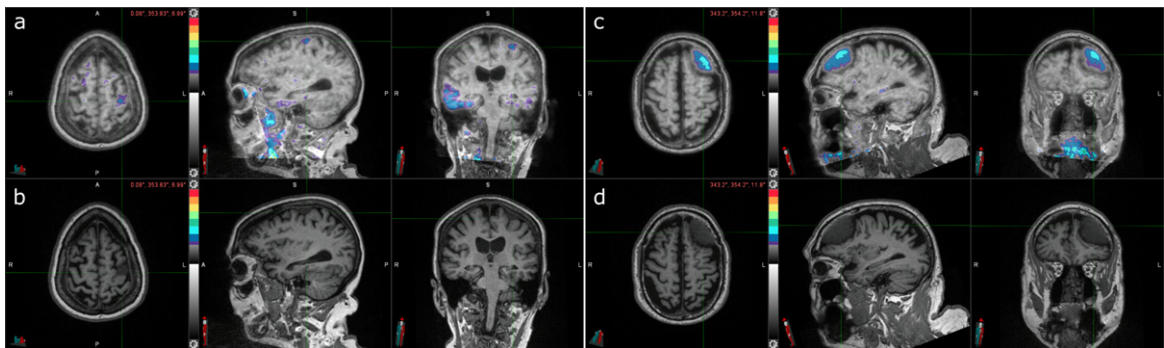


Fig. 3. Two cases of incidental meningioma with tau PET uptake. a) Tau PET coregistered with MRI of a female participant in her 70s with a left frontal posterior meningioma (indicated by the crosshair) with tau radiotracer uptake. b) Corresponding MRI image. c) Tau PET coregistered with MRI of a male participant in his 70s with a left frontal meningioma (indicated by the crosshair) with tau radiotracer uptake. d) Corresponding MRI image.

Table 3
AD biomarker status for cases with incidental meningioma

	Age	Sex	APOE	CDR [®]	A β PET (Centiloid)	Tau PET (SUVR)	CSF A β_{42} /A β_{40}	CSF p-tau181 (pg/ml)
Left posterior frontal meningioma	70s	Female	2/4	0	177	1.64	0.0481*	49.9*
Left frontal meningioma	70s	Male	2/3	0	8.94	1.16	0.0848*	30.3*

Numbers in bold denote positive biomarker status. *CSF lumbar punctures were performed approximately 10 years prior to tau PET.

Table 4
Participant characteristics for those who underwent lumbar puncture

		Cognitively normal	Cognitively impaired	Total
Number		126	18	144
Mean age in years (SD)		68.6 ± 8.32	76.1 ± 7.84	69.5 ± 8.60
Female (%)		70 (48.6)	10 (55.6)	80 (55.6)
Race	White	114	18	132
	Black or African American	11	0	11
	Asian	1	0	1
Mean MMSE (SD)		29.3 (1.07)	25.7 (3.90)	28.8 (2.06)
CDR®	0	126	0	126
	0.5	0	12	12
	1	0	5	5
	2	0	1	1
Clinical diagnosis	Cognitively normal	126	0	126
	Uncertain dementia	0	6	6
	AD dementia	0	12	12
APOE genotype	2/2	1	0	1
	2/3	22	1	23
	2/4	3	1	4
	3/3	62	7	69
	3/4	31	7	38
	4/4	6	2	8
	Unknown	1	0	1
Tau PET temporal meta-ROI SUVR	Mean ± SD [min, max]	1.15 ± 0.108 [0.924, 1.882]	1.47 ± 0.367 [1.042, 2.43]	1.19 ± 0.194 [0.924, 2.43]
	Positive (%)	3 (2.38)	11 (61.1)	14 (9.72)
Tau PET visual interpretation	Positive (%)	5 (3.97)	12 (66.7)	17 (11.8)
	Mean ± SD	19.0 ± 32.2	80.3 ± 46.7	26.7 ± 39.8
Aβ PET (Centiloid)	Positive (%)	34 (27.0)	16 (88.9)	50 (34.7)
	Mean ± SD	42.6 ± 30.4	88.7 ± 42.6	48.4 ± 35.5
CSF p-tau181	Positive (%)	19 (15.1)	14 (77.8)	33 (22.9)
	Mean ± SD	42.6 ± 30.4	88.7 ± 42.6	48.4 ± 35.5
CSF Aβ ₄₂ /Aβ ₄₀	Mean ± SD	0.0777 ± 0.0217	0.0502 ± 0.0196	0.0743 ± 0.0233
	Positive (%)	41 (32.5)	16 (88.9)	57 (39.6)

