

Hypothesis

The Adult Neurogenesis Theory of Alzheimer's Disease

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Accepted 6 April 2023

Pre-press 5 May 2023

Abstract. Alzheimer's disease starts in neural stem cells (NSCs) in the niches of adult neurogenesis. All primary factors responsible for pathological tau hyperphosphorylation are inherent to adult neurogenesis and migration. However, when amyloid pathology is present, it strongly amplifies tau pathogenesis. Indeed, the progressive accumulation of extracellular amyloid- β deposits in the brain triggers a state of chronic inflammation by microglia. Microglial activation has a significant pro-neurogenic effect that fosters the process of adult neurogenesis and supports neuronal migration. Unfortunately, this "reactive" pro-neurogenic activity ultimately perturbs homeostatic equilibrium in the niches of adult neurogenesis by amplifying tau pathogenesis in AD. This scenario involves NSCs in the subgranular zone of the hippocampal dentate gyrus in late-onset AD (LOAD) and NSCs in the ventricular-subventricular zone along the lateral ventricles in early-onset AD (EOAD), including familial AD (FAD). Neuroblasts carrying the initial seed of tau pathology travel throughout the brain via neuronal migration driven by complex signals and convey the disease from the niches of adult neurogenesis to near (LOAD) or distant (EOAD) brain regions. In these locations, or in close proximity, a focus of degeneration begins to develop. Then, tau pathology spreads from the initial foci to large neuronal networks along neural connections through neuron-to-neuron transmission.

Keywords: Alzheimer's disease, amyloid, microglia, neurofibrillary tangles, neurogenesis, tauopathies

INTRODUCTION

The amyloid hypothesis

The prevailing view in the field is that amyloid- β peptide ($A\beta$) exhibits a "toxic gain-of-function" when it forms oligomers and aggregates into plaques, directly contributing to the pathogenesis of Alzheimer's disease (AD) [1, 2]. In particular, the amyloid hypothesis, the prevalent theory of AD pathogenesis, suggests that the accumulation of pathological forms of $A\beta$ is the primary pathological process driven by an imbalance between $A\beta$

production and $A\beta$ clearance [3, 4]. In this pathway, microtubule-associated protein tau pathology with the formation of phospho-tau-immunoreactive neurofibrillary tangles (NFTs) and subsequent neuronal dysfunction and neurodegeneration, perhaps mediated via inflammation, are thought to be the downstream result [4]. The direct influence of $A\beta$ on tau pathogenesis is well documented. For example, injection of $A\beta$ fibrils [5] or $A\beta$ -containing brain extract [6] into mutant tau transgenic mice, crossed between mutant tau and amyloid precursor protein (APP) or 5x familial AD (FAD) transgenic mice, results in exacerbated tau pathology [5–16]. Moreover, "*in vitro*" [17] and "*in vivo*" [16] studies have demonstrated that $A\beta$ exerts its detrimental actions by activating a key kinase, glycogen synthase kinase 3 β (GSK-3 β) [17, 18], implicating this kinase as an

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important player in the amyloid cascade. Notably, GSK-3 β is the primary kinase that phosphorylates tau [18, 19]. In agreement, increased GSK-3 β activity has been observed in the brains of AD patients [20]. These data confirm GSK-3 β as a cornerstone of AD pathogenesis and support the notion that this kinase represents a crucial molecular link between A β and tau [18, 19, 21–24]. Accordingly, human A β oligomers induce hyperphosphorylation of tau at AD-relevant epitopes and cause neuritic dystrophy in cultured neurons [25].

The current view

In the current theory, I propose a shift in the paradigm wherein aggregates of the two key players in AD pathogenesis, i.e., A β and tau peptide, develop by two different and relatively independent processes. In particular, the central hypothesis is that tau pathogenesis is linked to adult neurogenesis and migration. All elements predisposing to pathological tau hyperphosphorylation are present in the niches of adult neurogenesis. In contrast, as already documented in the literature, metabolism plays a primary role in driving A β deposition [26–28]. Despite the fact that the two processes driving A β and tau pathogenesis are relatively independent, when A β pathology is present, it acts as a strong driving force for tau pathogenesis. Therefore, A β pathology also plays a crucial role in AD pathogenesis in the current theory. However, its detrimental effect is explained in quite a different way from the classical amyloid hypothesis. In particular, A β not only has a downstream effect on tau pathogenesis, especially when A β and tau colocalize, but also has an early indirect effect by influencing the process of adult neurogenesis and migration. In brief, the current theory depicts the following scenario. Progressive accumulation of extracellular A β deposits in the brain triggers a state of chronic inflammation by microglia. Microglial activation has a significant pro-neurogenic effect that fosters adult neurogenesis and supports neuronal migration. Unfortunately, this “reactive” pro-neurogenic pathway ultimately perturbs the delicate homeostatic equilibrium in the neurogenic niches by amplifying tau pathogenesis in AD. An imbalance between increased tau phosphorylation, already occurring at a high rate in neural stem cells (NSCs), coupled with less efficient clearance of the byproducts of tau hyperphosphorylation, as well as further increases in hyperphosphorylation during long migrations, could be the primary rea-

sons behind these detrimental effects. This scenario involves NSCs in the subgranular zone (SGZ) of the hippocampal dentate gyrus (DG) in late-onset AD (LOAD) and NSCs in the ventricular-subventricular zone (V-SVZ) along the lateral ventricles in early-onset AD (EOAD), including familial AD (FAD). Neuroblasts carrying the initial seed of tau pathology travel throughout the brain by neuronal migration driven by complex signals, bringing the disease from the niches of adult neurogenesis to near (LOAD) or distant (EOAD) brain regions. At these locations, or in close proximity, a focus of degeneration begins to develop. Then, tau pathology spreads from the initial foci to large neuronal networks along neural connections by neuron-to-neuron transmission.

Therefore, the new core statement of the current theory is that AD starts in NSCs in the niches of adult neurogenesis. Interestingly, recent findings suggest that the current paradigm and the classic amyloid hypothesis might not be incompatible. In particular, some authors found clear evidence of intracellular oligomers of A β generated in NSCs within the SGZ niche at a very early stage in a transgenic AD mouse model [29]. In the current theory, this finding could be interpreted as proof that A β pathology not only indirectly influences tau pathogenesis by fostering neurogenesis and migration but also directly contributes to pathological tau hyperphosphorylation within these niches, most likely by activating GSK-3 β . This view also reinforces the role of amyloid pathology as a leading factor in the current model, bridging the gap between the two theories.

In the first section of this manuscript, I propose the core hypothesis of tau pathogenesis in AD as linked to adult neurogenesis and migration. In the second section, I present the indirect microglia-mediated A β -tau interaction. In the third section, I consider the scaling of molecular pathology to the macroscopic brain. In the Discussion section, I put the new theory into context and consider the potential merits and limitations of the current proposal.

TAU PATHOGENESIS IS LINKED TO ADULT NEUROGENESIS AND MIGRATION

Tau isoform and phosphorylation during postnatal and adult neurogenesis

The tau isoform featured by the presence of three-repeat microtubule-binding domains (3R-tau) predominates at early developmental stages [30, 31].

The 3R-tau isoform exhibits a lower affinity for microtubules than the mature brain tau isoform [32], so it confers lower stability to the cytoskeleton and allows the morphological differentiation and migration of developing neurons. In contrast, 4R-tau is the predominant isoform at mature developmental stages [30, 31]. It binds microtubules with a greater affinity and displaces the previously bound 3R-tau from microtubules [30], guaranteeing the stability of the cytoskeleton required to maintain neuronal integrity. In addition to the presence or absence of exon 10 shaping the 4R- or 3R-tau isoform, tau phosphorylation is developmentally regulated: it is higher in fetal neurons and decreases with age during development [33–35]. As phosphorylation decreases the affinity of tau protein for microtubules [36], hyperphosphorylation of fetal tau [33, 37] contributes to maintaining a dynamic microtubule network as required by the outgrowth of axons during embryogenic neurogenesis [38].

In the adult human brain, both 3R- and 4R-tau are present, although in newborn neurons in the niches of adult neurogenesis, 3R-tau is the primary isoform [32, 39]. In particular, it has been demonstrated that 3R-tau is transiently expressed during the maturation of NSCs in the hippocampal SGZ [32, 38, 40, 41]. For instance, in rodents, individual new subgranular neurons exhibit the highest expression of 3R-tau when cells are 2 weeks old [40], and expression of this molecule is maintained until 4 weeks, a time point at which 3R-tau is replaced by 4R-tau [32]. Moreover, high tau phosphorylation in fetal epitopes is related to adult neurogenesis in both the V-SVZ and SGZ [39, 42], although fetal tau phosphorylation can be found in the adult brain in additional areas [35]. Transient expression of the 3R-tau isoform and fetal tau hyperphosphorylation in adult neurogenesis are not unexpected, considering that new neurons require a high degree of plasticity to migrate, differentiate, project axons, and integrate into the cell layer, and both the 3R-tau isoform and high phosphorylation guarantee a dynamic microtubule network [38, 39]. Furthermore, abnormally hyperphosphorylated tau in AD constitutes paired helical filaments (PHFs) of NFTs [43–47]. Interestingly, the 3R-tau isoform is said to facilitate PHFs, such as those seen in classical AD NFTs [48]. Additionally, several sites of hyperphosphorylation of the fetal 3R-tau during development were the same as those in the AD brain [33, 37, 49, 50]. Additionally, as already reported, GSK-3 β is the first identified tau kinase [51, 52] that plays a key role in AD-like tau hyper-

phosphorylation [18, 19, 21, 53–55]. Interestingly, during development, expression of GSK-3 β reaches its highest level in the late embryonic/early postnatal period, markedly decreasing with maturation into adulthood [56, 57]. More importantly, activated GSK-3 β is believed to be the primary tau kinase in newborn neurons during adult neurogenesis [39, 58].

In summary, tau isoform and phosphorylation in NSCs during postnatal, as well as adult, neurogenesis seem equivalent to those predisposing to the typical tau alterations observed in AD. In this regard, it is worth noting that the primary cause of the tau functional change and NFT formation in AD is believed to be abnormal hyperphosphorylation [59–67]. In addition, abnormal tau hyperphosphorylation seems to reflect exaggerated physiological phosphorylation rather than disorganized phosphorylation at random sites [68, 69]. Therefore, tau pathogenesis in AD seems to depend exclusively on the extent of phosphorylation and the combination of multiple specific phosphorylation sites [38, 70, 71].

Differences between adult and postnatal neurogenesis associated with tau pathogenesis

Interestingly, although 3R-tau is said to facilitate PHFs, such as those seen in classical AD NFTs [48], and several sites of high phosphorylation of the fetal 3R-tau are the same as those in the AD brain [33, 37, 49, 50], during development, fetal tau remains functional and does not polymerize into NFTs. At this point, I speculate that a further crucial factor responsible for tau pathogenesis in AD could be found among those aspects that distinguish adult and postnatal neurogenesis. In this regard, it is worth noting that although postnatal and adult neurogenesis share some niches and signals, there are some important differences between the two.

The primary difference is that the neurogenic niches are surrounded by different environments. In particular, in the large, evolutionarily developed brain of adult mammals, neuroblasts originating in the neurogenic niches must migrate long distances through a complex and generally inhibitory environment [72] made up of neuronal, glial, and vascular networks to reach their destination [73]. Considering that tau hyperphosphorylation in neuroblasts contributes to maintaining a dynamic microtubule network that is amenable to migration, demanding and long migration through the inhibitory environment of the adult brain could have the detrimental effect of further increasing tau hyperphosphorylation. I believe this

factor could be crucial to tau pathogenesis, especially in EOAD. Indeed, in the current theory, EOAD pathogenesis is linked to adult neurogenesis, especially in the V-SVZ. Here, migrating neuroblasts carrying the seeds of tau pathology deviate from the conventional rostral migration stream (RMS) to the olfactory bulb (OB) and take different and long migration paths toward various regions of the cortex, driven by complex signals released, in particular, by activated microglia.

I believe a further difference between adult and postnatal neurogenesis relevant to the current theory is related to clearance activity in the niches. In this respect, rapidly accumulating data suggest that autophagy fulfils some roles in NSC function [74, 75]. Specifically, autophagy may serve both a surveillance role by ensuring the quality of NSCs by degrading and eliminating intracellular components and aggregates and an elimination role by ensuring the removal of defective or damaged NSCs through its cell death promotion abilities [74, 75]. Consequently, reduced autophagy during aging compared to the postnatal period may contribute to the accumulation of hyperphosphorylated tau aggregates in NSCs. This event would be a crucial factor in AD pathogenesis, especially in LOAD. Indeed, in the current theory, LOAD is primarily linked to neurogenesis in the hippocampal SGZ, where neural precursors migrate very briefly to the granular layer of the DG. Accordingly, an increase in tau hyperphosphorylation through migration would be a less relevant factor in LOAD pathogenesis than in EOAD pathogenesis.

In summary, in the niches of adult neurogenesis, all conditions predispose patients to pathological tau hyperphosphorylation and accumulation of tau aggregates. When microglia activated by A β deposition foster neurogenesis in the niches and support long migrations, the situation is taken to extremes.

THE MICROGLIA-MEDIATED A β -TAU INTERACTION

According to the current theory, the slow progressive accumulation of A β deposits in the brain provokes microglial activation. Activated microglia foster the process of neurogenesis in niches and support neuronal migration. This “reactive” increase in neurogenesis and migration amplifies tau pathogenesis in AD, and available data in the literature seem to support this scenario.

Microglial activation in AD

Microglial activation in AD is well documented [76]. In the early AD brain, microglia are found in high densities surrounding A β plaques [77]. Both postmortem [78, 79] and “*in vivo*” clinical studies using PET ligands that bind to activated microglia [80, 81] have consistently confirmed the finding that microglia colocalize with amyloid plaques in AD. In particular, senile plaques are infiltrated by astrocytes and microglia in and around their central amyloid core [82, 83]. From this evidence, it has been proposed that A β plaques stimulate a chronic inflammatory reaction [84]. In other words, the activation and increased proliferation of microglia in AD [85] are thought to result from glial reactions to events related to the ongoing deposition of A β [86, 87]. In this regard, the high density of microglia found around A β plaques is consistent with their role in A β clearance pathways and their activation by A β itself [88]. Moreover, the finding that microglia possess a range of pattern recognition receptors, including Toll-like receptors, receptors for advanced glycation end products, and scavenger receptors, many of which can recognize different A β species through various interactions of differing affinities, support their role in A β clearance [89–91]. Once activated, microglia and astrocytes produce several proinflammatory signaling molecules, including cytokines, growth factors, complement molecules, cell adhesion molecules, and chemokines [84, 92, 93]. In agreement with these findings, increased levels of inflammatory cytokines and chemokines [78] as well as upregulated chemokine receptors [94], have been found in the AD brain.

Microglia and adult neurogenesis

At this point, it is relevant to the current theory to disclose that microglia have been found to have a role in adult neurogenesis.

Microglia modulates the production of new neurons

In particular, microglia can modulate the production of new neurons in the adult brain [88]. NSCs have been found to depend on signals from their niche to regulate their self-renewal, proliferation, and differentiation [88]. In the absence of microglia, NSCs progressively lose the capacity to undergo the differentiation process required for neurogenesis [95]. The role of microglia in neurogenesis can be seen

as instructive, with microglial-secreted factors, such as IGF-1 and trypsinogen, having the capacity to regulate adult NSC proliferation and differentiation, promoting neurogenesis [96, 97]. Interestingly, both acute and chronic microglial activation can modulate neurogenesis [88]. In particular, “*in vitro*”, microglia acutely activated with lipopolysaccharide (LPS) strongly expressing IL-1, IL-6, and TNF- α have been found to reduce neural progenitor cell survival [98]. The study of some neurological disorders confirmed that overactive microglia might inhibit adult hippocampal neurogenesis [99–101]. For instance, microglia-mediated neuroinflammation can disrupt neurogenic niches and undermine the integrity of neuronal population replenishment programs [102]. Conversely, “*in vitro*”, microglia chronically activated by LPS with a secretory profile dominated by IL-10 and prostaglandin E2 (PGE2) are highly permissive to the neurogenic cascade [98]. The finding that microglia chronically activated with a definite secretory profile can support adult neurogenesis is consistent with the current theory in which A β plaques stimulate a chronic inflammatory reaction by microglia. In addition, LPS, IL-10 and PGE2 are implicated in AD [103–111].

Microglia has phagocytic activity

Of note, microglia serve a further function in adult neurogenesis. In young adult rats, approximately 9,000 new cells are generated in the SGZ each day, but nearly half do not survive [112, 113], similar to what occurs in the V-SVZ [113]. In particular, the majority of newborn neural progenitors undergo apoptosis 1–4 days after they are generated in the SGZ, and microglia phagocytose apoptotic debris from these cells to help maintain the equilibrium of the neurogenic niche [114]. Interestingly, this microglial phagocytic activity is apparently unchanged by aging or acute neuroinflammation, suggesting that it is a mechanism that promotes a homeostatic neurogenic niche in both healthy and disease states [114].

Microglia directs the migration of neuroblasts

Finally, microglia seem to have the capacity to direct the migration of neuroblasts [115]. Interestingly, some authors found in a transgenic mouse strain that depletion of microglia in the V-SVZ was linked to a marked reduction in neuroblasts reaching the OB with a concomitant accumulation of immature cells in the V-SVZ and RMS [116]. These findings suggest that microglia residing in the V-SVZ/RMS regions are critical for neuroblast

survival and migration to the OB, possibly as a consequence of their release of the cytokines IL-4, IL-6, and IL-10 [116]. It is also important to note that microglia-mediated phagocytosis of neuroblasts is a rare phenomenon along the V-SVZ/RMS migratory pathway, and accordingly, markers of activated microglia, such as TREM2 and CD68, were undetectable in these regions [116]. In contrast, within the OB layers, where interneurons are continuously replaced by V-SVZ-generated precursors, microglia exhibit overt and robust phagocytosis. Therefore, microglia in the neurogenic areas of the V-SVZ/RMS are unique and specialized to support neural precursor proliferation and migration across significant distances to their final destination [117–119]. Further data supporting the role of microglia in sustaining and driving the migration of neuroblasts come from research on brain injury. In this regard, an invariant feature of damage to the CNS is the migration of microglial cells to the site of injury and their subsequent activation [120, 121]. Interestingly, several studies have shown that precursor cells preferentially migrate to sites of inflammation in animal models of multiple sclerosis and that these new cells preferentially differentiate into oligodendrocytes [122–124]. In contrast, in experimental models of more acute damage with neuronal loss, precursor cells, both extrinsically provided and endogenous precursor cells, migrated to the damaged area and differentiated into neurons [125–130]. More recently, some authors reported that precursor cells migrate from the V-SVZ and RMS to the injured cortex after traumatic brain injury (TBI) in mice and that prokineticin 2 (PROK2), a chemokine important for OB neurogenesis, is expressed exclusively by cortical microglia in the cortex as early as 24 h after injury [131]. In addition, the same authors demonstrated “*in vitro*” that cells expressing PROK2 directionally attract V-SVZ cells [131].