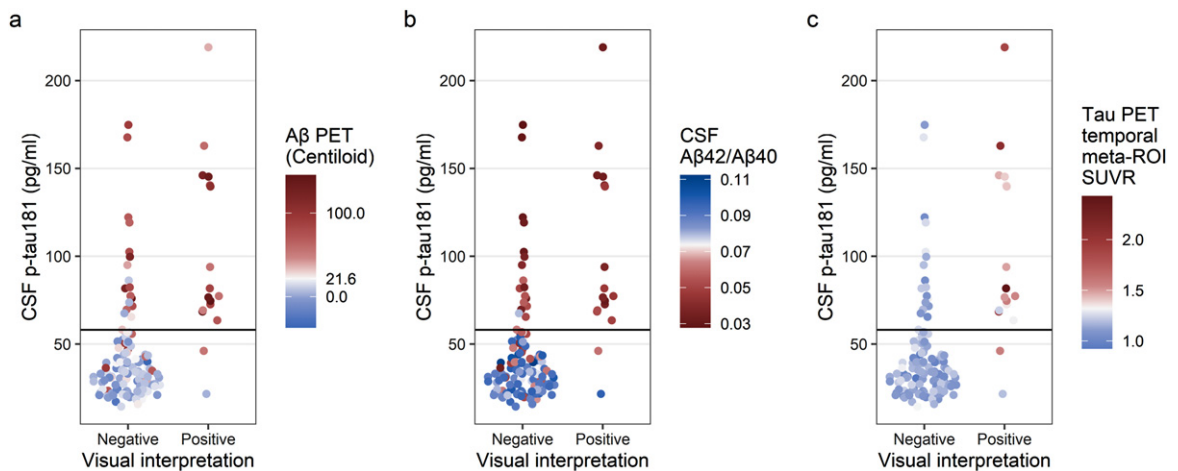


Fig. 4. Comparison of tau PET visual interpretation with CSF p-tau181 concentration. Each participant is plotted by visual interpretation (x-axis) and CSF p-tau181 concentration (y-axis); participants with p-tau181 ≥ 58.1 pg/ml were considered positive. In (a), the color indicates the Aβ PET status for each participant (positive Aβ PET, red; negative Aβ PET, blue; cutoff = 21.6 Centiloid). In (b), the color indicates the CSF Aβ₄₂/Aβ₄₀ status for each participant (positive CSF Aβ₄₂/Aβ₄₀, red; negative CSF Aβ₄₂/Aβ₄₀, blue; cutoff = 0.0737). In (c), the color indicates the tau PET temporal meta-ROI SUVR status for each participant (positive tau PET, red; negative tau PET, blue; cutoff = 1.32 SUVR).

Clinical and cognitive assessment

Six participants were assessed at baseline to be cognitively normal but tau-positive on visual interpretation (Table 5). One participant (Case 1) was previously mentioned to have PET radiotracer uptake colocalized to a parietal/precuneus hypointensity on T2*-weighted MRI and no other positive AD biomarkers (Fig. 2 and Table 2). The remaining five participants were all Aβ PET positive. No participant reliably converted from cognitively normal to AD dementia. One participant (Case 2) did convert to AD dementia at their three-year follow up but was reassessed to have a clinical diagnosis of uncertain dementia, more specifically, possible non-AD dementia of vascular origin at their five-year follow up. Another participant (Case 4) converted to AD dementia at their two-year follow up but was reassessed to have frontotemporal dementia (FTD) at their four-year follow up.

Four participants were assessed at baseline to be cognitively normal and tau-negative on visual interpretation but would convert to AD dementia at follow up (Table 5). Two participants (Case 7 and Case 10) converted to AD dementia at their one-year follow ups but were reassessed as cognitively normal at their two-year follow ups. The remaining two participants (Case 8 and Case 9) converted to AD dementia at their second- and fourth-year follow ups, respectively, but only Case 2 demonstrated Aβ PET positivity at baseline.

Twenty-three participants were assessed at baseline to have cognitive impairment (Table 6). Nine of these participants received a clinical assessment of uncertain dementia and two of the nine had a baseline tau-positive visual assessment. Both cases (Case 2 and Case 4) converted to AD dementia by their first- and second-year follow ups, respectively and were both Aβ PET positive. Nonetheless, three cases with a tau-negative visual interpretation at baseline (Case 5, Case 7, and Case 8) converted to AD dementia at their two-, two-, and three-year follow ups, respectively, although Case 5 was reassessed to be cognitively normal at their five-year follow up.