The role of astrocytes

The role of astrocytes, in addition to that of microglia, is in line with current theory, considering that astrocytes are early involved in AD [132–134] and have strong pro-neurogenic activity [135, 136].

Astrocytes in AD

Ramon y Cajal noticed reactive hypertrophic astrocytes surrounding senile plaques and blood vessels with amyloid deposits in post-mortem AD patients already in 1913 [137]. This observation has been

replicated several times in AD patients' brains [138–141] and in AD mouse models [142–144]. Within the CNS, astrocytes play a key role in the protection and repair of neuronal damage [145, 146]. Astrocytes respond to inflammatory substances and undergo a process known as reactive astrogliosis [147, 148] in various pathological conditions, including acute injury and progressive disorders such as tumors and AD [147]. Reactive astrocytes release molecules such as cytokines, chemokines, growth factors and gliotransmitters [149]. Notably, astrocytes release factors that promote axon growth, which are essential for synaptic formation and maturation in response to an injury [148, 150]. Moreover, astrocytes increase neuronal viability and mitochondrial biogenesis, protecting neural cells from oxidative stress and inflammation induced by amyloid peptides [151]. At the same time, astrocytes may exert neuroprotection at different stages of AD. Indeed, both astrogliosis and microgliosis, in response to amyloid, increase glial secretion of transforming growth factor- β (TGF- β), which protects neurons from amyloid toxicity and increases amyloid clearance by microglia [152, 153]. Furthermore, astrocytes surrounding amyloid plaques show phagocytic activity and are able to phagocytize neuritic dystrophies both in mouse models and in AD patients' brains [154]. Indeed, astrocytes are part of the brain's glymphatic system, a clearance system for proteins and soluble solutes [133]. The astrocyte water channel aquaporin-4, expressed at the ends of astrocytes, facilitates this process and is important for A β clearance [155, 156] and probably also for tau clearance [133].

Astrocytes and adult neurogenesis

Astrocyte is the main cell type in the hippocampal niche of neurogenesis by number [135]. In the molecular layer of the dentate gyrus and in the SVZ [157], astrocytes are in close contact with NSCs and contribute to the regulation of almost all stages of adult neurogenesis, from the proliferation of NSCs to the functional integration of new neurons [135]. In particular, molecules secreted by astrocytes increase the proliferation of adult NSCs. For example, *in vitro* studies found that the adenosine 50-triphosphate (ATP) through P2Y1-PLC-phosphatidylinositol 3-kinase (PI3K) signaling [158], the N-methyl-D-aspartate receptor co-agonist (NMDAR) D-serine [159–161] and the fibroblast growth factor-2 (FGF2) act as factors in the proliferative induction of adult NSCs [162, 163]. In addition, several miRNAs expressed in astrocyte exosomes are

known to regulate adult neurogenesis [164]. In addition to affecting proliferation, molecules secreted by astrocytes can also modulate other stages of adult neurogenesis, such as the migration and differentiation of progenitors into neurons, or the maturation, synaptic integration, and survival of newborn neurons [135]. For example, the first *in vitro* study examining the role of astrocytes in adult neurogenesis in the hippocampus showed that astrocyte-conditioned cell culture medium increases the differentiation of NSCs into neurons [165]. Neuronal differentiation of adult NSCs is also promoted *in vitro* through juxtacrine signaling by astrocyte secretion of ephrin-B2 and activation of EphB4 receptors on the stem cell [166]. In addition, astrocyte-derived soluble factor thrombospondin-1 (TSP1) is known for its antiangiogenic activity and promotion of synaptogenesis during brain development [167]; it also increases adult NSC proliferation and neuronal differentiation *in vitro* [167]. Consistently, adult TSP1-deficient mice exhibit reduced proliferation of adult NSCs [168]. Another secretory factor, neurogenesis-1, increases the neuronal fate of newly formed hippocampal cells [169], while IL-1b and IL-6 promote neuronal differentiation of adult NSCs/progenitors *in vitro* [170]. Finally, D-serine released from astrocytes has been shown to control dendritic maturation and functional integration of newborn hippocampal neurons [171].

Detrimental effects of glial cells in AD

The current theory focuses mainly on the neuroprotective functions of microglia and astrocytes in AD and their proneurogenic actions. However, it is worth noting that both types of glial cells have been found to contribute to the damaging effects in AD, mainly through the promotion of innate immunity and pro-inflammation and influencing the permeability of the blood-brain barrier [172, 173]. Specifically, both microglia and astrocytes interact with A β , and A β in turn activates microglia and astrocytes through TLRs to release neuroinflammatory mediators that promote neurodegeneration [174, 175]. Pro-inflammatory cytokines decrease the phagocytic activity of microglia and probably transform microglia into pro-inflammatory phenotypes [173]. In addition, pro-inflammatory microglia increase tau phosphorylation and aggravate tau pathology [176]. At the same time, reactive astrocytes have been found to release excessive amounts of GABA and glutamate, resulting in memory impairment and synaptic

loss in an animal model of AD [177, 178]. Moreover, these cells contribute to the microcirculation dysregulation and blood-brain barrier disruption, which facilitates A β accumulation and disease progression [179, 180]. Finally, reactive astrocytes might even pave the way for the formation of early amyloid plaques [181]. Considering that AD has a long pre-clinical phase, this dual role of glial cells [182] is not incompatible with the current theory that easily explains the very early stages of the disease. Moreover, some aspects of neuroinflammation, even under chronic conditions, continue to promote microglial phagocytosis and A β containment, resulting in a neuroprotective function [183].

THE SCALING OF MOLECULAR PATHOLOGY TO THE MACROSCOPIC BRAIN

Braak staging model

According to Braak's neuropathological staging [184–187], pathological tau aggregates in AD develop first in nerve cells of brainstem nuclei (subcortical stages a–c) that have projections ending in the cerebral cortex [188–190]. It appears that from the locus coeruleus (LC) of the pontine tegmentum [191–196], the lesions progress to a distinct portion of the cerebral cortex, the transentorhinal cortex (TEC) [197]. In cortical projection neurons, the resultant and originally nonargyrophilic pretangle protein, during cortical stages 1a and 1b, becomes transformed into argyrophilic neurofibrillary lesions that characterize subsequent NFT stages I–VI [189]. The neurofibrillary pathology advances from the TEC (NFT stage I) into the OB [198], the entorhinal cortex (EC), and the hippocampal formation (NFT stage II). During NFT stage III, tau pathology progresses from the TEC to the laterally adjoining basal temporal neocortex, and during NFT stage IV, it extends more widely to the temporal, insular, and frontal neocortices. In NFT stage V, cases display severe involvement of most neocortical association areas, leaving only the primary fields mildly involved or intact. In the end stage, NFT stage VI, even these areas become involved. The production of abnormal tau continues from the outset until the final stage of the pathological process [188, 189, 199]. In summary, in AD, the pathology progresses anterogradely from distinct predilection sites in the lower brainstem to distant but connected regions of the cerebral cortex, and it does so sequentially with little interindividual variation, albeit at

different rates [189]. Considering the mechanism implicated in tau spreading, a great deal of data suggest that transcellular propagation of tau aggregates, or seeds, could underlie disease progression [200–207].

The prion-like seeding and spreading hypothesis of tau

According to the prion-like hypothesis, pathological tau can distribute from one cell to another, thus propagating pathology from affected brain areas to interconnected healthy areas, involving mechanisms similar to those of prion diseases [208]. This hypothesis could explain the hierarchical pathway of neurodegeneration described in Braak's scheme [184]. The prion-like hypothesis involves two main stages, namely the seeding, that is the ability of abnormal tau to convert normal tau into a pathological form, and the propagation, that is the spread of pathological tau to connected neurons [209].

Abnormal tau has seeding capacity

Several studies support a seeding capacity of tau similar to that of prions [209]. In this regard, some authors showed that injection of tau aggregates extracted from mice overexpressing mutated tau (P301S) into mice overexpressing wild-type human tau is sufficient to induce tau pathology [201]. In particular, when a tau immunodepleted extract was injected, no pathology was detected, demonstrating that tau is the responsible factor of aggregation, as later confirmed by other research groups [210–212]. Most *in vitro* studies showed that incubated aggregates/seeds are internalized by endocytosis and promote aggregation of overexpressed tau in cell lines [203, 204, 210, 213–221]. Evidence that tau aggregates have prion-like seeding behavior come mostly from experimental models [222]. However, there is also evidence for seeding activity in tau aggregates derived from patients with tauopathy. Indeed, sarkosyl-insoluble PHFs extracted from AD brain tissue induce seeding in cultured cells and wild-type mice [223, 224]. In addition, in brain homogenates and cerebrospinal fluid (CSF) from AD cases, tau seeds have been found to induce non-aggregated tau aggregation in FRET-based biosensor assays, particularly in regions known to be devoid of phospho-tau deposits [220, 225]. Moreover, recent work showed that human CSF from AD patients can induce tau seeding in experimental models [226].

Seeded tau aggregation is templated

Other studies demonstrated that the seeded tau aggregation is templated [222]. Some authors, for example, observed that native P301S tau seeds derived from transgenic mice brains confer their highest seeding competence to less competent recombinant P301S tau seeds when co-incubated with them *in vitro* [227]. Under the light microscope, tau aggregates induced in cells or *in vivo* have the same morphological appearance as the parent tau seed, suggesting a templated conversion mechanism [222]. This has been demonstrated in many studies from Diamond's laboratory, in which the formation of morphologically distinct tau seeds resembles the parent tau seed both in cell culture [228] and, more recently, *in vivo* [225]. That a templated conversion mechanism may be relevant to tauopathies in humans has been demonstrated by studies from the laboratories of Goedert and Tolnay, in which injection of brain homogenates of different tauopathies into the brains of mice expressing unaggregated human tau resulted in the formation of only the inclusions of the corresponding tauopathy [229].

Neuroanatomical spread of tau aggregates

Trans-synaptic propagation of pathological tau has been demonstrated using a number of different approaches in transgenic mice [222]. Some authors showed not only the induction of tau aggregation in rodent brains following intracerebral injection of brain homogenates containing tau seeds, but also the time-dependent appearance of tau pathology in anatomically connected brain regions [201]. Others reported the appearance of pathological tau in areas connected to sites injected with tau seeds or tau-expressing viral vectors [230–233]. Some authors used a model in which human tau expression was restricted to the entorhinal cortex alone, showing that the tau pathology was evident in anatomically connected regions that did not express the human tau transgene [202, 205]. Further studies in tau transgenic mice indicated that tau seeds predict disease spread by appearing in brain regions before the occurrence of any other pathological changes [215]. Interestingly, this finding explains the histopathological observation made by some authors more than 20 years ago [234]. These authors reported the absence of pathological tau in a frontal cortical region that was anatomically disconnected from the limbic region following neurosurgery, decades before the patient developed AD. Conversely, the authors found an extensive tau pathology in the immedi-

ately adjacent brain regions and limbic and isocortical areas [234].

Propagation involves several steps

The propagation of pathological tau to connected neurons consists of at least four steps [235]. First, tau must be secreted or released from donor neurons; second, it must undergo aggregation before or after being released; third, tau must be taken up by recipient neurons; and fourth, tau aggregation must be induced in recipient cells [236].

Currently, there is evidence that misfolded tau is indeed secreted [209, 212, 237]. However, the nature of secreted tau is debated in the literature [222, 238]. Tau is secreted mainly in free form [239–242], but it is also found within nanotubes [243, 244] or associated with extracellular vesicles (EVs) [245], such as exosomes [242, 246, 247] and ectosomes [239]. While nanotubes may be difficult to visualize in the human brain, phospho-tau-containing EVs have been found not only in the brains of transgenic mice [248, 249], but also in peripheral fluids (CSFs) [242, 250] and blood [251–253] of AD patients [209].

Considering the uptake step of tau by an adjacent recipient cell, *in vitro* studies showed that extracellular aggregates of tau can be internalized by naïve cells by promoting fibrillation of intracellular tau [203, 204, 254]. Tau pathology can be transferred between co-cultured cells [203, 204, 254] and also through synaptic contacts between neurons that facilitate the propagation of pathology [255]. Intracranial or peripheral administration of pathological tau [222, 256] and *in vitro* experiments have shown that tau is mainly internalized by active endocytic processes [203, 257]. In particular, three types of endocytosis have been described: bulk-endocytosis, actin-dependent macropinocytosis mediated by HSPGs on the cell surface, and clathrin-mediated endocytosis [208].

Once internalized, tau can escape endosomal vesicles by inducing their rupture [258, 259] and accumulate in the cytoplasm where it becomes a potential template for tau misfolding [208]. Indeed, pathogenic misfolded tau proteins act as “seeds” that recruit soluble endogenous tau into larger aberrant conformations [260] that slowly propagate into interconnected brain regions, as demonstrated in various animal models [235]. Although the biochemical mechanisms that drive the conversion of normal tau to the pathological form remain unclear, several models of tau seeding have been proposed [261, 262]. Finally, transcellular transfer of tau aggregates between seri-

ally cultured cells in microfluidic chambers was demonstrated [263]. In addition, diffusion of tau from neuron to neuron through trans-synaptic connections via exosomes has been reported to seed aggregates [242, 249]. However, other mechanisms that do not require secretion but a direct connection between cytoplasm might be involved [222]. Indeed, a recent work showed that nanotubes promote the interneuronal transfer of tau fibrils into neurons [243, 244].

Limitations of the seeding and spreading hypothesis

Although several pieces of evidence seem to support the seeding and spreading hypothesis of tau, many points still remain to be clarified [68, 264, 265]. Firstly, some authors pointed out that the methods used and data collected in some studies supporting the hypothesis are not all without some limitations [68, 265]. Secondly, the biochemical mechanism that drives the conversion of normal tau to the pathological form is still not clear [208]. Thirdly, the exact nature of the tau seeds responsible for the propagation of tau pathology remains controversial [222]. Furthermore, the specific pathways and mechanisms underlying the spread of pathological tau, including the mechanism of releasing from donor neurons and subsequent uptake by recipient neurons in AD, remain unclear [209, 235, 266]. In addition, the molecular forms of extracellular tau are not fully understood, and the physiological or pathological functions of this extracellular tau remain unknown [266]. Further investigations are then needed to clarify the relationship between the propagation of tau aggregates and tau-induced toxicity and degeneration [222]. Furthermore, it cannot be ruled out that genetic variants identified as risk factors for tauopathies play a role in the propagation of tau pathology, but many more studies are needed to document this [222]. Finally, the contribution of selective vulnerability of neuronal populations as an alternative explanation of the spread of tau pathology needs to be clarified [222].

Microglia and astrocytes could be involved in the spread of pathological tau

It is worth noting that some studies highlighted the involvement of microglia in the spread of pathological tau [208]. Indeed, it has been reported that increased microglial activation accelerates the propagation of tau in the brain [267]. Furthermore, microglia were found to promote tau propagation [246, 268], as supported by the marked reduction in tau propagation through microglia depletion in two

independent models of tauopathy [246]. The mechanism involved in the promotion of tau propagation by microglia has not been fully elucidated. However, tau was found in the EVs in the CSF of individuals with AD [239], and microglia were found to internalize tau seeds and degrade them [182]. When microglia fail to degrade these tau seeds, deleterious consequences occur, including the secretion of tau-containing exosomes that can spread to neurons [182].

Interestingly, tau was also found in astrocytes of individuals with AD [269]. Although tau was found in glial cells [270], astrocytes do not express this protein under physiological conditions [271], and the origin of tau in astrocytes in AD is still unclear [272]. One unproven possibility is that AD progression induces the translation of tau from the mRNA present in astrocytes [273]. Alternatively, astrocytes could also capture extracellular tau [228, 274, 275]. In this regard, astrocytes have specific heparin sulfate proteoglycans (HSPGs) and receptors, such as low-density lipoprotein receptor-related protein 1 (LDR1), that can mediate the uptake of tau aggregates [133]. Aggregates can be internalized and processed by various mechanisms, including lysosomal degradation. Disruption of aquaporin-4 in perivascular astrocyte ends may contribute to the disruption of tau clearance and accumulation of tau aggregates in the CNS [133]. However, the microglial and especially astrocytic mechanisms that may contribute to pathological tau seeding are not yet fully understood [133]. As for microglia, it is sufficient to mention that they can also reduce the seeding activity of tau [268, 276–278], supporting the idea that microglia are indeed able to limit or promote the spread of tau [182]. Considering astrocytes, one hypothesis is that tau pathology spreads from one astrocyte to another, possibly through astrocyte gap junction networks and tunneling nanotubes across brain regions [266, 279]. Astrocyte engulfment of tau-containing synapses may be another pathway by which astrocytes contribute to the spread of tau in AD [133].

The possible involvement of glial cells in tau spreading is consistent with the current theory. In this case, microglia and astrocytes activated by A β deposition would not only promote tau pathogenesis through their proneurogenic effect, but also contribute to the spread of tau aggregates in the brain.

Open questions left in the Braak model

Despite the indubitable value of the Braak staging model, some open questions remain.

First, the view that nonthalamic nuclei would be the first site of tau pathology has been questioned [280]. Indeed, these nuclei are equipped with a type of termination (i.e., nonjunctional varicosities) [191, 196, 281, 282], supporting a diffusive mode of transmission [191, 196, 281, 283–285] that is not suitable for neuron-to-neuron transmission of abnormal tau as provided by the model. Indeed, recent findings have suggested that TEC/EC are actually the first site that develops early tau pathology [286]. In particular, tau seeding activity that precedes detectable NFTs was found in the LC only after it was already prominent in the TEC/EC, i.e., at later NFT stages (IV–VI), suggesting the idea that tau seeds spread from the TEC/EC to the LC and then to more distant cortical regions [286].

Second, to date, a clear explanation of why tau pathology begins in TEC/EC seems lacking. Indeed, it is not easy to contextualize this finding in the frame of classical amyloid theory, considering that amyloid plaques first appear in the association cortices of the temporal lobe, at some distance from the TEC/EC where damaged neurons containing NFTs are first found [187, 287]. Additionally, further studies examining the early degeneration of the lateral EC have reported that levels of amyloid peptides in this region are not higher than those in other, less affected regions [288, 289].

Third, despite the evidence that early tau pathology emerges in entorhinal layer II cells at Braak stage II [186] and that these cells project to the DG by the perforant path [290–294], the DG is not affected by tau pathology at early stages. Indeed, granule cells of the DG remain uninvolved in Braak stage III, and some tau pathology emerges only at stage IV [186]. At the same time, hippocampal CA1 cells receiving projections from entorhinal layer III cells [292, 293] are impacted far earlier, starting at Braak stage II [186].

Finally, a clear and accepted explanation of the peculiar regional distribution of tau lesions and subsequent neurodegeneration in EOAD, especially in the syndromic variants of AD, is lacking in the Braak model.

A new model of tau spreading in the medial temporal lobe

The current theory provides a new model of tau spreading in the medial temporal lobe (MTL) (Fig. 1). According to the new model, tau pathology begins in NSCs within the niches of adult neurogenesis. In particular, in LOAD, the initial tau pathology,

likely in the form of soluble aggregates of misfolded and hyperphosphorylated but nonfibrillar tau protein [280], originates in the SGZ in the hippocampal DG. From this site, seeds of tau pathology spread retrogradely to the EC through the perforant path. From here on out, anterograde transmission flanks retrograde transmission. Therefore, by virtue of reciprocal connections between the EC and TEC [294–297], the TEC receives tau seeds both anterogradely and retrogradely. In this regard, it is interesting to note that the EC projections to the TEC are input to layer II [296, 297], where NFTs are first found [185, 298]. Then, from EC/TEC, the pattern of distribution of tau pathology follows the Braak model. However, a further difference may occur. Indeed, according to the current model, it cannot be excluded that some further foci of degeneration could start locally in some regions of the cortex, provoked by neuroblasts carrying the seeds of tau pathology arising from the V-SVZ niche. In this case, in LOAD, the primary regional distribution pattern of tau pathology in the MTL would be complicated by concurrent foci of pathology that emerge locally in neocortical regions. In the same context, it is noteworthy that the OB is the arrival point of migrating neuroblasts from the V-SVZ along the RMS, and at the same time, many findings support early involvement of this region in AD [198, 299–301]. However, olfactory structures, including the OB, anterior olfactory nucleus, and piriform cortex, send projections to the superficial layers of EC [302, 303]. Therefore, retrograde transmission of tau pathology from the EC to the OB could be a more parsimonious explanation. The current model seems plausible and coherent with some findings in the literature. In this regard, it is worth noting that retrograde transmission has been found to be possible. Indeed, projection neurons generate long axons to transmit information from one site to another, and for this purpose, their axons have mechanisms for both anterograde and retrograde transport of various cargos [222, 280]. Moreover, the idea of retrograde transmission of tau pathology in the MTL in AD has already been suggested [304]. In addition, the pattern of connections among the DG, EC and TEC is compatible with the new model of both retrograde and anterograde transmission. In particular, the DG receives projections from entorhinal layer II cells, where tau pathology is found early in EC [186, 305]. Moreover, projections from the EC to the perirhinal cortex that includes the TEC terminate most heavily in and around layer II, where tau pathology is first found in the TEC [184, 298].

Possible solution to the questions left in Braak staging

Interestingly, the current model seems to offer possible explanations for the open questions left in Braak staging.

First, it is coherent with the recent view considering the LC and other nonthalamic nuclei as not the primary sites for tau spreading in the MTL. At the same time, the core hypothesis of tau pathogenesis as linked to adult neurogenesis and migration offers a speculative explanation for the emergence of tau pathology in these sites. In fact, the LC is highly connected to the hypothalamus, and constitutive neurogenesis in the adult hypothalamus of mammals, including rodents, rats, mice, voles [306–314], and sheep [315], has been documented [316].

Second, according to the current model, tau pathology emerges first in the TEC because this region receives the massive seeds of tau pathology from the EC, both anterogradely and retrogradely, through the multiple reciprocal connections between the two regions. Conversely, the EC receives the seeds of tau pathology from the DG only by retrograde transmission (Fig. 1, Stage 1). Therefore, in the current model, NFTs emerge first in TEC because the initial load of tau pathology would be greater than in the EC (Fig. 1, Stage 2).

Third, the absence of tau pathology in the DG at early Braak stages is due to the strong activity of clearance usually occurring in the neurogenic niches [74, 75]. According to this view, although seeds of tau pathology originate in NSCs in the SGZ niche and spread retrogradely toward the EC by a perforant path, the formation and accumulation of NFTs is suppressed or at least delayed in NSCs in the DG (Fig. 1, Stage 2). For the same reason, CA3 [118], which receives excitatory outputs from the DG, is not impacted by tau pathology at early stages. Instead, tau pathology emerges first in CA3 at Braak stages III–IV [186]. In the same context, it is relevant to note that adult-generated neurons in the SGZ receive local connections from multiple types of GABAergic interneurons [317], whose inputs to the niche are fundamental for maintaining a healthy level of neurogenesis under normal conditions [318, 319]. Interestingly, these same GABAergic interneurons have been shown to be particularly vulnerable to AD pathologies, such as NFTs of phosphorylated tau protein [317, 320–323]. Therefore, GABAergic interneurons could be plausible candidates to convey transmission of the first tau seeds originating in SGZ

NSCs. Strictly related to the previous point, the current model seems to offer a plausible explanation for why tau pathology spreads early from the EC overall to the hippocampal CA1 region, while it seems not to target the DG, despite that the EC and the DG are highly connected by the perforant path. Damage to the perforant path between the lateral EC and DG occurs unusually early in AD [324]. The long axons of projection neurons are in fact not well equipped to degrade or eliminate pathological proteinaceous aggregates [189, 325]. Based on the current model, the seeds of tau pathology, after which the EC, and more so the TEC, have been impacted, would spread anterogradely from the EC to multiple regions, as in the classical Braak model. However, at this point, the current view predicts that the DG would be primarily disconnected from the EC because the entorhinal perforant projections toward the DG would already be deteriorated due to the precedent retrograde transmission of tau pathology along the same projections in the opposite direction, from the DG to the EC, during the first stage of disease. Consequently, the connections between the EC and CA1 (and subiculum) are unique undamaged fibers in the perforant path available for tau seed transmission at this stage (Fig. 1, Stage 3).

Finally, the current model seems to offer a plausible explanation for the peculiar distribution of tau pathology in EOAD. Individuals with EOAD may present with striking neurobehavioral phenotypes, reflecting damage to the language systems [326], visual systems [327], or frontal-executive systems [328]. In general, EOAD is more likely to present with atypical clinical phenotypes than LOAD patients [329]. In one study, approximately 25% of EOAD patients presented with a nonamnestic phenotype in whom visual or apraxic and language phenotypes predominated [330]. In another study, almost 60–70% of EOAD patients exhibited atypical patterns of brain atrophy [329]. Interestingly, the focus and system-specific neurobehavioral features in EOAD variants do not reflect regional accentuation of A β , but they do show strong correlations with the pattern of glucose hypometabolism and atrophy [331–334]. More generally, despite their differences, in autosomal dominant AD (ADAD), EOAD and LOAD, the distribution of A β deposition throughout the brain is similar (with the exception of A β deposition in the striatum in ADAD), affecting large confluent areas of the association cortex and overlapping with a set of brain regions active at rest [335–339]. Therefore, phenotypic heterogeneity in

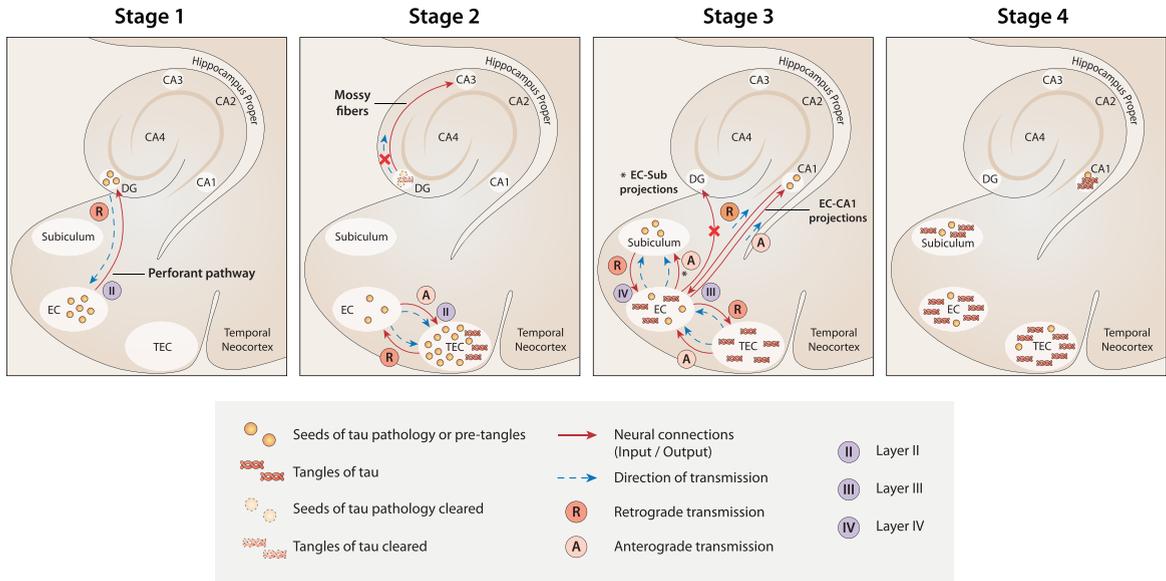


Fig. 1. A new model of the spread of tau pathology in the MTL according to current theory. Stage 1: The first seeds of tau pathology develop in NSCs in the SGZ niche of the DG. Then, they spread from the DG to the EC by retrograde transmission along the connections of the perforant pathway. Stage 2: Seeds of tau pathology spread from the EC to the TEC by both anterograde and retrograde transmission along the multiple connections between the two regions. Because of the massive load of tau pathology accumulated in the TEC, tangles develop here first. At the same time, tangle formation is suppressed (or delayed) in the DG because of the strong clearance activity that usually occurs in the neurogenesis niches. As a result, the transmission of tau pathology from the DG to CA3 is nullified (red cross). Stage 3: Seeds of tau pathology spread from the EC to the CA1 and subiculum along the EC-CA1 and EC-subiculum projections, respectively. In contrast, the seeds of tau pathology would not spread anterogradely to the DG (red cross), because the neuronal connections between the EC and DG are already deteriorated at this stage due to the transmission of tau pathology in the reverse (retrograde) direction in the previous stage (Stage 1). Stage 4. When tau tangles emerge in the CA1 and subiculum (starting from Braak stage II), the DG and CA3 are not yet affected. Figure 1 was produced by Antonio Garcia, scientific illustrator from Bio-Graphics.

AD is not easy to explain considering both the frame of the classical amyloid theory and the primary pattern of regional distribution of tau pathology starting from the MTL according to Braak staging. In the current model, EOAD and LOAD exhibit different regional distributions of tau pathology and subsequent degeneration because the V-SVZ niche is primarily active in EOAD, while the SGZ niche is primarily active in LOAD. Accordingly, EOAD especially impacts regions on the dorsal cortex, whereas LOAD impacts the MTL [340] (Fig. 2). Moreover, activated microglia surrounding $A\beta$ plaques release chemokines that attract and drive migrating neuroblasts toward the regions of $A\beta$ deposition, similar to what happens in brain injury [125–131]. Consequently, especially in EOAD, migrating neuroblasts deviate from the RMS to the OB and take different paths toward various regions of the cortex, carrying the seeds of tau pathology to those locations. In summary, the redirection of migrating neuroblasts to multiple possible destinations in the cortex is the basis of heterogeneity in the regional distribution of tau pathology and subsequent degeneration

in atypical EOAD syndromes (Fig. 2). In addition, as already reported, long-distance migration throughout the inhibitory environment of the adult brain could contribute to augmenting tau phosphorylation.

Interestingly, recent *in vivo* tau-PET imaging studies in AD have revealed substantial heterogeneity in tau deposition patterns with significant deviations from Braak's scheme [334, 341, 342]. These findings are in line with the four subtypes previously identified from neuropathology and neuroimaging studies based on the distribution of NFTs and patterns of brain atrophy, respectively: hippocampal-sparing AD, limbic-predominant AD, typical AD, and minimal atrophy AD [343–354]. In addition, some studies confirmed atypical patterns of tau deposition with elevated tau-PET signal in the occipital and parietal cortex [355], left temporo-parietal areas (logopenic) [356] and, similarly, perirolandic areas (corticobasal syndrome due to AD) [357, 358] reflecting the clinical variants most frequently associated with EOAD [359, 360]. Interestingly, some of these tau-PET imaging studies in AD found that pathological tau accumulates in the associative cortex, completely

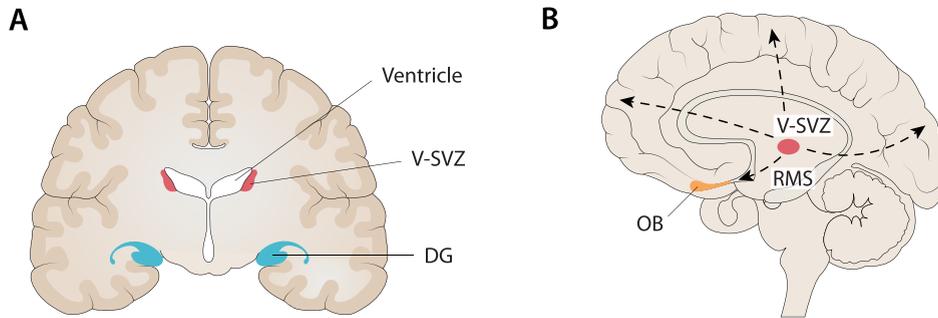


Fig. 2. Compatibility between the localization of the main niches of adult neurogenesis and the core regions targeted in AD. A) One of the main niches of adult neurogenesis is the sub-granular zone (SGZ) of the dentate gyrus (DG) in the hippocampus. At the same time, the hippocampus is the first major region targeted in AD, especially when late-onset AD (LOAD) is considered. B) Another main niche in adult neurogenesis is the ventricular subventricular zone (V-SVZ) along the lateral ventricles. From this niche, through several long migrations to the cortex (dashed lines), it is possible to reach every cortical region (e.g., frontal, fronto-parietal, occipital) that is targeted by AD, especially when considering early-onset AD (EOAD) and syndromic variants of AD. In addition, it is noteworthy that the olfactory bulb (OB) is the end point of neuroblasts migrating from the V-SVZ along the rostral migratory stream (RMS) and, at the same time, many findings support an early involvement of this region in AD. Figure 2 was produced by Antonio Garcia, scientific illustrator from Bio-Graphics.

sparing the hippocampus [342, 350, 359, 361, 362]. This finding strongly supports the idea of distinct foci of early tau deposition and multiple pathways of tau diffusion in AD, including cases without any involvement of the hippocampus and/or entorhinal cortex [342, 348].

This scenario does not fit well with Braak's staging system and seems more consistent with the current theory that predicts different niches of adult neurogenesis and multiple pathways of migration to the cortex.

Cortical arealization in development and AD

Considering that migration paths throughout the cortex, including the RMS, are mostly quiescent in the human adult cortex, the current model would be plausible only provided that a strong pro-neurogenic action would foster neurogenesis and support highly demanding migrations. In this respect, the accumulation of A β deposits and consequent microglial activation are key factors. However, the distribution of A β throughout the brain is diffuse and similar in LOAD, EOAD, and FAD, as well as in AD variants. Therefore, it is not plausible that microglia surrounding A β plaques signal a precise direction to migrating neuroblasts, similar to focal brain insults, such as stroke. Furthermore, the current model cannot explain why only some directions of migration are undertaken—those corresponding to the paths toward the regions impacted in well-known AD variants, e.g., posterior, frontal, and left perisylvian—and not others. At this point, I speculate that not only should a further source of signals drive migration through-

out the cortex in EOAD but also that this source should contain information about brain topography, likely at the macroscopic level of hemispheres, lobes, and gyres, to efficiently work. Surprisingly, I found that the program under cortical arealization in development perfectly fits this idea. In particular, there is a complex mechanism regulating the progressive patterning and correct localization of brain areas during development [363], which necessarily uses some spatial information related to brain topography to work. This mechanism would be mostly, even if not exclusively, under the genetic control of factors with discrete expression in the cortical field (protomap models). Moreover, some findings have suggested that the main spatial information used is related to simple brain axes. In particular, animal studies have demonstrated that there is an anterior-posterior (A-P) gradient of gene expression of morphogens or transcription factors, such that specific genetic factors enlarge rostral (motor) areas at the expense of caudal (sensory) areas, and vice versa [363]. In addition to this A-P gradient, there is evidence for graded expression patterns along with other distributions, including the medial-lateral (M-L) and dorsal-ventral (D-V) axes.

The failure of certain processes (e.g., cell proliferation, migration, and abnormal organization) during cortex development has been associated with several cortical malformations [364]. What is interesting is that most malformations do not involve the entire cortex uniformly but have regions of maximum severity. For example, some malformations (schizencephaly, megalencephaly) may alternately involve one or both hemispheres. Another type of malformation (e.g.,

lissencephaly) may have two forms, one with maximum severity in the frontal lobes and the other with maximum severity in the occipital lobes [364]. Another more diverse malformation (i.e., polymicrogyria) shows a highly heterogeneous topographic distribution (e.g., frontal, frontoparietal, perisylvian, parasagittal parieto-occipital, parietal, generalized), with a predilection for the perisylvian cortex [365]. As might be expected considering that the malformations are due to the failure of certain processes during cortical development, by observing the distribution over the cortex of some of these developmental malformations, we can easily recognize the structure of the A-P, D-V, and M-L axes underlying the cortical arealization process (Fig. 3). Interestingly, the regions targeted by degeneration during early stages in AD, considering LOAD, EOAD, FAD and all the syndromic variants together, seem to be arranged at opposite locations along the same A-P, D-V, and M-L brain axes [366] (Fig. 3). In other words, AD (and more specifically EOAD) and the program of cortical arealization in development seem to use the same alphabet of spatial information on brain topography (Fig. 3).

In summary, I speculate that in EOAD, when neuroblasts leave the V-SVZ niche and start migrating, the signals from microglia activated by A β deposition provoke path redirection from the RMS and, at the same time, sustain long-distance migration. However, reactivation of the genetic program of arealization during development would contribute to signaling the direction for migrating neuroblasts to follow.

DISCUSSION

Adult neurogenesis in brain injury and AD

The study of adult neurogenesis and migration in brain injury, keeping in mind the peculiarity of A β deposition compared to other types of injury, seems to support the notions of increased neurogenesis, promotion, and redirection of neuroblast migration, as well as reactivation of quiescent paths, recognized in the current theory. Indeed, in rodents, various pathological changes and injuries, e.g., ischemia or TBI, stimulate neurogenesis in the V-SVZ [367–369] and in the DG [370–374]. In addition, in the injured adult brain, neuroblasts generated in the V-SVZ migrate toward the site of injury [125, 126, 375–377], driven by various guidance cues, such as chemoattractants secreted by injury-activated astrocytes, microglia, and vascular endothelial cells in the injured area [73,

378]. Accordingly, multiple studies have shown significant intensification of neuroblast migration [125, 126, 367, 379] under these conditions. Therefore, the migratory paths from the V-SVZ, which are largely quiescent in the adult brain [380], could be reactivated in response to injury [367].

Over the years epileptic seizures, as well as stroke and TBI, were demonstrated to provoke functional alterations in the hippocampal neurogenic cascade that were characterized under the umbrella term “aberrant neurogenesis” [381, 382]. In particular, aberrant neurogenesis encompasses multiple (dys)functional outcomes, including excessive activation of NSCs [383, 384], alterations in NSC fate [383, 385] with a shift from neurogenesis to astrogenesis, downregulation of the proliferative capacity of NSCs, neural progenitor cells or neuroblasts [385–387], abnormal development and length of the dendritic tree of newborn neurons [388, 389], and ectopic migration of newborn neurons [390]. Interestingly, some authors, both in postmortem AD patients and in a transgenic (3xTg) AD mouse model, found that hyperphosphorylated tau, especially when expressed in GABAergic interneurons in the DG, was related to multiple alterations in SGZ NSCs strictly resembling the cardinal features of aberrant neurogenesis [317]. This finding suggests that tau-mediated aberrant neurogenesis also occurs in AD.