Thirteen of the 23 participants with baseline cognitive impairment received a clinical assessment of AD dementia. All 13 participants were Aβ PET positive (Table 6). Twelve of these participants had tau-positive visual interpretation; the remaining participant (Case 18) was tau PET negative, but at their one-year follow up had their clinical assessment changed to FTD. Additionally, Case 21 was tau PET

Table 5
Cognitively normal participant follow up

Age	Sex	APOE	Baseline tau PET temporal meta-ROI	Baseline tau PET visual interpretation	Asymmetry	Baseline Aβ PET (Centiloid)	Yearly follow-up clinical diagnosis				
							1	2	3	4	5
Baseline cognitively normal and tau PET visual interpretation positive											
1*	80s	Male	3/4	1.19	Right	3.87	CN	CN	CN	CN	CN
2	70s	Male	2/3	1.35	Left	86.6	CN	CN	AD	AD	UD
3**	70s	Female	3/4	1.23	Left	17.0 → 50.0	CN	CN	CN	CN	CN
4	80s	Male	3/3	1.88	Left	37.6	UD	AD	AD	FTD	FTD
5	80s	Male	3/4	1.54	Left	57.3	CN	CN	AD	AD	FTD
6***	70s	Female	2/4	1.64	Right	177	CN	CN	CN	CN	CN
Baseline cognitively normal (and tau PET visual interpretation negative), but converts at follow up											
7	70s	Female	3/4	1.22	Negative	-2.35	AD	CN	CN	CN	CN
8	80s	Female	3/4	1.21	Negative	120	CN	AD	AD	AD	AD
9	50s	Male	3/3	0.93	Negative	3.38	AD	UD	UD	AD	AD
10****	70s	Male	2/3	1.16	Negative	8.94	AD	CN	CN	CN	CN

Numbers in bold denote positive biomarker status. AD, Alzheimer's disease (dementia); CN, cognitively normal; FTD, frontotemporal dementia; UD, uncertain dementia. *Same case as the "Parietal/precuneus hemorrhagic infarct" case in Table 2. **Same case as the "Left occipital" case in Table 2. ***Same case as the "Left posterior frontal meningioma" case in Table 3. ****Same case as the "Left frontal meningioma" case in Table 3.

Table 6
Cognitively impaired participant follow up

Age	Sex	APOE	Baseline tau PET temporal meta-ROI	Baseline tau PET visual interpretation	Asymmetry	Baseline A β PET (Centiloid)	Yearly follow-up clinical diagnosis				
							1	2	3	4	5
Baseline uncertain dementia											
1	60s	Male	3/4	1.11	Negative			UD	UD	CN	UD
2	70s	Female	3/4	1.86	Positive		AD	AD	AD		
3	60s	Female	3/3	1.14	Negative	Right	CN		CN	CN	CN
4*	70s	Female	3/3	1.31	Positive	Right	UD	AD	AD	AD	
5	70s	Male	3/4	1.10	Negative	Right	CN	UD	AD	UD	CN
6	70s	Female	2/3	1.16	Negative		UD				
7	80s	Male	2/4	1.04	Negative		CN	AD			
8	70s	Female	4/4	1.29	Negative			UD	AD	AD	
9	70s	Male	3/4	1.02	Negative	Right	CN	CN	CN	CN	
Baseline 0.5 in memory only											
10	80s	Male	3/3	1.07	Negative		CN	CN			
Baseline AD dementia											
11	70s	Female	3/4	1.57	Positive		AD	AD			
12	60s	Male	3/3	2.43	Positive						
13	70s	Female	3/4	2.08	Positive		AD	AD	AD	AD	
14	80s	Male	3/3	1.46	Positive		AD	AD			
15	80s	Male	3/4	1.35	Positive		AD	AD	AD	AD	
16	70s	Female	4/4	1.58	Positive	Left	AD	AD			
17	70s	Female	3/4	1.52	Positive		AD				
18	70s	Female	3/4	1.18	Negative		FTD				
19	70s	Male	3/4	1.40	Positive	Right	AD	AD	AD	AD	
20	80s	Male	3/4	1.51	Positive		AD	AD			
21	80s	Female	3/3	1.41	Positive	Right	CN	CN			
22	70s	Female	3/4	1.48	Positive	Left	AD	AD			
23	50s	Male	3/3	2.01	Positive						

Numbers in bold denote positive biomarker status. AD, Alzheimer disease (dementia); CN, cognitively normal; FTD, frontotemporal dementia; UD, uncertain dementia. *Same case as the “Right occipital” case in Table 2.

positive, but was reassessed to be cognitively normal at their one- and two-year follow ups.