The link between AD and adult neurogenesis

A link between AD and adult neurogenesis has been recognized for some time. Indeed, AD and adult neurogenesis are not only linked by common sites where early pathology occurs and newly born neurons integrate in the preexisting circuitry (e.g., MTL, OB) but also share a number of common molecules in both processes [391–395]. In particular, molecular players in AD, including apolipoprotein E (ApoE), APP, and presenilin 1 (PS1), as well as their metabolites, play a role in adult neurogenesis [392, 394]. Further critical signals in AD have been found to regulate neurogenesis, such tau [38, 317, 396], Notch1 [384, 397], cyclic AMP response element binding protein (CREB) [398–406], and Wnt/ β -catenin [407–410]. Furthermore, some authors have shown that blocking adult hippocampal neurogenesis in an AD mouse model exacerbated neuronal loss and cognitive impairment, while inducing adult hippocampal neurogenesis together with brain-derived neurotrophic factor improved cognition in AD mice [411]. Another study observed markers of increased

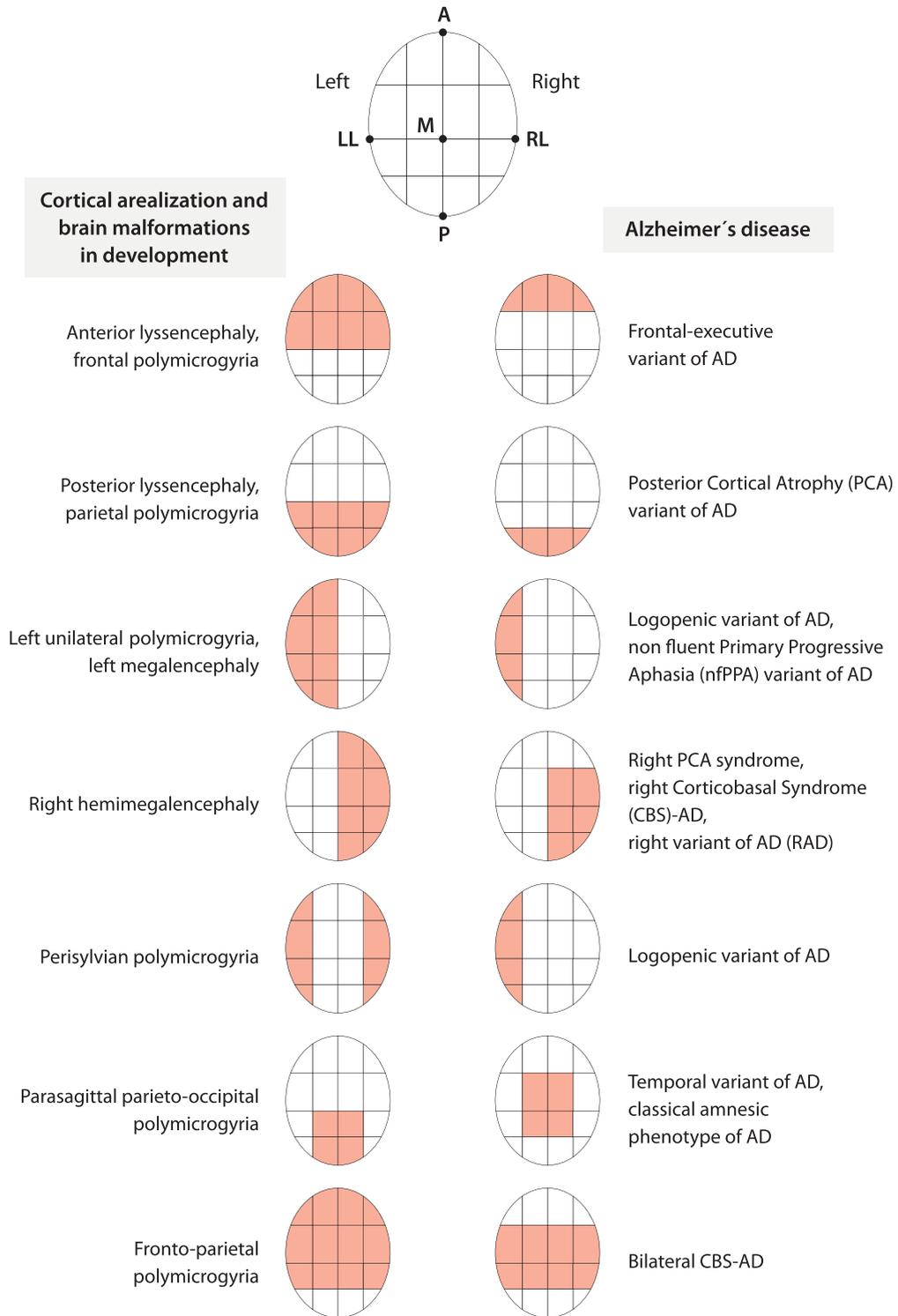


Fig. 3. Cortical arealization in development and AD appear to share the same alphabet of spatial information about brain topography. The coarse distribution over the cortex of brain malformations due to the failure of the arealization program during development and the key regions targeted by AD, considering all phenotypes, seem to follow the same few topographical instructions related to anterior-posterior, medial-lateral, dorsal-ventral brain axes and a simple left-right hemisphere specification. Figure 3 was produced by Antonio Garcia, scientific illustrator from Bio-Graphics. This is a modified version of Fig. 1 in Abbate (2018) [366].

neurogenesis in the DG of rare “resilient” individuals who remained cognitively intact, despite the presence of neuropathological features associated with AD, compared to AD and mild cognitive impairment patients [412]. Therefore, the prevailing view in the field is that impaired neurogenesis is a key contributing factor to AD pathology-driven neuronal dysfunction [394, 413–415]. Actually, the study of adult neurogenesis in postmortem AD patients and AD animal models has yielded conflicting results, frequently reporting a decrease [392, 416–420], but sometimes also an increase [392, 420, 421], in adult neurogenesis. The current theory predicts a complex relationship between the rate of neurogenesis and AD, depending on disease stage. In the first stage, there is a long-lasting tonic phase of reactive neurogenesis promoted by activated microglia triggered by A β deposition. Accordingly, the rate of neurogenesis is augmented, and tau pathogenesis is amplified. Then, the accumulation of tau aggregates in the niches start to have detrimental effects, likely ultimately reducing the rate of neurogenesis. This represents a phase of aberrant neurogenesis in AD that is most likely tau-mediated. A further phase could start when degeneration first occurs in the brain. Indeed, similar to what happens in stroke or TBI, cell death in the injured area stimulates the release of signals that could provoke a second cycle of aberrant neurogenesis in AD. The contrasting results found in the study of neurogenesis in AD seem to reflect this suspected complex relationship.

Adult neurogenesis, AD, and primary age-related tauopathy

The relative independence between the two processes driving amyloid and tau pathology in the new paradigm allows us to consider tau pathogenesis without A β deposition. Consequently, the finding of tau pathology uncoupled from amyloid pathology, as found, for example, in primary age-related tauopathy (PART) [422], seems compatible with the current paradigm. Moreover, the current model seems to fit well in explaining tau pathogenesis in PART. Indeed, the topography of tau lesions in PART is consistent with a possible origin in the NSCs within the SGZ niche. In particular, NFT changes in PART are usually restricted to the MTL and adjacent regions [132, 422, 423]. Later age of onset [422, 424–427], as well as limited spreading of tau pathology outside the MTL in PART compared to AD, would be due to the lack of a promoting effect on both neurogen-

esis and tau spreading by microglia in the absence of A β deposition. Therefore, the core hypothesis of tau pathogenesis as linked to adult neurogenesis and migration seems capable of combining AD and PART in a unique scenario. Some significant similarities identified between the two diseases support this idea. For example, NFTs in both disorders are identical, sharing both 3 repeat and 4 repeat tau isoforms and a 22–25 nm paired helical filamentous ultrastructure [422, 424, 428]. Moreover, phosphorylated tau lesions have the same topographic distribution in both PART and early AD [184, 424, 429]. In more detail, it has been reported that neurons in layer II of the TEC, a crucial region of early involvement in AD [184, 298], are also affected by neurofibrillary degeneration in PART [184, 298].

Adult neurogenesis, AD, and chronic traumatic encephalopathy

Unexpectedly, some findings from the study of a different disease, chronic traumatic encephalopathy (CTE) [430], seem to support the core hypothesis of tau pathogenesis as linked to adult neurogenesis and migration. CTE is a progressive tauopathy with distinctive clinical and pathological features that occurs after repetitive mild TBIs [430]. Microscopically, CTE is characterized primarily by NFTs and astrocytic tangles, with a relative absence of A β peptide deposits [431–447]. The evidence suggests that CTE begins focally, usually perivascularly, especially around small cerebral vessels, and at the depths of the sulci in the cerebral cortex [430, 446, 448–450]. Interestingly, some data have shown that the perivascular regions and the depths of the cerebral sulci are the most stressed regions, when the brain is subjected to rapid acceleration, deceleration, or rotational forces, such as occurs in mild TBI [451]. Thus, the highest concentration of phosphorylated tau correlates to the highest areas of stress in CTE [451]. Subsequently, pathological NFTs spread from these areas to adjacent superficial cortical layers. Considering the findings of an increase in the generation of new neurons in the niches [368, 452] and redirection of neuroblast migration to the injury site after TBI [73], V-SVZ neuroblast migration is ultimately redirected to the perivascular regions and the depths of the cerebral sulci in CTE because these regions are the primary sites of injury. Therefore, the current model of tau pathogenesis based on adult neurogenesis and migration in a special case of brain injury fits well for explaining the peculiar sites of

pathological tau deposition in CTE. Indeed, the distinct feature of prominent periventricular NFTs in CTE is in agreement with the location of the V-SVZ niche. Therefore, this model also seems to combine neurodegeneration (AD) with traumatic degenerative dementia (CTE) in a unique scenario. The fact that the isoform profile and phosphorylation state of CTE are very similar to those in AD [285, 308, 309] agrees with this idea. In particular, neuronal tau pathology in CTE shows immunoreactivity to both 3R and 4R tau, as in AD [430, 453, 454]. In addition, tau in both AD and CTE is phosphorylated at the same amino acids, including tau phosphorylated at threonine 231, and all six isoforms are present, leading to the observation that NFTs associated with AD are indistinguishable from those that occur in TBI [453].

Adult neurogenesis, AD, and the antimicrobial protection hypothesis

The current theory does not focus on the process that drives the initial deposition of A β . Consequently, it is compatible with a recent etiologic model of AD, namely the Antimicrobial Protection hypothesis, which views A β deposition in a new light compared to the classic amyloid hypothesis [455]. In this model, A β deposition is an innate immune response that normally protects against genuine, or misperceived, microbial infection in the brain. A β first traps and neutralizes invading pathogens in A β . Fibrillation of A β stimulates neuroinflammatory pathways that help fight infection and clear A β /pathogen deposits. In AD, chronic activation of this pathway leads to sustained inflammation and neurodegeneration. This new model is supported by several lines of evidence.

Indeed, many studies documented the presence of abnormal levels of pathogens in the AD brain, including viral, bacterial, and fungal infections [456], particularly herpes simplex virus type 1 (HSV1) [457], Chlamydia pneumoniae, and several types of spirochetes [458–461].

Most importantly, A β showed consistent antimicrobial activity. Soscia et al. (2010) [462] were the first to demonstrate antibacterial and antifungal activity of A β peptide against numerous pathogens. These authors found that A β can act as an antimicrobial peptide and that A β deposition can be rapidly induced in mice, in *Caenorhabditis elegans* models, and in AD-based neural cell models as an innate immune defense mechanism against microbial pathogens [462, 463].

Interestingly, synthetic A β can reduce the growth of common pathogens up to 200-fold *in vitro* [462]. Some authors reported that A β peptide strongly inhibits the infectivity of influenza A virus in cell culture [464], while others [465, 466] reported similar results for herpes simplex virus type 1 (HSV-1). Further studies showed that A β peptides can protect the host against brain infections with Salmonella enterica serovar Typhimurium, HSV-1, and HHV-6 [463, 467].

Moreover, several evidence have mainly linked HSV-1 infections to the pathogenesis of AD [468–470]. In fact, HSV-1 DNA has been detected more frequently in the brains of AD patients than in healthy controls and has been found to be co-localized with A β [471]. In addition, some studies verified the presence of IgM anti-HSV-1 antibodies in most people with AD [472]. Moreover, high titers of anti-HSV-1 antibodies have been found to be positively correlated with the development of AD-like cognitive dysfunction [473], with symptoms of mild cognitive impairment [474], and with bilateral temporal and orbitofrontal cortical gray matter volume [475]. In addition, some studies suggested that in people carrying the *APOE* ϵ 4 allele and, therefore, predisposed to develop AD, HSV-1 infection significantly increases the risk of AD [476, 477]. Furthermore, HSV-1 was shown to produce calcium-dependent GSK-3 β activation, which results in hyperphosphorylation of tau and A β PP proteins as well as A β accumulation [478, 479]. Also, HSV-1 reactivation was associated with neuroinflammation and the appearance of several markers of neurodegeneration [478–480]. In addition, the brain regions mainly affected during acute encephalitis produced by active replication of HSV-1 in neuronal cells of the brain (herpes simplex encephalitis) [481], both in humans and in experimental rodent models, are the same regions impaired in AD (limbic system, frontal and temporal cortex) [482–488]. Consistently, a high percentage of brains of elderly people contain latent HSV-1 DNA especially in CNS regions critically involved in AD [476, 489]. More recently, some authors found that a mouse model of HSV-1 infection and recurrent reactivation showed a picture resembling the phenotype of sporadic AD [490]. Indeed, after infection and multiple rounds of reactivation of the virus promoting its spread within the brain, infected mice showed accumulation of A β and hyperphosphorylated tau proteins in several brain areas, including the hippocampus, and these molecular changes were accompanied by memory deficits [490].

Interestingly, the current adult neurogenesis theory of AD is not only compatible with the antimicrobial protection hypothesis, but also shows some relevant points of convergence with it. It is noteworthy that HSV-1 latency has been observed mainly within the lateral ventricles in the SVZ, hippocampus, and brainstem before being detected in the neurons of the trigeminal ganglion [491, 492], and also in the olfactory bulbs, frontal cortex, and cerebellum in some studies [492]. More generally, the olfactory nerve, which leads to the lateral entorhinal cortex, is a portal of entry of HSV-1 [493] and other viruses [494], as well as *Chlamydia pneumoniae*, into the brain [495]. In addition, brainstem areas that harbor latent HSV directly irrigate these brain regions [190], and from the brainstem, neurons project to the thalamus and eventually reach the sensory cortex. Thus, it is interesting to note that the sites of HSV-1 latency (hippocampus and SVZ) overlap with the major niches of adult neurogenesis, and the pathways of HSV-1 infection seem consistent with the network of connected regions considered relevant in current theory in both LOAD (hippocampus - EC - OB - brainstem) and EOAD (SVZ - OB). Based on the finding that the preferred sites of HSV-1 latency are in the niches of adult neurogenesis (hippocampus and SVZ), ependymal cells and neural progenitor cells turn out to be highly susceptible to HSV-1 infection [491, 496–498]. Indeed, HSV-1 readily replicates in these cells during acute encephalitis [496, 498], and viral lytic-associated proteins were detected in these cells during latency [498]. The presence of HSV-1 in lateral ventricle ependymal cells and neural progenitor cells during latent infection alters the proliferation of NPCs as a consequence of fibroblast growth factor 2 deficiency [499], whereas HSV-1 replication during acute encephalitis results in their loss and altered differentiation [496, 498]. Interestingly, a recent study showed that HSV-1 affects adult hippocampal neurogenesis *in vitro* and *in vivo* by reducing the proliferation of NSCs and their neuronal differentiation in the SGZ of the hippocampal DG, through intracellular accumulation of A β , without inducing cell death [500, 501]. Indeed, anti-A β antibodies or experimental mouse models lacking APP (and thus unable to form A β) reverse the impairment of neurogenesis induced by HSV-1 infection [500]. Furthermore, impairment of adult hippocampal neurogenesis occurs when cognitive dysfunction induced by HSV-1 infection is not yet present, suggesting a role of adult hippocampal neurogenesis in the pathogenesis of AD [501].

Main unresolved questions in the field and possible solutions

The scenario proposed in the current theory seems to offer possible explanations for some unresolved questions in dementia and AD research (Table 1).

A β -tau interaction

While amyloid and tau pathology are clearly critical in the pathogenesis of AD, a major unresolved question at this time is how the two interact [2, 335]. According to the current theory, amyloid and tau have a twofold interaction. At a very early preclinical stage, A β deposition has an indirect microglia-mediated effect of fostering adult neurogenesis and supporting migration, amplifying tau hyperphosphorylation in the niches or during long migrations. Later, when the two are in close interaction, A β has a documented downstream effect on tau pathogenesis. The idea that A β deposition acts as a driving force for tau pathogenesis by fostering neurogenesis and supporting migration is consistent with several findings showing that A β deposition facilitates both the pathogenicity [502–506] and spread [335, 507, 508] of tau in the brain, even when A β and tau are not topographically closely related [509, 510].

Phenotypic heterogeneity in AD

Comparisons between EOAD and LOAD are particularly puzzling because, although both disorders show a generalized pattern of A β distribution, tau seems to accumulate preferentially in the memory system in aging and LOAD but predominates in a number of other neural systems in EOAD [335, 508]. Even within EOAD, different syndromes exhibit different distributions of tau, despite similar distributions of A β [335, 508]. The hypothesis of tau pathogenesis as linked to adult neurogenesis seems to combine EOAD and LOAD, as well as different EOAD syndromes, in a unique scenario. Tau pathology develops in the niches of adult neurogenesis in all types of AD, but in EOAD, it is especially active in the V-SVZ. Some neuroblasts carrying the seeds of tau pathology migrate from this niche toward different regions of the cortex guided by complex signals, after which microglia activated by A β deposition redirect migration from the RMS toward the site of injury, similar to what occurs in stroke or TBI. When these neuroblasts arrive at target regions in the occipital, parietal, frontal, or left perisylvian cortex, degeneration begins to develop. Therefore, in this model, a common mechanism through distinct migratory

Table 1
Unresolved questions in the field and possible explanations according to current theory

Open question in the field	Possible solution according to current theory
A β -tau interaction	Amyloid and tau have a twofold interaction. At a very early preclinical stage, A β deposition has an indirect microglia-mediated effect of promoting adult neurogenesis and supporting migration, with the effect of amplifying hyperphosphorylation of tau in niches or during long migrations. Later, when the two are in close interaction, A β has the documented downstream effect on tau pathogenesis.
Difference between EOAD and LOAD	Tau pathology develops in the niches of adult neurogenesis in all types of AD, but in EOAD the V-SVZ is particularly active. NSCs carrying the seeds of tau pathology migrate from this niche to different regions of the cortex driven by complex signals after microglia activated by A β deposition redirect migration from the canonical RMS. In LOAD, in accordance with an age-related decline in neurogenesis, the SGZ of the hippocampal DG remains mainly, or often exclusively, active. NSCs briefly migrate to the granular layer in the DG, where the seeds of tau pathology begin to develop and spread within the MTL.
Phenotypic heterogeneity in (EO)AD	Microglia activated by A β deposition promote neurogenesis so that more newborn neurons are present mainly in the V-SVZ niche in the EOAD. When these neuroblasts leave the V-SVZ niche and begin to migrate, microglia signals cause a redirection of the pathway from the canonical RMS and support long-distance migration. Meanwhile, the genetic program of arealization during development is reactivated and helps signal the direction for migrating neuroblasts to follow. Specifically, a mechanism based on gradients of gene expression of morphogens or transcription factors specifies certain localizations along the simple brain axes A-P, D-V and L-M. Thus, some neuroblasts carrying the seeds of tau pathology migrate to these locations in the cortex and, when they arrive at their final destination in the occipital, parietal, frontal, or left perisylvian cortex, a focus of degeneration begins to develop.
Scaling of molecular pathology to the macroscopic brain	According to current theory, adult NSCs in the SGZ niche are the first cells that carry the initial seeds of tau pathology. However, NFT formation is suppressed and delayed in the DG because of the strong clearance activity in the niche. The seeds of tau pathology, probably after some GABAergic interneurons carry transmission from SGZ NSCs, spread retrogradely from the DG to the EC along the perforant pathway connections. Then, the seeds of tau pathology spread from EC to TEC through both retrograde and anterograde transmission due to the multiple reciprocal connections between the two regions. Thus, the TEC receives a massive load of tau pathology from the EC, and this fact would explain why NFTs appear first here.
Not specificity of tau pathology in AD	According to the new theory, tau pathology would develop in the niches of adult neurogenesis in all three diseases. However, in CTE, migration is redirected to sites of injury that lie at the depth of sulci and at the perivascular level. In PART, the lack of a promoter effect on both neurogenesis and tau spreading by microglia, in the absence of A β deposition, causes limited spread of tau pathology outside MTL compared with AD.