Conclusions

^{18}F -flortaucipir PET visual interpretation was found to be consistent between readers in this study ($n = 189/189$, 100%, $\kappa = 1$) and highly consistent with SUVR quantification ($n = 186/189$, 98.4%, $\kappa = 0.910$), suggesting these two approaches to determining tau PET positivity give similar results despite their different methodologies. However, three participants had discordant visual interpretations and SUVRs, likely due to a hemorrhagic infarct with elevated radiotracer uptake, an atypical, occipital-predominant presentation of AD NFT deposition, and a highly lateralized presentation of AD NFT deposition, respectively. These cases suggest the need for MR imaging to accompany ^{18}F -flortaucipir PET visual interpretation, and the need to consider regions outside the temporal meta-ROI for SUVR quantification.

Among non-AD sources of ^{18}F -flortaucipir uptake, the most studied is off-target binding in the choroid plexus, striatum, brainstem, and bone/meninges [24, 25]. In this study, off-target binding did not mimic the appearance of the AD tau pattern when assessed by visual readers, nor did it cause any tau PET temporal meta-ROI SUVR to be falsely positive when compared to visual interpretation. However, we observed two other sources of off-target binding that were not mentioned in the manufacturer's guidelines for ^{18}F -flortaucipir PET visual interpretation and which can potentially confound tau PET interpretations: hemorrhagic infarcts and meningiomas. The hemorrhagic infarct case was the case previously described as having a tau-positive visual interpretation and a tau-negative SUVR quantification. The two meningioma cases demonstrated elevated levels of radiotracer uptake in the frontal lobe, which is immaterial when assessing tau PET positivity by visual interpretation, but meningiomas in the posterolateral temporal, occipital, or parietal/precuneus regions might plausibly interfere with visual interpretation and SUVR quantification.

^{18}F -flortaucipir PET visual interpretation was found to be moderately consistent with CSF p-tau181 ($n = 124/144$, 86.1%, $\kappa = 0.526$). Most discordant cases ($n = 18/20$) are amyloid-positive and CSF p-tau181 positive, but tau-negative on visual interpretation. This suggests that the discordance between ^{18}F -flortaucipir PET and CSF p-tau181 may be

attributed to participants with early changes in AD pathophysiology. Moloney and colleagues have found in an autopsy study that p-tau181, 205, 217, and 231 fluid biomarker sites are present in the early stages of NFT maturity [26]. Wennström and colleagues have found that p-tau217 can be found within NFTs and neuropil threads along with p-tau181, 231, 202, 202/205, and 369/404, and that p-tau217 area fraction correlated with antemortem plasma p-tau217 in individuals with confirmed A β plaque pathology [27]. Furthermore, plasma p-tau and CSF p-tau have been shown to be strongly correlated [28–30]. Taken together, these findings suggest a possibility that CSF p-tau181 is tracking changes in AD pathophysiology that occur earlier than the more advanced stages of NFT aggregation that ^{18}F -flortaucipir PET may be more sensitive to.

When interpreting tau PET visual interpretation alongside clinical diagnosis after the study (both visual interpretation and clinical diagnosis were performed independently) a few relationships between the two kinds of AD diagnoses were remarkable. First, a baseline tau-positive visual interpretation in participants who were cognitively normal at baseline did not reliably predict conversion to AD dementia at follow up. If anything, tau PET positivity in cognitively normal participants was more likely to be either a sign of atypical AD, of related dementias (vascular dementia or FTD), or of resilience to AD dementia. Second, a baseline tau-negative visual interpretation in participants who were cognitively normal at baseline did not rule out conversion to AD dementia at follow up. Four cases were found to demonstrate conversion to AD dementia at follow up under these circumstances, although two of these were later reassessed to be cognitively normal. Third, baseline tau PET positivity in cognitively impaired participants did not guarantee a diagnosis of AD dementia at follow up: one participant was assessed to be cognitively normal at follow up even under these circumstances and another was reassessed to have FTD. Finally, baseline tau PET negativity in cognitively impaired participants cannot be used to rule out conversion to AD dementia at follow up: three such participants converted to AD dementia at their follow up visits, respectively, although one was reassessed to be cognitively normal at a later date.

A bias of the current study lies in the inclusion of cognitively normal participants. In a clinical setting, ^{18}F -flortaucipir PET is indicated for use in patients with cognitive impairment. Two of the three cases discordant between visual interpretation and

SUVR quantification in this study were from cognitively normal participants and would not warrant the use of ^{18}F -flortaucipir visual interpretation in a clinical setting to begin with. Six of the 20 cases discordant between visual interpretation and CSF p-tau181 quantification were from cognitively normal participants and also would not warrant the use of ^{18}F -flortaucipir visual interpretation in a clinical setting. Furthermore, the inclusion of cognitively normal participants, who represent the majority of the cases studied, also introduces a negative case bias, as they also represent a majority of the tau PET negative cases. Since most of these cases have tau PET SUVRs much lower than the SUVR positivity threshold, the agreement between visual interpretation and SUVRs is higher in the current study compared to a more challenging study comparing exclusively borderline cases. Nonetheless, exploring tau positivity in cognitively normal participants in this study identified individuals who have atypical AD tau and clinical progression.

The current study is focused on ^{18}F -flortaucipir PET, and its conclusions do not necessarily apply to other tau PET radiotracers, which may have their own distinctive characteristics regarding off-target binding and sensitivity, which need to be accounted for on a tracer-by-tracer basis in studies of visual interpretation guidelines [31].

Future studies may also explore the discordances between tau PET visual interpretation and tau PET SUVR more thoroughly by investigating those participants with questionable or very mild dementia. Future studies may also explore the consequences of using regions of interest beyond the temporal meta-ROI used in the current study, such as the MUBADA [32]. These alternative ROIs may be appropriate for increasing concordance between visual interpretation and SUVR quantification, especially if crucial differences between the two lie in elevated radiotracer uptake outside the temporal meta-ROI. That said, the most direct way of harmonizing visual interpretation and SUVR quantification may be to construct an entirely new ROI for SUVR quantification that covers the critical regions in visual interpretation (posterolateral temporal, occipital, and parietal/precuneus regions) and develop a quantification method sensitive to contiguous uptake within this ROI.

In conclusion, ^{18}F -flortaucipir PET visual interpretation can identify atypical AD NFT deposition that may be missed by SUVR quantification depending upon the regions of interest used. However, while the manufacturer's guidelines for ^{18}F -flortaucipir

PET visual interpretation address non-AD sources of uptake such as off-target binding, they do not address other non-AD sources of uptake such as hemorrhagic infarcts and meningiomas. Temporal meta-ROI SUVR was highly concordant with visual interpretation for the cohort considered in this study. However, SUVR analyses could not detect lateralized occipital-predominant AD NFT deposition because the occipital lobe falls outside the temporal meta-ROI. CSF p-tau181 concentration was moderately concordant with visual interpretation and enabled detection of early changes in AD pathophysiology associated with tau hyperphosphorylation. However, these changes cannot be seen on PET. Finally, a positive visual interpretation did not make a follow up diagnosis of AD dementia inevitable, and a negative visual interpretation did not exclude the possibility of a follow up diagnosis of AD dementia. Additional work is needed to understand how multiple AD PET and CSF biomarkers might conceivably be used in tandem in a clinical setting alongside AD clinical evaluation in order to correctly diagnose and treat all individuals, not just those who demonstrate AD biomarker and clinical findings concordant with group-level trends.

ACKNOWLEDGMENTS

We thank the altruism of participants and their families and contributions of Knight ADRC support staff. Avid Radiopharmaceuticals provided technology transfer and precursor for ^{18}F -flortaucipir and ^{18}F -florbetapir.

FUNDING

C.D.C. received support from the Knight ADRC T32 Fellowship (5T32AG058518-04) and the NSF GRFP Fellowship (DGE-1745038 and DGE-2139839). S.E.S. received support from NIA R01AG070941 and Barnes-Jewish Hospital Foundation. A.M.F. received support from various NIH grants (P30AG066444, P01AG003991, P01AG026276, U19AG032438). J.C.M. received support from various NIH grants (P30AG066444, P01AG003991, P01AG026276, U19AG032438).