(Continued)

Table 1
(Continued)

	Open question in the field	Possible solution according to current theory
Selective vulnerability (region)	Why some definite brain regions but not others are impacted by degeneration?	The selection of a particular brain region as a target would be determined by the localization of the SGZ niche in the MTL or the direction of migrations to the cortex made by neuroblasts carrying the seeds of tau pathology from the V-SVZ. Thus, the selection of a cortical region depends on the location of the niches or the combined outcome of the complex signals guiding the migrating neuroblasts rather than on a regional vulnerability. Therefore, the concept of regional selective vulnerability appears superfluous in the new scenario.
Selective vulnerability (cell)	Why proteins that usually show widespread expression should accumulate in one set of cells but not in apparently similar neighboring cells?	Tau pathology develops in NSCs when neurons are immature and undifferentiated and their fate, as well as their final localization, have not yet been fully decided. Therefore, in this scenario, the concept of selective cellular vulnerability appears superfluous
Preponderance in humans	Why AD develops mainly in humans although all vertebrates produce APP, β -secretase, A β , and tau protein, and neurogenesis and neuronal migration in the adult brain are well conserved from fish to primates?	The cause is the incredible development of the neocortex in humans compared with other vertebrates. Both processes implicated in the pathogenesis of AD according to current theory, i.e., metabolism driving amyloid deposition and adult neurogenesis/migration driving tau pathogenesis, are particularly stressed in the extended and interconnected cortex of the human brain. The longevity revolution, another distinct aspect of humans, is likely to contribute to the extremes of this scenario.

EOAD, early onset Alzheimer's disease; LOAD, late onset Alzheimer's disease; V-SVZ, ventricular subventricular zone; NSCs, neural stem cells; RMS, rostral migratory stream; SGZ, subgranular zone; DG, dentate gyrus; MTL, medial temporal lobe; A-P, anterior-posterior; D-V, dorsal-ventral; M-L, medial-lateral; NFTs, neurofibrillary tangles; EC, entorhinal cortex; TEC, transentorhinal cortex; CTE, chronic traumatic encephalopathy; PART, primary age-related tauopathy.

paths would explain the phenotypic heterogeneity in EOAD. In LOAD, in agreement with an age-related decline in neurogenesis [511–514], it remains active, especially, or often exclusively, the SGZ of the hippocampal DG. Neuroblasts migrate a brief distance to the granular layer in the DG, where the seeds of tau pathology start to develop and spread inside the MTL.

Not specificity of tau pathology in AD

The model of tau pathogenesis proposed in the current theory seems not to be specific for AD. For example, it was able to combine three different diseases, i.e., AD, CTE, and PART, into a unique scenario. Equally, it could be conveniently applied to other proteinopathies in different neurodegenerative diseases. In this regard, considering, for example, Parkinson's disease dementia and limbic-predominant age-related TDP-43 encephalopathy, it is intriguing to report that there is some evidence of the presence of neurogenic niches in mammals in the substantia nigra and amygdala [316].

Selective vulnerability

The clinical manifestation of a particular neurodegenerative disease reflects the region of the brain and the specific population of cells within it that are affected [515]. Major neurodegenerative diseases differ from each other not only in the type of pathological protein that accumulates but also in the regions impacted and the types of neurons that are vulnerable. Why proteins that usually show widespread expression should accumulate in one set of cells but not in apparently similar neighboring cells and why some definite brain regions but not others are impacted are fundamental questions remaining in the field [515]. These questions have been primarily conceptualized in the notion of selective vulnerability (cellular and regional) [515–517]. The current conceptualization seems to offer a simple and unexpectedly rapid solution to the question of selective vulnerability in dementia. Indeed, the selection of a definite brain region as a target would be determined either by localization of the neurogenic niches or by direction of the migration paths toward the cortex taken by neuroblasts carrying the seeds of tau pathology. Therefore, the selection would be established in advance or actively driven rather than emerging passively by virtue of regional vulnerability. In other words, according to the current theory, there would not be any regional vulnerability to determine. We can draw the same conclusion considering selective cell

vulnerability. Indeed, according to the main hypothesis, tau pathology develops in NSCs when neurons are immature, and their fate, as well as final localization, have not yet been totally decided. In this context, it is evident that the concept of selective cell vulnerability appears unnecessary.

Preponderance in humans

The current view suggests a possible explanation for another relevant question in the field. In particular, although all vertebrates produce A β PP, β -secretase, A β , and tau protein [518–520] and neurogenesis and neuronal migration in the adult brain are well conserved from fish to primates [73, 521–523], AD develops mainly in humans. The reason is most likely found in the primary distinctive aspect of the human brain, that is, the incredible development of the neocortex. In this regard, it is worth noting that both the processes implicated in AD pathogenesis in the current theory, i.e., metabolism driving amyloid deposition and adult neurogenesis/migration driving tau pathogenesis, are particularly stressed in an extended and interconnected cortex. In particular, advanced cognitive performance is highly demanding on the metabolism. At the same time, adult neurogenesis strains try to supply an adequate pool of new neurons to guarantee high plasticity and repair from injury. Finally, neuronal migration is long-distance and demanding. The longevity revolution [524], a further distinct aspect of humans, likely contributes to extremes in this scenario. Therefore, amyloid deposition and NFT formation are inevitable consequences in extremely old and advanced human brains. Unfortunately, amyloid and tau pathology interact, with the first fosters and amplifying the second, resulting in emergent AD.

Limitations

The current theory is highly speculative. Many parts of the scenario depicted are based on suggestions and hypotheses that require more concrete evidence. Moreover, I did not find any data in the literature supporting some of the hypotheses proposed. For example, one hypothesis is that long migration of neuroblasts from the V-SVZ to different regions of the cortex are implicated in the development of EOAD. However, evidence for organized, long-distance migration of newly generated neurons in the adult human brain is lacking [525–528]. Additionally, useful methods and techniques are needed to verify some hypotheses proposed in the theory

that are not yet available. In particular, the study of adult neurogenesis in humans faces many challenges. For example, it is currently not possible to confirm the existence of adult neurogenesis in the living human brain [529]. Additionally, some authors noticed that adult hippocampal neurogenesis markers degrade rapidly in fixed postmortem tissues and could thus be undetectable if the tissues are not stored and processed quickly [530]. Moreover, considering the study of neurogenesis in AD, other authors observed that data from postmortem human tissues are intrinsically controversial and difficult to interpret because as a rule, postmortem material reflects the late stages of the disease [420]. In addition, artifacts and misinterpretations can arise due to the stage of the disease and treatments provided [420]. Furthermore, these discrepancies also depend on the methods used for labeling proliferating cells [420].

ACKNOWLEDGMENTS

The theory presented in this article was awarded one of four gold prizes in the Oskar Fischer Prize competition organized by the University of Texas, College of Sciences, at San Antonio in June 2022. The prize was awarded thanks to a generous donation from Dr. James Truchard. I would like to express my immense gratitude to Dr. James Truchard and all the UTSA staff involved in organizing the competition. I would also like to express my sincere congratulations to the other winners of the Oskar Fischer Prize competition. Finally, I would like to thank Antonio Garcia, Bio-Graphics illustrator for the valuable work in making the Figures.

FUNDING

The author has no funding to report.

CONFLICT OF INTEREST

Carlo Abbate is an Editorial Board Member of this journal but was not involved in the peer-review process nor had access to any information regarding its peer-review.

REFERENCES

- [1] Masters CL, Selkoe DJ (2012) Biochemistry of amyloid β -protein and amyloid deposits in Alzheimer disease. *Cold Spring Harb Perspect Med* **2**, a006262.

- [2] Lane CA, Hardy J, Schott JM (2018) Alzheimer's disease. *Eur J Neurol* **25**, 59-70.
- [3] Hardy JA, Higgins GA (1992) Alzheimer's disease: The amyloid cascade hypothesis. *Science* **256**, 184-185.
- [4] Hardy J, Selkoe DJ (2002) The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science* **297**, 353-356.
- [5] Gotz J, Chen F, van Dorpe J, Nitsch RM (2001) Formation of neurofibrillary tangles in P301 L tau transgenic mice induced by A β 42 fibrils. *Science* **293**, 1491-1495.
- [6] Bolmont T, Clavaguera F, Meyer-Luehmann M, Herzog MC, Radde R, Staufenbiel M, Lewis J, Hutton M, Tolnay M, Jucker M (2007) Induction of tau pathology by intracerebral infusion of amyloid-beta containing brain extract and by amyloid-beta deposition in APP \times Tau transgenic mice. *Am J Pathol* **171**, 2012-2020.
- [7] Vergara C, Houben S, Suain V, Yilmaz Z, De Decker R, Dries VV, Boom A, Mansour S, Leroy K, Ando K, Brion J-P (2019) Amyloid- β pathology enhances pathological fibrillary tau seeding induced by Alzheimer PHF *in vivo*. *Acta Neuropathol* **137**, 397-412.
- [8] Bennett RE, DeVos SL, Dujardin S, Corjuc B, Gor R, Gonzalez J, Roe AD, Frosch MP, Pitstick R, Carlson GA, Hyman BT (2017) Enhanced tau aggregation in the presence of amyloid β . *Am J Pathol* **187**, 1601-1612.
- [9] Héraud C, Goufak D, Ando K, Leroy K, Suain V, Yilmaz Z, De Decker R, Authélet M, Laporte V, Octave J-N, Brion J-P (2014) Increased misfolding and truncation of tau in APP/PS1/tau transgenic mice compared to mutant tau mice. *Neurobiol Dis* **62**, 100-112.
- [10] Hurtado DE, Molina-Porcel L, Iba M, Aboagye AK, Paul SM, Trojanowski JQ, Lee VM-Y (2010) A β accelerates the spatiotemporal progression of tau pathology and augments tau amyloidosis in an Alzheimer mouse model. *Am J Pathol* **177**, 1977-1988.
- [11] Lewis J, Dickson DW, Lin WL, Chisholm L, Corral A, Jones G, Yen S-H, Sahara N, Skipper L, Yager D, Eckman C, Hardy J, Hutton M, McGowan E (2001) Enhanced neurofibrillary degeneration in transgenic mice expressing mutant tau and APP. *Science* **293**, 1487-1491.
- [12] Perez M, Ribe E, Rubio A, Lim F, Moran MA, Ramos PG, Ferrer I, Islab MTG, Avila J (2005) Characterization of a double (amyloid precursor protein-tau) transgenic: Tau phosphorylation and aggregation. *Neuroscience* **130**, 339-347.
- [13] Pooler AM, Polydoro M, Maury EA, Nicholls SB, Reddy SM, Wegmann S, William C, Saqran L, Cagsal-Getkin O, Pitstick R, Beier DR, Carlson GA, Spire-Jones TL, Hyman BT (2015) Amyloid accelerates tau propagation and toxicity in a model of early Alzheimer's disease. *Acta Neuropathol Commun* **3**, 14.
- [14] Seino Y, Kawarabayashi T, Wakasaya Y, Watanabe M, Takamura A, Yamamoto-Watanabe Y, Tomoko Kurata T, Koji Abe K, Ikeda M, Westaway D, Murakami T, St. George Hyslop P, Matsubara E, Shoji M (2010) Amyloid β accelerates phosphorylation of tau and neurofibrillary tangle formation in an amyloid precursor protein and tau double-transgenic mouse model. *J Neurosci Res* **88**, 3547-3554.
- [15] Stancu IC, Ris L, Vasconcelos B, Marinangeli C, Goeminne L, Laporte V, Laetitia E, Haylani LE, Julien Couturier J, Olivier Schakman O, Gailly P, Pierrot N, Kienlen-Campard P, Octave J-N, Dewachter I (2014) Tauopathy contributes to synaptic and cognitive deficits

- in a murine model for Alzheimer's disease. *FASEB J* **28**, 2620-2631.
- [16] Terwel D, Muyllaert D, Dewachter I, Borghgraef P, Croes S, Devijver H, Van Leuven F (2008) Amyloid activates GSK-3beta to aggravate neuronal tauopathy in bigenic mice. *Am J Pathol* **172**, 786-798.
- [17] Takashima A, Sato M, Mercken M, Tanaka S, Kondo S, Honda T, Sato K, Murayama M, Noguchi K, Nakazato Y, Takahashi H (1996) Localization of Alzheimer-associated presenilin 1 in transfected COS-7 cells. *Biochem Biophys Res Commun* **227**, 423-426.
- [18] Takashima A, Noguchi K, Michel G, Mercken M, Hoshi M, Ishiguro K, Imahori K (1996) Exposure of rat hippocampal neurons to amyloid beta peptide (25-35) induces the inactivation of phosphatidyl inositol-3 kinase and the activation of tau protein kinase I/glycogen synthase kinase-3beta. *Neurosci Lett* **203**, 33-36.
- [19] Llorens-Martin M, Jurado J, Hernandez F, Avila J (2014) GSK-3beta, a pivotal kinase in Alzheimer disease. *Front Mol Neurosci* **7**, 46.
- [20] Leroy K, Yilmaz Z, Brion JP (2007) Increased level of active GSK-3beta in Alzheimer's disease and accumulation in argyrophilic grains and in neurones at different stages of neurofibrillary degeneration. *Neuropathol Appl Neurobiol* **33**, 43-55.
- [21] Avila J, Hernandez F (2007) GSK-3 inhibitors for Alzheimer's disease. *Expert Rev Neurother* **7**, 1527-1533.
- [22] Engel T, Goni-Oliver P, Lucas JJ, Avila J, Hernandez F (2006) Chronic lithium administration to FTDP-17 tau and GSK-3beta overexpressing mice prevents tau hyperphosphorylation and neurofibrillary tangle formation, but pre-formed neurofibrillary tangles do not revert. *J Neurochem* **99**, 1445-1455.
- [23] Hernandez F, de Barreda EG, Fuster-Matanzo A, Goni-Oliver P, Lucas JJ, Avila J (2009) The role of GSK3 in Alzheimer disease. *Brain Res Bull* **80**, 248-250.
- [24] Hernandez F, Gomez de Barreda E, Fuster-Matanzo A, Lucas JJ, Avila J (2010) GSK3: A possible link between beta amyloid peptide and tau protein. *Exp Neurol* **223**, 322-325.
- [25] Jin M, Shepardson N, Yang T, Chen G, Walsh D, Selkoe DJ (2011) Soluble amyloid-protein dimers isolated from Alzheimer cortex directly induce tau hyperphosphorylation and neuritic degeneration. *Proc Natl Acad Sci U S A* **108**, 5819-5824.
- [26] Cohen AD, Price JC, Weissfeld LA, James J, Rosario BL, Bi W, Nebes RD, Saxton JA, Snitz BE, Aizenstein HA, Wolk DA, DeKosky ST, Mathis CA, Klunk WE (2009) Basal cerebral metabolism may modulate the cognitive effects of Aβ in mild cognitive impairment: An example of brain reserve. *J Neurosci* **29**, 14770-14778.
- [27] Johnson SC, Christian BT, Okonkwo OC, Oh JM, Harding S, Xu G, Hillmer AT, Wooten DW, Murali D, Barnhart TE, Hall LT, Racine AM, Klunk WE, Mathis CA, Bendlin BB, Gallagher CL, Carlsson CM, Rowley HA, Hermann BP, Dowling NM, Asthana S, Sager MA (2014) Amyloid burden and neural function in people at risk for Alzheimer's disease. *Neurobiol Aging* **35**, 576-584.
- [28] Oh H, Madison C, Baker S, Rabinovici G, Jagust W (2016) Dynamic relationships between age, amyloid-β deposition, and glucose metabolism link to the regional vulnerability to Alzheimer's disease. *Brain* **139**, 2275-2289.
- [29] Scopa C, Marrocco F, Latina V, Ruggeri F, Corvaglia V, La Regina F, Ammassari-Teule M, Middei S, Amadoro G, Meli G, Scardigli R, Cattaneo A (2020) Impaired adult neurogenesis is an early event in Alzheimer's disease neurodegeneration, mediated by intracellular Aβ oligomers. *Cell Death Differ* **27**, 934-948.
- [30] Lu M, Kosik KS (2001) Competition for microtubule-binding with dual expression of tau missense and splice isoforms. *Mol Biol Cell* **12**, 171-184.
- [31] Avila J, Lucas JJ, Perez M, Hernandez F (2004) Role of tau protein in both physiological and pathological conditions. *Physiol Rev* **84**, 361-384.
- [32] Bullmann T, de Silva R, Holzer M, Mori H, Arendt T (2007) Expression of embryonic tau protein isoforms persist during adult neurogenesis in the hippocampus. *Hippocampus* **17**, 98-102.
- [33] Brion JP, Smith C, Couck AM, Gallo JM, Anderton BH (1993) Developmental changes in tau phosphorylation: Fetal tau is transiently phosphorylated in a manner similar to paired helical filament-tau characteristic of Alzheimer's disease. *J Neurochem* **61**, 2071-2080.
- [34] Brion JP, Octave JN, Couck AM (1994) Distribution of the phosphorylated microtubule-associated protein tau in developing cortical neurons. *Neuroscience* **63**, 895-909.
- [35] Yu Y, Run X, Liang Z, Li Y, Liu F, Liu Y, Iqbal K, Grundke-Iqbal I, Gong CX (2009) Developmental regulation of tau phosphorylation, tau kinases, and tau phosphatases. *J Neurochem* **108**, 1480-1494.
- [36] Lovestone S, Hartley CL, Pearce J, Anderton BH (1996) Phosphorylation of tau by glycogen synthase kinase-3 beta in intact mammalian cells: The effects on the organization and stability of microtubules. *Neuroscience* **73**, 1145-1157.
- [37] Kenessey A, Yen SH (1993) The extent of phosphorylation of fetal tau is comparable to that of PHF-tau from Alzheimer paired helical filaments. *Brain Res* **629**, 40-46.
- [38] Fuster-Matanzo A, Llorens-Martin M, Jurado-Arjona J, Avila J, Hernandez F (2012) Tau protein and adult hippocampal neurogenesis. *Front Neurosci* **6**, 104.
- [39] Fuster-Matanzo A, De Barreda EG, Dawson HN, Vitek MP, Avila J, Hernandez F (2009) Function of tau protein in adult newborn neurons. *FEBS Lett* **583**, 3063-3068.
- [40] Llorens-Martin M, Teixeira CM, Fuster-Matanzo A, Jurado-Arjona J, Borrell V, Soriano E, Avila J, Hernandez F (2012) Tau isoform with three microtubule binding domains is a marker of new axons generated from the subgranular zone in the hippocampal dentate gyrus: Implications for Alzheimer's disease. *J Alzheimers Dis* **29**, 921-930.
- [41] Teixeira CM, Pallas-Bazarra N, Bolos M, Terreros-Roncal J, Avila J, Llorens-Martin M (2018) Untold new beginnings: Adult hippocampal neurogenesis and Alzheimer's disease. *J Alzheimers Dis* **64**, S497-S505.
- [42] Hong XP, Peng CX, Wei W, Tian Q, Liu YH, Cao FY, Wang Q, Wang JZ (2011) Relationship of adult neurogenesis with tau phosphorylation and GSK-3beta activity in subventricular zone. *Neurochem Res* **36**, 288-296.
- [43] Grundke-Iqbal I, Iqbal K, Quinlan M, Tung YC, Zaidi MS, Wisniewski HM (1986) Microtubule-associated protein tau. A component of Alzheimer paired helical filaments. *J Biol Chem* **261**, 6084-6089.
- [44] Grundke-Iqbal I, Iqbal K, Tung YC, Quinlan M, Wisniewski HM, Binder LI (1986) Abnormal phosphorylation of the microtubule-associated protein tau (tau) in Alzheimer cytoskeletal pathology. *Proc Natl Acad Sci U S A* **83**, 4913-4917.