CONFLICT OF INTEREST

C.D.C., J.A.L., and S.F. have no disclosures. M.R.P. received consulting fees from NFL Con-

cussion Settlement Program; and received payment/honoraria from the University of Texas Medical School Galveston (Cooley Visiting Professor for the Department of Radiology). S.E.S. received support from NIA R01AG070941 and Barnes-Jewish Hospital Foundation; received payment/honoraria from the University of Wisconsin, University of Washington, University of Indiana, and St. Luke's Hospital; is a board member of Greater Missouri Alzheimer's Association; and received data on behalf of Washington University from C2N Diagnostics at no cost. A.M.F. received support from various NIH grants (P30AG066444, P01AG003991, P01AG026276, U19AG032438); received consulting fees from DiamiR and Siemens Healthcare Diagnostics Inc.; and participated on an advisory board at Roche Diagnostics, Genentech, and Diadem. J.C.M. received support from various NIH grants (P30AG066444, P01AG003991, P01AG026276, U19AG032438); received consulting fees from Barcelona Brain Research Center and TS Srinivasan Advisory Board; received payment or honoraria from Montefiore Grand Rounds and Tetra-Inst ADRC seminar series; and participated on an advisory board at Cure Alzheimer's Fund. T.L.S.B. has investigator-initiated research funding from NIH, Alzheimer's Association, Barnes-Jewish Hospital foundation, and Avid Radiopharmaceuticals (a wholly owned subsidiary of Eli Lilly); participates as a site investigator in clinical trials sponsored by Avid Radiopharmaceuticals, Eli Lilly, Biogen, Eisai, Janssen, and Roche; serves as an unpaid consultant to Eisai and Siemens; and is on the Speaker's Bureau for Biogen.

DATA AVAILABILITY

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

SUPPLEMENTARY MATERIAL

The supplementary material is available in the electronic version of this article: <https://dx.doi.org/10.3233/JAD-230032>.

REFERENCES

[1] Braak H, Alafuzoff I, Arzberger T, Kretschmar H, Del Tredici K (2006) Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sec-

tions and immunocytochemistry. *Acta Neuropathol* **112**, 389-404.

- [2] Braak H, Thal DR, Ghebremedhin E, Del Tredici K (2011) Stages of the pathologic process in Alzheimer disease: Age categories from 1 to 100 years. *J Neuropathol Exp Neurol* **70**, 960-969.
- [3] Blennow K, Wallin A, Agren H, Spenger C, Siegfried J, Vanmechelen E (1995) Tau protein in cerebrospinal fluid: A biochemical marker for axonal degeneration in Alzheimer disease? *Mol Chem Neuropathol* **26**, 231-245.
- [4] Chien DT, Bahri S, Szardenings AK, Walsh JC, Mu F, Su M-Y, Shankle WR, Elizarov A, Kolb HC (2013) Early clinical PET imaging results with the novel PHF-tau radioligand [F-18]-T807. *J Alzheimers Dis* **34**, 457-468.
- [5] Villemagne VL, Furumoto S, Fodero-Tavoletti MT, Mulligan RS, Hodges J, Harada R, Yates P, Piguet O, Pejoska S, Doré V, Yanai K, Masters CL, Kudo Y, Rowe CC, Okamura N (2014) *In vivo* evaluation of a novel tau imaging tracer for Alzheimer's disease. *Eur J Nucl Med Mol Imaging* **41**, 816-826.
- [6] Betthausen TJ, Cody KA, Zammit MD, Murali D, Converse AK, Barnhart TE, Stone CK, Rowley HA, Johnson SC, Christian BT (2019) *In vivo* characterization and quantification of neurofibrillary tau PET radioligand 18F-MK-6240 in humans from Alzheimer disease dementia to young controls. *J Nucl Med* **60**, 93-99.
- [7] Jack CR, Wiste HJ, Weigand SD, Therneau TM, Lowe VJ, Knopman DS, Gunter JL, Senjem ML, Jones DT, Kantarci K, Machulda MM, Mielke MM, Roberts RO, Vemuri P, Reyes DA, Petersen RC (2017) Defining imaging biomarker cut-points for brain aging and Alzheimer's disease. *Alzheimers Dement* **13**, 205-216.
- [8] Schwarz CG, Therneau TM, Weigand SD, Gunter JL, Lowe VJ, Przybelski SA, Senjem ML, Botha H, Vemuri P, Kantarci K, Boeve BF, Whitwell JL, Josephs KA, Petersen RC, Knopman DS, Jack CR (2021) Selecting software pipelines for change in flortaucipir SUVR: Balancing repeatability and group separation. *Neuroimage* **238**, 118259.
- [9] Boerwinkle AH, Wisch JK, Chen CD, Gordon BA, Butt OH, Schindler SE, Sutphen C, Flores S, Dincer A, Benzinger TLS, Fagan AM, Morris JC, Ances BM (2021) Temporal correlation of CSF and neuroimaging in the amyloid-tau-neurodegeneration model of Alzheimer disease. *Neurology* **97**, e76-e87.
- [10] La Joie R, Bejanin A, Fagan AM, Ayakta N, Baker SL, Bourakova V, Boxer AL, Cha J, Karydas A, Jerome G, Maass A, Mensing A, Miller ZA, O'Neil JP, Pham J, Rosen HJ, Tsai R, Visani AV, Miller BL, Jagust WJ, Rabinovici GD (2018) Associations between [18F]AV1451 tau PET and CSF measures of tau pathology in a clinical sample. *Neurology* **90**, e282-e290.
- [11] Bucci M, Chiotis K, Nordberg A (2021) Alzheimer's disease profiled by fluid and imaging markers: Tau PET best predicts cognitive decline. *Mol Psychiatry* **26**, 5888-5898.
- [12] Ossenkopppele R, Reimand J, Smith R, Leuzy A, Strandberg O, Palmqvist S, Stomrud E, Zetterberg H, Alzheimer's Disease Neuroimaging Initiative, Scheltens P, Dage JL, Bouwman F, Blennow K, Mattsson-Carlsson N, Janelidze S, Hansson O (2021) Tau PET correlates with different Alzheimer's disease-related features compared to CSF and plasma p-tau biomarkers. *EMBO Mol Med* **13**, e14398.
- [13] Weintraub S, Salmon D, Mercaldo N, Ferris S, Graff-Radford NR, Chui H, Cummings J, DeCarli C, Foster NL, Galasko D, Peskind E, Dietrich W, Beekly DL, Kukull WA,

- Morris JC (2009) The Alzheimer's Disease Centers' Uniform Data Set (UDS): The neuropsychological test battery. *Alzheimer Dis Assoc Disord* **23**, 91-101.
- [14] Morris JC (1993) The Clinical Dementia Rating (CDR): Current version and scoring rules. *Neurology* **43**, 2412-2414.
- [15] Folstein MF, Folstein SE, McHugh PR (1975) "Mini-mental state": A practical method for grading the cognitive state of patients for the clinician. *J Psychiatr Res* **12**, 189-198.
- [16] Su Y, Blazey TM, Snyder AZ, Raichle ME, Marcus DS, Ances BM, Bateman RJ, Cairns NJ, Aldea P, Cash L, Christensen JJ, Friedrichsen K, Hornbeck RC, Farrar AM, Owen CJ, Mayeux R, Brickman AM, Klunk W, Price JC, Thompson PM, Ghetti B, Saykin AJ, Sperling RA, Johnson KA, Schofield PR, Buckles V, Morris JC, Benzinger TLS, Dominantly Inherited Alzheimer Network (2015) Partial volume correction in quantitative amyloid imaging. *Neuroimage* **107**, 55-64.
- [17] Su Y, D'Angelo GM, Vlassenko AG, Zhou G, Snyder AZ, Marcus DS, Blazey TM, Christensen JJ, Vora S, Morris JC, Mintun MA, Benzinger TLS (2013) Quantitative analysis of PiB-PET with FreeSurfer ROIs. *PLoS One* **8**, e73377.
- [18] Fischl B (2012) FreeSurfer. *Neuroimage* **62**, 774-781.
- [19] Su Y, Flores S, Hornbeck RC, Speidel B, Vlassenko AG, Gordon BA, Koeppe RA, Klunk WE, Xiong C, Morris JC, Benzinger TLS (2018) Utilizing the Centiloid scale in cross-sectional and longitudinal PiB PET studies. *Neuroimage Clin* **19**, 406-416.
- [20] Klunk WE, Koeppe RA, Price JC, Benzinger TL, Devous MD, Jagust WJ, Johnson KA, Mathis CA, Minhas D, Pontecorvo MJ, Rowe CC, Skovronsky DM, Mintun MA (2015) The Centiloid Project: Standardizing quantitative amyloid plaque estimation by PET. *Alzheimers Dement* **11**, 1-15.e1-4.
- [21] Zhu X, Melnykov V (2018) Manly transformation in finite mixture modeling. *Comput Stat Data Anal* **121**, 190-208.
- [22] R Core Team (2021) *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria.
- [23] Vogel JW, Young AL, Oxtoby NP, Smith R, Ossenkoppele R, Strandberg OT, La Joie R, Aksman LM, Grothe MJ, Iturria-Medina Y, Pontecorvo MJ, Devous MD, Rabinovici GD, Alexander DC, Lyoo CH, Evans AC, Hansson O (2021) Four distinct trajectories of tau deposition identified in Alzheimer's disease. *Nat Med* **27**, 871-881.
- [24] Baker SL, Harrison TM, Maass A, Joie RL, Jagust WJ (2019) Effect of off-target binding on 18F-Flortaucipir variability in healthy controls across the life span. *J Nucl Med* **60**, 1444-1451.
- [25] Smith R, Strandberg O, Leuzy A, Bethausen TJ, Johnson SC, Pereira JB, Hansson O (2021) Sex differences in off-target binding using tau positron emission tomography. *Neuroimage Clin* **31**, 102708.