- [45] Ihara Y, Nukina N, Miura R, Ogawara M (1986) Phosphorylated tau protein is integrated into paired helical filaments in Alzheimer's disease. *J Biochem (Tokyo)* **99**, 1807-1810.
- [46] Montejo de Garcini E, Serrano L, Avila J (1986) Self-assembly of microtubule associated protein tau into filaments resembling those found in Alzheimer disease. *Biochem Biophys Res Commun* **141**, 790-796.
- [47] Lee VM, Balin BJ, Otvos L Jr, Trojanowski JQ (1991) A68: A major subunit of paired helical filaments and derivatized forms of normal tau. *Science* **251**, 675-678.
- [48] Goedert M, Jakes R, Spillantini MG, Hasegawa M, Smith MJ, Crowther RA (1996) Assembly of microtubule associated protein tau into Alzheimer-like filaments induced by sulphated glycosaminoglycans. *Nature* **383**, 550-553.
- [49] Goedert M (1993) Tau protein and the neurofibrillary pathology of Alzheimer's disease. *Trends Neurosci* **16**, 460-465.
- [50] Morishima-Kawashima M, Hasegawa M, Takio K, Suzuki M, Yoshida H, Watanabe A, Titani K, Ihara Y (1995) Hyperphosphorylation of tau in PHF. *Neurobiol Aging* **16**, 365-371.
- [51] Ishiguro K, Omori A, Takamatsu M, Sato K, Arioka M, Uchida T, Imahori K (1992) Phosphorylation sites on tau by tau protein kinase I, a bovine derived kinase generating an epitope of paired helical filaments. *Neurosci Lett* **148**, 202-206.
- [52] Ishiguro K, Takamatsu M, Tomizawa K, Omori A, Takahashi M, Arioka M, Uchida T, Imahori K (1992) Tau protein kinase I converts normal tau protein into A68-like component of paired helical filaments. *J Biol Chem* **267**, 10897-10901.
- [53] Wang JZ, Wu Q, Smith A, Grundke-Iqbal I, Iqbal K (1998) Tau is phosphorylated by GSK-3 at several sites found in Alzheimer disease and its biological activity markedly inhibited only after it is prephosphorylated by A-kinase. *FEBS Lett* **436**, 28-34.
- [54] Liu SJ, Zhang AH, Li HL, Wang Q, Deng HM, Netzer WJ, Xu H, Wang JZ (2003) Overactivation of glycogen synthase kinase-3 by inhibition of phosphoinositol-3 kinase and protein kinase C leads to hyperphosphorylation of tau and impairment of spatial memory. *J Neurochem* **87**, 1333-1344.
- [55] Plattner F, Angelo M, Giese KP (2006) The roles of cyclin-dependent kinase 5 and glycogen synthase kinase 3 in tau hyperphosphorylation. *J Biol Chem* **281**, 25457-25465.
- [56] Takahashi M, Tomizawa K, Kato R, Sato K, Uchida T, Fujita SC, Imahori K (1994) Localization and developmental changes of tau protein kinase I/glycogen synthase kinase-3 beta in rat brain. *J Neurochem* **63**, 245-255.
- [57] Leroy K, Brion JP (1999) Developmental expression and localization of glycogen synthase kinase-3beta in rat brain. *J Chem Neuroanat* **16**, 279-293.
- [58] Hong XP, Peng CX, Wei W, Tian Q, Liu YH, Yao XQ, Zhang Y, Cao F-Y, Wang Q, Wang J-Z (2010) Essential role of tau phosphorylation in adult hippocampal neurogenesis. *Hippocampus* **20**, 1339-1349.
- [59] Gong CX, Liu F, Grundke-Iqbal I, Iqbal K (2006) Dysregulation of protein phosphorylation/dephosphorylation in Alzheimer's disease: A therapeutic target. *J Biomed Biotechnol* **31825**, 31825.
- [60] Alonso A, Del C, Li B, Grundke-Iqbal I, Iqbal K (2008) Mechanism of tau-induced neurodegeneration in Alzheimer disease and related tauopathies. *Curr Alzheimer Res* **5**, 375-384.
- [61] Avila J (2006) Tau phosphorylation and aggregation in Alzheimer's disease pathology. *FEBS Lett* **580**, 2922-2927.
- [62] Gong CX, Iqbal K (2008) Hyperphosphorylation of microtubule associated protein tau: A promising therapeutic target for Alzheimer disease. *Curr Med Chem* **15**, 2321-2328.
- [63] Iqbal K, Grundke-Iqbal I, Zaidi T, Merz PA, Wen GY, Shaikh SS, Wisniewski HM, Alafuzoff I, Winblad B (1986) Defective brain microtubule assembly in Alzheimer's disease. *Lancet* **2**, 421-426.
- [64] Yoshida H, Ihara Y (1993) Tau in paired helical filaments is functionally distinct from fetal tau: Assembly incompetence of paired helical filament-tau. *J Neurochem* **61**, 1183-1186.
- [65] Alonso AD, Zaidi T, Grundke-Iqbal I, Iqbal K (1994) Role of abnormally phosphorylated tau in the breakdown of microtubules in Alzheimer disease. *Proc Natl Acad Sci U S A* **91**, 5562-5566.
- [66] Alonso AD, Zaidi T, Novak M, Grundke-Iqbal I, Iqbal K (2001) Hyperphosphorylation induces self-assembly of tau into tangles of paired helical filaments/straight filaments. *Proc Natl Acad Sci U S A* **98**, 6923-6928.
- [67] Alonso AD, Li B, Grundke-Iqbal I, Iqbal K (2006) Polymerization of hyperphosphorylated tau into filaments eliminates its inhibitory activity. *Proc Natl Acad Sci U S A* **23**, 8864-8869.
- [68] Castellani RJ, Perry G (2019) Tau biology, tauopathy, traumatic brain injury, and diagnostic challenges. *J Alzheimers Dis* **67**, 447-467.
- [69] Morris M, Knudsen GM, Maeda S, Trinidad JC, Ioanoviciu A, Burlingame AL, Mucke L (2015) Tau post-translational modifications in wild-type and human amyloid precursor protein transgenic mice. *Nat Neurosci* **18**, 1183-1189.
- [70] Liu F, Li B, Tung EJ, Grundke-Iqbal I, Iqbal K, Gong CX (2007) Site-specific effects of tau phosphorylation on its microtubule assembly activity and self-aggregation. *Eur J Neurosci* **26**, 3429-3436.
- [71] Wang JZ, Grundke-Iqbal I, Iqbal K (2007) Kinases and phosphatases and tau sites involved in Alzheimer neurofibrillary degeneration. *Eur J Neurosci* **25**, 59-68.
- [72] Ghashghaei HT, Lai C, Anton ES (2007) Neuronal migration in the adult brain: Are we there yet? *Nat Rev Neurosci* **8**, 141-151.
- [73] Kaneko N, Sawada M, Sawamoto K (2017) Mechanisms of neuronal migration in the adult brain. *J neurochem* **141**, 835-847.
- [74] Casares-Crespo L, Calatayud-Baselga I, García-Corzo L, Mira H (2018) On the role of basal autophagy in adult neural stem cells and neurogenesis. *Front Cell Neurosci* **12**, 339.
- [75] Hong CJ, Park H, Yu SW (2016) Autophagy for the quality control of adult hippocampal neural stem cells. *Brain Res* **1649**, 166-172.
- [76] Moore Z, Taylor JM, Crack PJ (2019) The involvement of microglia in Alzheimer's disease: A new dog in the fight. *Br J Pharmacol* **176**, 3533-3543.
- [77] Femminella GD, Dani M, Wood M, Fan Z, Calsolaro V, Atkinson R, Edginton T, Hinz R, Brooks DJ, Edison P (2019) Microglial activation in early Alzheimer trajectory is associated with higher gray matter volume. *Neurology* **92**, 1331-1343.

- [78] Heneka MT, Kummer MP, Latz E (2014) Innate immune activation in neurodegenerative disease. *Nat Rev Immunol* **14**, 463-477.
- [79] Calsolaro V, Edison P (2016) Neuroinflammation in Alzheimer's disease: Current evidence and future directions. *Alzheimers Dement* **12**, 719-732.
- [80] Femminella GD, Ninan S, Atkinson R, Fan Z, Brooks DJ, Edison P (2016) Does microglial activation influence hippocampal volume and neuronal function in Alzheimer's disease and Parkinson's disease dementia? *J Alzheimers Dis* **51**, 1275-1289.
- [81] Hamelin L, Lagarde J, Dorothee G, Leroy C, Labit M, Comley RA, Cruz de Souza L, Corne H, Dauphinot L, Bertoux M, Dubois B, Gervais P, Colliot O, Potier MC, Bottlaender M, Sarazin M, the Clinical IMABio3 team (2016) Early and protective microglial activation in Alzheimer's disease: A prospective study using 18 F-DPA-714 PET imaging. *Brain* **139**, 1252-1264.
- [82] Selkoe DJ (1994) Amyloid beta-protein precursor: New clues to the genesis of Alzheimer's disease. *Curr Opin Neurobiol* **4**, 708-716.
- [83] Mrak RE, Griffin WS (2005) Glia and their cytokines in progression of neurodegeneration. *Neurobiol Aging* **26**, 349-354.
- [84] Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, Cooper NR, Eikelenboom P, Emmerling M, Fiebich BL, Finch CE, Frautschy S, Griffin WST, Hampel H, Hull M, Landreth G, Lue L-F, Mrak R, Mackenzie IR, McGeer PL, O'Banion MK, Pachter J, Pasinetti G, Plata-Salaman C, Rogers J, Rydel R, Shen Y, Streit W, Strommeyer R, Tooyoma I, Van Muiswinkel FL, Veerhuis R, Walker D, Webster S, Wegrzyniak B, Wenk G, Wyss-Coray T (2000) Inflammation and Alzheimer's disease. *Neurobiol Aging* **21**, 383-421.
- [85] Gomez-Nicola D, Fransen NL, Suzzi S, Perry VH (2013) Regulation of microglial proliferation during chronic neurodegeneration. *J Neurosci* **33**, 2481-2493.
- [86] Wyss-Coray T (2006) Inflammation in Alzheimer disease: Driving force, bystander or beneficial response? *Nat Med* **12**, 1005-1015.
- [87] Heneka MT, O'Banion MK (2007) Inflammatory processes in Alzheimer's disease. *J Neuroimmunol* **184**, 69-91.
- [88] Gray SC, Kinghorn KJ, Woodling NS (2020) Shifting equilibria in Alzheimer's disease: The complex roles of microglia in neuroinflammation, neuronal survival and neurogenesis. *Neural Regen Res* **15**, 1208.
- [89] El Khoury J, Hickman SE, Thomas CA, Cao L, Silverstein SC, Loike JD (1996) Scavenger receptor-mediated adhesion of microglia to β -amyloid fibrils. *Nature* **382**, 716-719.
- [90] Landreth GE, Reed-Geaghan EG (2009) Toll-like receptors in Alzheimer's disease. *Curr Top Microbiol Immunol* **336**, 137-153.
- [91] Venegas C, Heneka MT (2017) Danger-associated molecular patterns in Alzheimer's disease. *J Leukoc Biol* **101**, 87-98.
- [92] Gebicke-Haerter PJ (2001) Microglia in neurodegeneration: Molecular aspects. *Microsc Res Tech* **54**, 47-58.
- [93] Streit WJ (2002) Microglia as neuroprotective, immunocompetent cells of the CNS. *Glia* **40**, 133-139.
- [94] Mines M, Ding Y, Fan GH (2007) The many roles of chemokine receptors in neurodegenerative disorders: Emerging new therapeutical strategies. *Curr Med Chem* **14**, 2456-2470.
- [95] Walton NM, Sutter BM, Laywell ED, Levkoff LH, Kearns SM, Marshall GP 2nd, Scheffler B, Steindler DA (2006) Microglia instruct subventricular zone neurogenesis. *Glia* **54**, 815-825.
- [96] Nikolakopoulou AM, Dutta R, Chen Z, Miller RH, Trapp BD (2013) Activated microglia enhance neurogenesis via trypsinogen secretion. *Proc Natl Acad Sci USA* **110**, 8714-8719.
- [97] Mir S, Cai W, Carlson SW, Saatman KE, Andres DA (2017) IGF-1 mediated neurogenesis involves a novel RIT1/Akt/Sox2 cascade. *Sci Rep* **7**, 3283.
- [98] Cacci E, Ajmone-Cat MA, Anelli T, Biagioni S, Minghetti L (2008) *In vitro* neuronal and glial differentiation from embryonic or adult neural precursor cells are differently affected by chronic or acute activation of microglia. *Glia* **425**, 412-425.
- [99] Ekdahl CT, Claassen JH, Bonde S, Kokaia Z, Lindvall O (2003) Inflammation is detrimental for neurogenesis in adult brain. *Proc Natl Acad Sci USA* **100**, 13632-13637.
- [100] Pluchino S, Muzio L, Imitola J, Deleidi M, Alfaro-Cervello C, Salani G, Porcheri C, Brambilla E, Cavasinni F, Bergamaschi A, Garcia-Verdugo JM, Comi G, Khoury SJ, Martino G (2008) Persistent inflammation alters the function of the endogenous brain stem cell compartment. *Brain* **131**, 2564-2578.
- [101] Tepavčević V, Lazarini F, Alfaro-Cervello C, Kernion C, Yoshikawa K, Garcia-Verdugo JM, Lledo PM, Nait-Oumesmar B, Baron-Van Evercooren A (2011) Inflammation-induced subventricular zone dysfunction leads to olfactory deficits in a targeted mouse model of multiple sclerosis. *J Clin Invest* **121**, 4722-4734.
- [102] Fan LW, Pang Y (2017) Dysregulation of neurogenesis by neuroinflammation: Key differences in neurodevelopmental and neurological disorders. *Neural Regen Res* **12**, 366-371.
- [103] Bhattacharjee S, Lukiw WJ (2013) Alzheimer's disease and the microbiome. *Front Cell Neurosci* **7**, 153.
- [104] Bagyinszky E, Giau VV, Shim K, Suk K, An SSA, Kim S (2017) Role of inflammatory molecules in the Alzheimer's disease progression and diagnosis. *J Neurol Sci* **376**, 242-254.
- [105] Zhao Y, Cong L, Jaber V, Lukiw WJ (2017) Microbiome-derived lipopolysaccharide enriched in the perinuclear region of Alzheimer's disease brain. *Front Immunol* **8**, 1064.
- [106] Zhao Y, Jaber V, Lukiw WJ (2017) Secretory products of the human GI tract and their potential impact on Alzheimer's disease (AD): Detection of lipopolysaccharide (LPS) in AD Hippocampus. *Front Cell Infect Microbiol* **7**, 318.
- [107] Lio D, Licastro F, Scola L, Chiappelli M, Grimaldi LM, Crivello A, Colonna-Romano G, Candore G, Franceschi C, Caruso C (2003) Interleukin-10 promoter polymorphism in sporadic Alzheimer's disease. *Genes Immun* **4**, 234-238.
- [108] Arosio B, Trabattoni D, Galimberti L, Bucciarelli P, Fasano F, Calabresi C, Cazzullo CL, Vergani C, Annoni G, Clerici M (2004) Interleukin-10 and interleukin-6 gene polymorphisms as risk factors for Alzheimer's disease. *Neurobiol Aging* **25**, 1009-1015.
- [109] Speciale L, Calabrese E, Saresella M, Tinelli C, Mariani C, Sanvito L, Longhi R, Ferrante P (2007) Lymphocyte subset patterns and cytokine production in Alzheimer's disease patients. *Neurobiol Aging* **28**, 1163-1169.
- [110] Montine TJ, Sidell KR, Crews BC, Markesbery WR, Marnett LJ, Roberts LJ, Morrow JD (1999) Elevated cere-