- [26] Moloney CM, Labuzan SA, Crook JE, Siddiqui H, Castanedes-Casey M, Lachner C, Petersen RC, Duara R, Graff-Radford NR, Dickson DW, Mielke MM, Murray ME (2023) Phosphorylated tau sites that are elevated in Alzheimer's disease fluid biomarkers are visualized in early neurofibrillary tangle maturity levels in the post mortem brain. *Alzheimers Dement* **19**, 1029-1040.
- [27] Wennström M, Janelidze S, Nilsson KPR, Serrano GE, Beach TG, Dage JL, Hansson O, The Netherlands Brain Bank (2022) Cellular localization of p-tau217 in brain and its association with p-tau217 plasma levels. *Acta Neuropathol Commun* **10**, 3.
- [28] Barthélemy NR, Horie K, Sato C, Bateman RJ (2020) Blood plasma phosphorylated-tau isoforms track CNS change in Alzheimer's disease. *J Exp Med* **217**, e20200861.
- [29] Janelidze S, Mattsson N, Palmqvist S, Smith R, Beach TG, Serrano GE, Chai X, Proctor NK, Eichenlaub U, Zetterberg H, Blennow K, Reiman EM, Stomrud E, Dage JL, Hansson O (2020) Plasma P-tau181 in Alzheimer's disease: Relationship to other biomarkers, differential diagnosis, neuropathology and longitudinal progression to Alzheimer's dementia. *Nat Med* **26**, 379-386.
- [30] Suárez-Calvet M, Karikari TK, Ashton NJ, Rodríguez JL, Milà-Alomà M, Gispert JD, Salvadó G, Mingüillon C, Fauria K, Shekari M, Grau-Rivera O, Arenaza-Urquijo EM, Sala-Vila A, Sánchez-Benavides G, González-de-Echavarrri JM, Kollmorgen G, Stoops E, Vanmechelen E, Zetterberg H, Blennow K, Molinuevo JL (2020) Novel tau biomarkers phosphorylated at T181, T217 or T231 rise in the initial stages of the preclinical Alzheimer's continuum when only subtle changes in A β pathology are detected. *EMBO Mol Med* **12**, e12921.
- [31] Shuping JL, Matthews DC, Adamczuk K, Scott D, Rowe CC, Kreisl WC, Johnson SC, Lukic AS, Johnson KA, Rosa-Neto P, Andrews RD, Van Laere K, Cordes L, Ward L, Wilde CL, Barakos J, Purcell DD, Devanand DP, Stern Y, Luchsinger JA, Sur C, Price JC, Brickman AM, Klunk WE, Boxer AL, Mathotaarachchi SS, Lao PJ, Evelhoch JL (2023) Development, initial validation, and application of a visual read method for [18F]MK-6240 tau PET. *Alzheimers Dement (N Y)* **9**, e12372.
- [32] Pontecorvo MJ, Devous MD, Kennedy I, Navitsky M, Lu M, Galante N, Salloway S, Doraiswamy PM, Southehal S, Arora AK, McGeehan A, Lim NC, Xiong H, Trucocchio SP, Joshi AD, Shcherbinin S, Teske B, Fleisher AS, Mintun MA (2019) A multicentre longitudinal study of flortaucipir (18F) in normal ageing, mild cognitive impairment and Alzheimer's disease dementia. *Brain* **142**, 1723.