- brospinal fluid prostaglandin E2 levels in patients with probable Alzheimer's disease. *Neurology* **53**, 1495-1498.
- [111] Combrinck M, Williams J, De Berardinis MA, Warden D, Puopolo M, Smith AD, Minghetti L (2006) Levels of CSF prostaglandin E2, cognitive decline, and survival in Alzheimer's disease. *J Neurol Neurosurg Psychiatry* **77**, 85-88.
- [112] Cameron HA, McKay RD (2001) Adult neurogenesis produces a large pool of new granule cells in the dentate gyrus. *J Comp Neurol* **435**, 406-417.
- [113] Abrous DN, Koehl M, Le Moal M (2005) Adult neurogenesis: From precursors to network and physiology. *Physiol Rev* **85**, 523-569.
- [114] Sierra A, Encinas JM, Deudero JJ, Chancey JH, Enikolopov G, Overstreet-Wadiche LS, Tsirka SE, Maletic-Savatic M (2010) Microglia shape adult hippocampal neurogenesis through apoptosis-coupled phagocytosis. *Cell Stem Cell* **7**, 483-495.
- [115] Aarum J, Sandberg K, Haerberlein SLB, Persson MA (2003) Migration and differentiation of neural precursor cells can be directed by microglia. *Proc Natl Acad Sci U S A* **100**, 15983-15988.
- [116] Xavier ALR, Kress BT, Goldman SA, de Menezes JRL, Nedergaard M (2015) A distinct population of microglia supports adult neurogenesis in the subventricular zone. *J Neurosci* **35**, 11848-11861
- [117] Menezes JR, Smith CM, Nelson KC, Luskin MB (1995) The division of neuronal progenitor cells during migration in the neonatal mammalian forebrain. *Mol Cell Neurosci* **6**, 496-508.
- [118] Zhao C, Teng EM, Summers RG Jr, Ming GL, Gage FH (2006) Distinct morphological stages of dentate granule neuron maturation in the adult mouse hippocampus. *J Neurosci* **26**, 3-11.
- [119] Platel JC, Dave KA, Bordey A (2008) Control of neuroblast production and migration by converging GABA and glutamate signals in the postnatal forebrain. *J Physiol* **586**, 3739-3743
- [120] Aloisi F (2001) Immune function of microglia. *Glia* **36**, 165-179.
- [121] Streit WJ (2000) Microglial response to brain injury: A brief synopsis. *Toxicol Pathol* **28**, 28-30.
- [122] Pluchino S, Quattrini A, Brambilla E, Gritti A, Salani G, Dina G, Galli R, Del Carro U, Amadio S, Bergami A, Furlan R, Comi G, Vescovi AL, Martino G (2003) Injection of adult neurospheres induces recovery in a chronic model of multiple sclerosis. *Nature* **422**, 688-694.
- [123] Ben-Hur T, Einstein O, Mizrahi-Kol R, Ben-Menachem O, Reinhartz E, Karussis D, Abramsky O (2003) Transplanted multipotential neural precursor cells migrate into the inflamed white matter in response to experimental autoimmune encephalomyelitis. *Glia* **41**, 73-80.
- [124] Picard-Riera N, Decker L, Delarasse C, Goude K, Nait-Oumesmar B, Liblau R, Pham-Dinh D, Baron-Van Evercooren A (2002) Experimental autoimmune encephalomyelitis mobilizes neural progenitors from the subventricular zone to undergo oligodendrogenesis in adult mice. *Proc Natl Acad Sci U S A* **99**, 13211-13216.
- [125] Arvidsson A, Collin T, Kirik D, Kokaia Z, Lindvall O (2002) Neuronal replacement from endogenous precursors in the adult brain after stroke. *Nat Med* **8**, 963-970.
- [126] Nakatomi H, Kuriu T, Okabe S, Yamamoto SI, Hatano O, Kawahara N, Tamura A, Kirino T, Nakafuku M (2002) Regeneration of hippocampal pyramidal neurons after ischemic brain injury by recruitment of endogenous neural progenitors. *Cell* **110**, 429-441.
- [127] Young MJ, Ray J, Whiteley SJ, Klassen H, Gage FH (2000) Neuronal differentiation and morphological integration of hippocampal progenitor cells transplanted to the retina of immature and mature dystrophic rats. *Mol Cell Neurosci* **16**, 197-205.
- [128] Aboody KS, Brown A, Rainov NG, Bower KA, Liu S, Yang W, Small JE, Herrlinger U, Ourednik V, Black PMcL, Breakefield XO, Snyder EY (2000) Neural stem cells display extensive tropism for pathology in adult brain: Evidence from intracranial gliomas. *Proc Natl Acad Sci U S A* **97**, 12846-12851.
- [129] Snyder EY, Yoon C, Flax JD, Macklis JD (1997) Multipotent neural precursors can differentiate toward replacement of neurons undergoing targeted apoptotic degeneration in adult mouse neocortex. *Proc Natl Acad Sci U S A* **94**, 11663-11668.
- [130] Svendsen CN, Caldwell MA, Shen J, ter Borg MG, Rosser AE, Tyers P, Karmioli S, Dunnett SB (1997) Long-term survival of human central nervous system progenitor cells transplanted into a rat model of Parkinson's disease. *Exp Neurol* **148**, 135-146.
- [131] Mundim MV, Zamproni LN, Pinto AAS, Galindo LT, Xavier AM, Glezer I, Porcionatto M (2019) A new function for Prokineticin 2: Recruitment of SVZ-derived neuroblasts to the injured cortex in a mouse model of traumatic brain injury. *Mol Cell Neurosci* **94**, 1-10.
- [132] Monterey MD, Wei H, Wu X, Wu JQ (2021) The many faces of astrocytes in Alzheimer's disease. *Front Neurol* **12**, 619626.
- [133] Reid MJ, Beltran-Lobo P, Johnson L, Perez-Nievas BG, Noble W (2020) Astrocytes in tauopathies. *Front Neurol* **11**, 572850.
- [134] Preman P, Alfonso-Triguero M, Alberdi E, Verkhatsky A, Arranz AM (2021) Astrocytes in Alzheimer's disease: Pathological significance and molecular pathways. *Cell* **10**, 540.
- [135] Cassé F, Richetin K, Toni N (2018) Astrocytes' contribution to adult neurogenesis in physiology and Alzheimer's disease. *Front Cell Neurosci* **12**, 432.
- [136] Sung PS, Lin PY, Liu CH, Su HC, Tsai KJ (2020) Neuroinflammation and neurogenesis in Alzheimer's disease and potential therapeutic approaches. *Int J Mol Sci* **21**, 701.
- [137] Garcia-Marin V, Garcia-Lopez P, Freire M (2007) Cajal's contributions to the study of Alzheimer's disease. *J Alzheimers Dis* **12**, 161-174.
- [138] Bouvier DS, Jones EV, Quesseveur G, Davoli MA, Ferreira TA, Quirion R, Mechawar N, Murai KK (2016) High resolution dissection of reactive glial nets in Alzheimer's disease. *Sci Rep* **6**, 24544.
- [139] Osborn LM, Kamphuis W, Wadman WJ, Hol EM (2016) Astrogliosis: An integral player in the pathogenesis of Alzheimer's disease. *Prog Neurobiol* **144**, 121-141.
- [140] Serrano-Pozo A, Muzikansky A, Gómez-Isla T, Growdon JH, Betensky RA, Frosch MP, Hyman BT (2013) Differential relationships of reactive astrocytes and microglia to fibrillar amyloid deposits in Alzheimer disease. *J Neuropathol Exp Neurol* **72**, 462-471.
- [141] Beach T, McGeer E (1988) Lamina-specific arrangement of astrocytic gliosis and senile plaques in Alzheimer's disease visual cortex. *Brain Res* **463**, 357-361.
- [142] Verkhatsky A, Zorec R, Rodríguez JJ, Parpura V (2016) Astroglia dynamics in ageing and Alzheimer's disease. *Curr Opin Pharm* **26**, 74-79.

- [143] Rodríguez JJ, Olabarria M, Chvatal A, Verkhratsky A, Rodr JJ (2008) Astroglia in dementia and Alzheimer's disease. *Cell Death Differ* **16**, 378-385.
- [144] Verkhratsky A, Zorec R, Parpura V (2017) Stratification of astrocytes in healthy and diseased brain. *Brain Pathol* **27**, 629-644.
- [145] Frost GR, Li YM (2017) The role of astrocytes in amyloid production and Alzheimer's disease. *Open Biol* **7**, 170228.
- [146] Burda JE, Sofroniew MV (2014) Reactive gliosis and the multicellular response to CNS damage and disease. *Neuron* **81**, 229-248.
- [147] Escartin C, Galea E, Lakatos A, O'Callaghan JP, Petzold GC, Serrano-Pozo A, Steinhäuser C, Volterra A, Carmignoto G, Agarwal A, Allen NJ, Araque A, Barbeito L, Barzilai A, Bergles DE, Bonvento G, Butt AM, Chen W-T, Cohen-Salmon M, Cunningham C, Deneen B, De Strooper B, Díaz-Castro B, Farina C, Freeman M, Gallo V, Goldman JE, Goldman SA, Götz M, Gutiérrez A, Haydon PG, Heiland DH, Hol EM, Holt MG, Iino M, Kastanenka KV, Kettenmann H, Khakh BS, Koizumi S, Lee CJ, Liddelow SA, MacVicar BA, Magistretti P, Messing A, Mishra A, Molofsky AV, Murai KK, Norris CM, Okada S, Oliet SHR, Oliveira JF, Panatier A, Parpura V, Pekna M, Pekny M, Pellerin L, Perea G, Pérez-Nievas BG, Pfrieger FW, Poskanzer KE, Quintana FJ, Ransohoff RM, Riquelme-Perez M, Robel S, Rose CR, Rothstein JD, Rouach N, Rowitch DH, Semyanov A, Sirko S, Sontheimer H, Swanson RA, Vitorica J, Wanner I-B, Wood LB, Wu J, Zheng B, Zimmer ER, Zorec R, Sofroniew MV, Verkhratsky A (2021) Reactive astrocyte nomenclature, definitions, and future directions. *Nat Neurosci* **24**, 312-325.
- [148] Pekny M, Pekna M, Messing A, Steinhäuser C, Lee JM, Parpura V, Hol EM, Sofroniew MV, Verkhratsky A (2016) Astrocytes: A central element in neurological diseases. *Acta Neuropathol* **131**, 323-345.
- [149] Li K, Li J, Zheng J, Qin S (2019) Reactive astrocytes in neurodegenerative diseases. *Aging Dis* **10**, 664-675.
- [150] Chen Y, Swanson RA (2003) Astrocytes and brain injury. *J Cereb Blood Flow Metab* **23**, 137-149.
- [151] Verkhratsky A, Olabarria M, Noristani HN, Yeh CY, Rodríguez JJ (2010) Astrocytes in Alzheimer's disease. *Neurotherapeutics* **7**, 399-412.
- [152] Diniz LP, Tortelli V, Matias I, Morgado J, Araujo APB, Melo HM, Da Silva GSS, Alves-Leon SV, De Souza JM, Ferreira ST, De Felice FG, Gomes FCA (2017) Astrocyte transforming growth factor beta 1 protects synapses against Aβ oligomers in Alzheimer's disease model. *J Neurosci* **37**, 6797-6809.
- [153] Lian H, Zheng H (2016) Signaling pathways regulating neuron-glia interaction and their implications in Alzheimer's disease. *J Neurochem* **136**, 475-491.
- [154] Gomez-Arboledas A, Davila JC, Sanchez-Mejias E, Navarro V, Nuñez-Díaz C, Sanchez-Varo R, Sanchez-Mico MV, Trujillo-Estrada L, Fernandez-Valenzuela JJ, Vizuete M, Comella JX, Galea E, Vitorica J, Gutierrez A (2018) Phagocytic clearance of presynaptic dystrophies by reactive astrocytes in Alzheimer's disease. *Glia* **66**, 637-653.
- [155] Iliiff JJ, Wang M, Liao Y, Plogg BA, Peng W, Gundersen GA, Benveniste H, Vates GE, Deane R, Goldman SA, Nagelhus EA, Nedergaard M (2012) A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid-β. *Sci Transl Med* **4**, 147ra111.
- [156] Benarroch EE (2007) Aquaporin-4, homeostasis, and neurologic disease. *Neurology* **69**, 2266-2268.
- [157] Lim DA, Alvarez-Buylla A (1999) Interaction between astrocytes and adult subventricular zone precursors stimulates neurogenesis. *Proc Natl Acad Sci U S A* **96**, 7526-7531.
- [158] Cao X, Li L-P, Qin X-H, Li S-J, Zhang M, Wang Q, Hu H-H, Fang Y-Y, Gao Y-B, Li X-W, Sun L-R, Xiong W-C, Gao T-M, Zhu X-H (2013) Astrocytic adenosine 5'-triphosphate release regulates the proliferation of neural stem cells in the adult hippocampus. *Stem Cells* **31**, 1633-1643.
- [159] Mothet J-P, Pollegioni L, Ouanounou G, Martineau M, Fossier P, Baux G (2005) Glutamate receptor activation triggers a calcium-dependent and SNARE protein-dependent release of the gliotransmitter D-serine. *Proc Natl Acad Sci U S A* **102**, 5606-5611.
- [160] Papouin T, Henneberger C, Rusakov DA, Oliet SHR (2017) Astroglial versus neuronal D-serine: Fact checking. *Trends Neurosci* **40**, 517-520.
- [161] Sultan S, Gebara E, Toni N (2013) Doxycycline increases neurogenesis and reduces microglia in the adult hippocampus. *Front Neurosci* **7**, 131.
- [162] Palmer TD, Ray J, Gage FH (1995) FGF-2-responsive neuronal progenitors reside in proliferative and quiescent regions of the adult rodent brain. *Mol Cell Neurosci* **6**, 474-486.
- [163] Kirby ED, Muroy SE, Sun WG, Covarrubias D, Leong MJ, Barchas LA, Kaufer D (2013) Acute stress enhances adult rat hippocampal neurogenesis and activation of newborn neurons via secreted astrocytic FGF2. *eLife* **2**, e00362.
- [164] Jovčić A, Gitler AD (2017) Distinct repertoires of microRNAs present in mouse astrocytes compared to astrocyte-secreted exosomes. *PLoS One* **12**, e0171418.
- [165] Song H, Stevens CF, Gage FH (2002) Astroglia induce neurogenesis from adult neural stem cells. *Nature* **417**, 39-44.
- [166] Ashton RS, Conway A, Pangarkar C, Bergen J, Lim K-I, Shah P, Bissell M, Schaffer DV (2012) Astrocytes regulate adult hippocampal neurogenesis through ephrin-B signaling. *Nat Neurosci* **15**, 1399-1406.
- [167] Christopherson KS, Ullian EM, Stokes CCA, Mullen CE, Hell JW, Agah A, Lawler J, Moshier DF, Bornstein P, Barres BA (2005) Thrombospondins are astrocyte-secreted proteins that promote CNS synaptogenesis. *Cell* **120**, 421-433.
- [168] Lu Z, Kipnis J (2010) Thrombospondin 1-a key astrocyte-derived neurogenic factor. *FASEB J* **24**, 1925-1934.
- [169] Ueki T, Tanaka M, Yamashita K, Mikawa S, Qiu Z, Maragakis NJ, Hevner RF, Miura N, Sugimura H, Sato K (2003) A novel secretory factor, Neurogenin-1, provides neurogenic environmental cues for neural stem cells in the adult hippocampus. *J Neurosci* **23**, 11732-11740.
- [170] Barkho BZ, Song H, Aimone JB, Smrt RD, Kuwabara T, Nakashima K, Gage FH, Zhao X (2006) Identification of astrocyte-expressed factors that modulate neural stem/progenitor cell differentiation. *Stem Cells Dev* **15**, 407-421.
- [171] Sultan S, Li L, Moss J, Petrelli F, Cassé F, Gebara E, Lopatar J, Pfrieger FW, Bezzi P, Bischofberger J, Toni N (2015) Synaptic integration of adult-born hippocampal neurons is locally controlled by astrocytes. *Neuron* **88**, 957-972.

- [172] Leng F, Edison P (2021) Neuroinflammation and microglial activation in Alzheimer disease: Where do we go from here? *Nat Rev Neurol* **17**, 157-172.
- [173] Kwon HS, Koh SH (2020) Neuroinflammation in neurodegenerative disorders: The roles of microglia and astrocytes. *Transl Neurodegener* **9**, 1-12.
- [174] Kempuraj D, Thangavel R, Natteru PA, Selvakumar GP, Saeed D, Zahoor H, Zaheer S, Iyer SS, Zaheer A (2016) Neuroinflammation induces neurodegeneration. *J Neurol Neurosurg Spine* **20161**, 1003.
- [175] Glass CK, Saijo K, Winner B, Marchetto MC, Gage FH (2010) Mechanisms underlying inflammation in neurodegeneration. *Cell* **140**, 918-934.
- [176] Lee DC, Rizer J, Selenica ML, Reid P, Kraft C, Johnson A, Blair L, Gordon MN, Dickey CA, Morgan D (2010) LPS-induced inflammation exacerbates phospho-tau pathology in rTg4510 mice. *J Neuroinflammation* **7**, 56.
- [177] Jo S, Yarishkin O, Hwang YJ, Chun YE, Park M, Woo DH, Bae JY, Kim T, Lee J, Chun H, Park HJ, Lee DY, Hong J, Kim HY, Oh S-J, Park SJ, Lee H, Yoon B-E, Kim YS, Jeong Y, Shim I, Bae YC, Cho J, Kowall NW, Ryu H, Hwang E, Kim D, Lee CJ (2014) GABA from reactive astrocytes impairs memory in mouse models of Alzheimer's disease. *Nat Med* **20**, 886-896.
- [178] Chang J, Liu F, Lee M, Wu B, Ting K, Zara JN, Soo C, Al Hezaimi K, Zou W, Chen X, Mooney DJ, Wang C-Y (2013) NF- κ B inhibits osteogenic differentiation of mesenchymal stem cells by promoting β -catenin degradation. *Proc Natl Acad Sci U S A* **110**, 9469-9474.
- [179] Winkler EA, Nishida Y, Sagare AP, Rege SV, Bell RD, Perlmutter D, Sengillo JD, Hillman S, Kong P, Nelson AR, Sullivan JS, Zhao Z, Meiselman HJ, Wenby RB, Soto J, Abel ED, Makshanoff J, Zuniga E, De Vivo DC, Zlokovic BV (2015) GLUT1 reductions exacerbate Alzheimer's disease vasculo-neuronal dysfunction and degeneration. *Nat Neurosci* **18**, 521-530.
- [180] Kisler K, Nelson AR, Montagne A, Zlokovic BV (2017) Cerebral blood flow regulation and neurovascular dysfunction in Alzheimer disease. *Nat Rev Neurosci* **18**, 419-434.
- [181] Heneka MT, Sastre M, Dumitrescu-Ozimek L, Dewachter I, Walter J, Klockgether T, Van Leuven F (2005) Focal glial activation coincides with increased BACE1 activation and precedes amyloid plaque deposition in APP[V717I] transgenic mice. *J Neuroinflammation* **2**, 22.
- [182] Odfalk KF, Bieniek KF, Hopp SC (2022) Microglia: Friend and foe in tauopathy. *Prog Neurobiol* **216**, 102306.
- [183] Da Gray SC, Kinghorn KJ, Woodling NS (2020) Shifting equilibriums in Alzheimer's disease: The complex roles of microglia in neuroinflammation, neuronal survival and neurogenesis. *Neural Regen Res* **15**, 120.
- [184] Braak H, Braak E (1991) Neuropathological stageing of Alzheimer-related changes. *Acta Neuropathol* **82**, 239-259.
- [185] Braak H, Braak E (1997) Frequency of stages of Alzheimer-related lesions in different age categories. *Neurobiol Aging* **18**, 351-357.
- [186] Braak H, Alafuzoff I, Arzberger T, Kretschmar H, Del Tredici K (2006) Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. *Acta Neuropathol* **112**, 389-404.
- [187] Braak H, Thal DR, Ghebremedhin E, Del Tredici K (2011) Stages of the pathologic process in Alzheimer disease: Age categories from 1 to 100 years. *J Neuropathol Exp Neurol* **70**, 960-969.
- [188] Braak H, Del Tredici K (2011) The pathological process underlying Alzheimer's disease in individuals under thirty. *Acta Neuropathol* **121**, 171-181.
- [189] Braak H, Del Tredici K (2015) Neuroanatomy and pathology of sporadic Alzheimer's disease. *Adv Anat Embryol Cell Biol* **215**, 1-162.
- [190] Braak H, Del Tredici K (2015) The preclinical phase of the pathological process underlying sporadic Alzheimer's disease. *Brain* **138**, 2814-2833.
- [191] Marien MR, Colpaert FC, Rosenquist AC (2004) Noradrenergic mechanisms in neurodegenerative diseases: A theory. *Brain Res Rev* **45**, 38-78.
- [192] Aston-Jones G, Cohen JD (2005) Adaptive gain and the role of the locus coeruleus-norepinephrine system in optimal performance. *J Comp Neurol* **493**, 99-110.
- [193] Samuels ER, Szabadi E (2009) Functional neuroanatomy of the noradrenergic locus coeruleus: Its roles in the regulation of arousal and autonomic function. Part I: Principles of functional organization. *Curr Neuropharmacol* **6**, 235-253.
- [194] Sara SJ (2009) The locus coeruleus and noradrenergic modulation of cognition. *Nature Rev Neurosci* **10**, 211-223.
- [195] Counts SE, Mufson EJ (2012) Locus coeruleus. In *The human nervous system*, 3rd ed, Mai JK, Paxinos G, eds. Academic, New York, pp. 425-438.
- [196] O'Donnell J, Zeppenfeld D, McConnell E, Pena S, Nedergaard M (2012) Norepinephrine: A neuromodulator that boosts the function of multiple cell types to optimize CNS performance. *Neurochem Res* **37**, 2496-2512.
- [197] Braak H, Braak E (1992) The human entorhinal cortex: Normal morphology and lamina-specific pathology in various diseases. *Neurosci Res* **15**, 6-31.
- [198] Attems J, Jellinger KA (2006) Olfactory tau pathology in Alzheimer's disease and mild cognitive impairment. *Clin Neuropathol* **25**, 265-271.
- [199] Braak H, Del Tredici K (2014) Are cases with tau pathology in the absence of Ab deposits part of the AD-related pathological process? *Acta Neuropathol* **128**, 767-772.
- [200] Furman JL, Vaquer-Alicea J, White CL, Cairns NJ, Nelson PT, Diamond MI (2017) Widespread tau seeding activity at early Braak stages. *Acta Neuropathol* **133**, 91-100.
- [201] Clavaguera F, Bolmont T, Crowther RA, Abramowski D, Frank S, Probst A, Fraser G, Stalder AK, Beibel M, Staufenbiel M, Jucker M, Goedert M, Tolnay M (2009) Transmission and spreading of tauopathy in transgenic mouse brain. *Nat Cell Biol* **11**, 909-913.
- [202] de Calignon A, Polydoro M, Suárez-Calvet M, William C, Adamowicz David H, Kathy KJ, Pittstick R, Sahara N, Ashe KH, Carlson GA, Spire-Jones TL, Hyman BT (2012) Propagation of tau pathology in a model of early Alzheimer's disease. *Neuron* **73**, 685-697.
- [203] Frost B, Jacks RL, Diamond MI (2009) Propagation of tau misfolding from the outside to the inside of a cell. *J Biol Chem* **284**, 12845-12852.
- [204] Guo JL, Lee VMY (2011) Seeding of normal tau by pathological tau conformers drives pathogenesis of Alzheimer-like tangles. *J Biol Chem* **286**, 15317-15331.
- [205] Liu L, Drouet V, Wu JW, Witter MP, Small SA, Clelland C, Duff K (2012) Trans-synaptic spread of tau pathology *in vivo*. *PLoS One* **7**, e31302.
- [206] Prusiner SB (1984) Some speculations about prions, amyloid, and Alzheimer's disease. *N Engl J Med* **310**, 661-663.

- [207] Weaver CL, Espinoza M, Kress Y, Davies P (2000) Conformational change as one of the earliest alterations of tau in Alzheimer's disease. *Neurobiol Aging* **21**, 719-727.
- [208] d'Errico P, Meyer-Luehmann M (2020) Mechanisms of pathogenic tau and A β protein spreading in Alzheimer's disease. *Front Aging Neurosci* **12**, 265.
- [209] Colin M, Dujardin S, Schraen-Maschke S, Meno-Tetang G, Duyckaerts C, Courade JP, Buée L (2020) From the prion-like propagation hypothesis to therapeutic strategies of anti-tau immunotherapy. *Acta Neuropathol* **139**, 3-25.
- [210] Nobuhara CK, DeVos SL, Commins C, Wegmann S, Moore BD, Roe AD, Costantino I, Frosch MP, Pitstick R, Carlson GA, Hock C, Nitsch RM, Montrasio F, Grimm J, Cheung AE, Dunah AW, Wittmann M, Bussiere T, Weinreb PH, Hyman BT (2017) Tau antibody targeting pathological species blocks neuronal uptake and interneuron propagation of tau *in vitro*. *Am J Pathol* **187**, 1399-1412.
- [211] Vandermeeren M, Borgers M, Van Kolen K, Theunis C, Vasconcelos B, Bottelbergs A, Wintmolders C, Daneels G, Willems R, Dockx K, Delbroek L, Marreiro A, Ver Donck L, Sousa C, Nanjunda R, Lacy E, Van De Castele T, Van Dam D, De Deyn PP, Kemp JA, Thomas MJ, Mercken MH (2018) Anti-tau monoclonal antibodies derived from soluble and filamentous tau show diverse functional properties *in vitro* and *in vivo*. *J Alzheimers Dis* **65**, 265-281.
- [212] Yanamandra K, Kfoury N, Jiang H, Mahan TE, Ma S, Maloney SE, Wozniak DF, Diamond MI, Holtzman DM (2013) Anti-tau antibodies that block tau aggregate seeding *in vitro* markedly decrease pathology and improve cognition *in vivo*. *Neuron* **80**, 402-414.
- [213] Evans LD, Wassmer T, Fraser G, Smith J, Perkinson M, Billinton A, Livesey FJ (2018) Extracellular monomeric and aggregated tau efficiently enter human neurons through overlapping but distinct pathways. *Cell Rep* **22**, 3612-3624.
- [214] Holmes BB, Devos SL, Kfoury N, Li M, Jacks R, Yanamandra K, Ouidja MO, Brodsky FM, Marasa J, Bagchi DP, Kotzbauer PT, Miller TM, Papy-Garcia D, Diamond MI (2013) Heparan sulfate proteoglycans mediate internalization and propagation of specific proteopathic seeds. *Proc Natl Acad Sci U S A* **110**, E3138-3147.
- [215] Holmes BB, Furman JL, Mahan TE, Yamasaki TR, Mirbaha H, Eades WC, Belaygorod L, Cairns NJ, Holtzman DM, Diamond MI (2014) Proteopathic tau seeding predicts tauopathy *in vivo*. *Proc Natl Acad Sci U S A* **111**, E4376-4385.
- [216] Nonaka T, Watanabe ST, Iwatsubo T, Hasegawa M (2010) Seeded aggregation and toxicity of alpha-synuclein and tau: Cellular models of neurodegenerative diseases. *J Biol Chem* **285**, 34885-34898.
- [217] Reilly P, Winston CN, Baron KR, Trejo M, Rockenstein EM, Akers JC, Kfoury N, Diamond M, Masliah E, Rissman RA, Yuan SH (2017) Novel human neuronal tau model exhibiting neurofibrillary tangles and transcellular propagation. *Neurobiol Dis* **106**, 222-234.
- [218] Santa-Maria I, Varghese M, Ksiezak-Reding H, Dzhun A, Wang J, Pasinetti GM (2012) Paired helical filaments from Alzheimer disease brain induce intracellular accumulation of tau protein in aggregates. *J Biol Chem* **287**, 20522-20533.
- [219] Takahashi M, Miyata H, Kametani F, Nonaka T, Akiyama H, Hisanaga S, Hasegawa M (2015) Extracellular association of APP and tau fibrils induces intracellular aggregate formation of tau. *Acta Neuropathol* **129**, 895-907.
- [220] Takeda S, Commins C, DeVos SL, Nobuhara CK, Wegmann S, Roe AD, Costantino I, Fan Z, Nicholls SB, Sherman AE, Trisini Lipsanopoulos AT, Scherzer CR, Carlson GA, Pitstick R, Peskind ER, Raskind MA, Li G, Montine TJ, Frosch MP, Hyman BT (2016) Seed-competent high-molecular-weight tau species accumulates in the cerebrospinal fluid of Alzheimer's disease mouse model and human patients. *Ann Neurol* **80**, 355-367.
- [221] Takeda S, Wegmann S, Cho H, DeVos SL, Commins C, Roe AD, Nicholls SB, Carlson GA, Pitstick R, Nobuhara CK, Costantino I, Frosch MP, Müller DJ, Irimia D, Hyman BT (2015) Neuronal uptake and propagation of a rare phosphorylated high-molecular-weight tau derived from Alzheimer's disease brain. *Nat Commun* **6**, 849.
- [222] Mudher A, Colin M, Dujardin S, Medina M, Dewachter I, Naini A, Mandelkow E-M, Mandelkow E, Buée L, Goedert M, Brion JP (2017) What is the evidence that tau pathology spreads through prion-like propagation? *Acta Neuropathol Commun* **5**, 99.
- [223] Audouard E, Houben S, Masaracchia C, Yilmaz Z, Suain V, Authélet M, De Decker R, Buée L, Boom A, Leroy K, Ando K, Brion J-P (2016) High-molecular-weight paired helical filaments from Alzheimer brain induce seeding of wild-type mouse tau into an argyrophillic 4R tau pathology *in vivo*. *Am J Pathol* **186**, 2709-2722.
- [224] Guo JL, Narasimhan S, Changolkar L, He Z, Stieber A, Zhang B, Gathagan RJ, Iba M, McBride JD, Trojanowski JQ, Lee VMY (2016) Unique pathological tau conformers from Alzheimer's brains transmit tau pathology in non-transgenic mice. *J Exp Med* **213**, 2635-2654.
- [225] Kaufman SK, Thomas TL, Del Tredici K, Braak H, Diamond MI (2017) Characterization of tau prion seeding activity and strains from formaldehyde-fixed tissue. *Acta Neuropathol Commun* **5**, 41.
- [226] Skachokova Z, Martinisi A, Flach M, Sprenger F, Naegelin Y, Steiner-Monard V (2019) Cerebrospinal fluid from Alzheimer's disease patients promotes tau aggregation in transgenic mice. *Acta Neuropathol Commun* **7**, 72.
- [227] Falcon B, Cavallini A, Angers R, Glover S, Murray TK, Barnham L, Jackson S, O'Neill MJ, Isaacs AM, Hutton ML, Szekeres PG, Goedert M, Bose S (2015) Conformation determines the seeding potencies of native and recombinant tau aggregates. *J Biol Chem* **290**, 1049-1065.
- [228] Sanders DW, Kaufman SK, DeVos SL, Sharma AM, Mirbaha H, Li A, Barker SJ, Foley AC, Thorpe JR, Serpell LC, Miller TM, Grinberg LT, Seeley WW, Diamond MI (2014) Distinct tau prion strains propagate in cells and mice and define different tauopathies. *Neuron* **82**, 1271-1288.
- [229] Clavaguera F, Akatsu H, Fraser G, Crowther RA, Frank S, Hench J, Probst A, Winkler DT, Reichwald J, Staufenbiel M, Ghetti B, Goedert M, Tolnay M (2013) Brain homogenates from human tauopathies induce tau inclusions in mouse brain. *Proc Natl Acad Sci U S A* **110**, 9535-9540.
- [230] Ahmed Z, Cooper J, Murray TK, Garn K, McNaughton E, Clarke H, Parhizkar S, Ward MA, Cavallini A, Jackson S, Bose S, Clavaguera F, Tolnay M, Lavenir I, Goedert M, Hutton ML, O'Neill MJ (2014) A novel *in vivo* model of tau propagation with rapid and progressive neurofibrillary tangle pathology: The pattern of spread is determined by connectivity, not proximity. *Acta Neuropathol* **127**, 667-683.
- [231] Dujardin S, Lécolle K, Cailliez R, Bégard S, Zommer N, Lachaud C, Carrier S, Dufour N, Aurégan G, Winderickx

- J, Hantraye P, Déglon N, Colin M, Buée L (2014) Neuron-to-neuron wild-type tau protein transfer through a trans-synaptic mechanism: Relevance to sporadic tauopathies. *Acta Neuropathol Commun* **2**, 14.
- [232] Iba M, McBride JD, Guo JL, Zhang B, Trojanowski JQ, Lee VM-Y (2015) Tau pathology spread in PS19 tau transgenic mice following locus coeruleus (LC) injections of synthetic tau fibrils is determined by the LC's afferent and efferent connections. *Acta Neuropathol* **130**, 349-362.
- [233] Peeraer E, Böttelbergs A, Van Kolen K, Stancu I-C, Vasconcelos B, Mahieu M, Duytschaever H, Ver Donck L, Torremans A, Sluydts E, Van Acker N, Kemp JA, Mercken M, Brunden KR, Trojanowski JQ, Dewachter I, Lee VMY, Moechars D (2015) Intracerebral injection of preformed synthetic tau fibrils initiates widespread tauopathy and neuronal loss in the brains of tau transgenic mice. *Neurobiol Dis* **73**, 83-95.
- [234] Duyckaerts C, Uchihara T, Seilhean D, He Y, Hauw J-J (1997) Dissociation of Alzheimer type pathology in a disconnected piece of cortex. *Acta Neuropathol* **93**, 501-507.
- [235] Zhang H, Cao Y, Ma L, Wei Y, Li H (2021) Possible mechanisms of tau spread and toxicity in Alzheimer's disease. *Front Cell Dev Biol* **9**, 707268.
- [236] Kanmert D, Cantlon A, Muratore C, Jin M, O'malley T, Lee G, Young-Pearse TL, Selkoe DJ, Walsh DM (2015) C-terminally truncated forms of tau, but not full-length tau or its C-terminal fragments, are released from neurons independently of cell death. *J Neurosci* **35**, 10851-10865.
- [237] Yanamandra K, Patel TK, Jiang H, Schindler S, Ulrich JD, Boxer AL, Miller BL, Kerwin DR, Gallardo G, Stewart F, Finn MB, Cairns NJ, Verghese PB, Fogelman I, West T, Braunstein J, Robinson G, Keyser J, Roh J, Knapik SS, Hu Y, Holtzman DM (2017) Anti-tau antibody administration increases plasma tau in transgenic mice and patients with tauopathy. *Sci Transl Med* **9**, eaal2029.
- [238] Perez M, Medina M, Hernandez F, Avila J (2018) Secretion of full-length tau or tau fragments in cell culture models. Propagation of tau in vivo and in vitro. *Biomol Concepts* **9**, 1-11.
- [239] Dujardin S, Begard S, Cailliez R, Lachaud C, Delattre L, Carrier S, Loyens A, Galas M-C, Bousset L, Melki R, Aurégan G, Hantraye P, Brouillet E, Buée L, Colin M (2014) Ectosomes: A new mechanism for non-exosomal secretion of tau protein. *PLoS One* **9**, e100760.
- [240] Katsinelos T, Zeitler M, Dimou E, Karakatsani A, Muller HM, Nachman E, Steringer JP, Ruiz de Almodovar C, Nickel W, Jahn TR (2018) Unconventional secretion mediates the trans-cellular spreading of tau. *Cell Rep* **23**, 2039-2055.
- [241] Merezhko M, Brunello CA, Yan X, Vihinen H, Jokitalo E, Uronen R-L, Huttunen HJ (2018) Secretion of tau via an unconventional non-vesicular mechanism. *Cell Rep* **25**, 2027-2035.
- [242] Wang Y, Balaji V, Kaniyappan S, Kruger L, Irsen S, Tepper K, Chandupatla RR, Maetzler W, Schneider A, Mandelkow E, Mandelkow E-M (2017) The release and trans-synaptic transmission of Tau via exosomes. *Mol Neurodegener* **12**, 5.
- [243] Abounit S, Wu JW, Duff K, Victoria GS, Zurzolo C (2016) Tunneling nanotubes: A possible highway in the spreading of tau and other prion-like proteins in neurodegenerative diseases. *Prion* **10**, 344-351.
- [244] Tardivel M, Begard S, Bousset L, Dujardin S, Coens A, Melki R, Buée L, Colin M (2016) Tunneling nanotube (TNT)-mediated neuron-to neuron transfer of pathological Tau protein assemblies. *Acta Neuropathol Commun* **4**, 117.
- [245] Perez M, Avila J, Hernandez F (2019) Propagation of tau via extracellular vesicles. *Front Neurosci* **13**, 698.
- [246] Asai H, Ikezu S, Tsunoda S, Medalla M, Luebke J, Haydar T, Wolozin B, Butovsky O, Kügler S, Ikezu T (2015) Depletion of microglia and inhibition of exosome synthesis halt tau propagation. *Nat Neurosci* **18**, 1584-1593.
- [247] Simón D, García-García E, Royo F, Falcón-Pérez JM, Avila J (2012) Proteostasis of tau. Tau overexpression results in its secretion via membrane vesicles. *FEBS Lett* **586**, 47-54.
- [248] Baker S, Polanco JC, Götz J (2016) Extracellular vesicles containing P301L mutant tau accelerate pathological tau phosphorylation and oligomer formation but do not seed mature neurofibrillary tangles in ALZ17 mice. *J Alzheimers Dis* **54**, 1207-1217.
- [249] Polanco JC, Scicluna BJ, Hill AF, Götz J (2016) Extracellular vesicles isolated from the brains of rTg4510 mice seed tau protein aggregation in a threshold-dependent manner. *J Biol Chem* **291**, 12445-12466.
- [250] Saman S, Kim W, Raya M, Visnick Y, Miro S, Jackson B, McKee AC, Alvarez VE, Lee NCY, Hall GF (2012) Exosome-associated tau is secreted in tauopathy models and is selectively phosphorylated in cerebrospinal fluid in early Alzheimer disease. *J Biol Chem* **287**, 3842-3849.
- [251] Fiandaca MS, Kapogiannis D, Mapstone M, Boxer A, Eitan E, Schwartz JB, Abner EL, Petersen RC, Federoff HJ, Miller BL, Goetzl EJ (2015) Identification of preclinical Alzheimer's disease by a profile of pathogenic proteins in neurally derived blood exosomes: A case-control study. *Alzheimers Dement* **11**, 600-607.
- [252] Guix FX, Corbett GT, Cha DJ, Mustapic M, Liu W, Mengel D, Chen Z, Aikawa E, Young-Pearse T, Kapogiannis D, Selkoe DJ, Walsh DM (2018) Detection of aggregation-competent tau in neuron-derived extracellular vesicles. *Int J Mol Sci* **19**, 663.
- [253] Winston CN, Goetzl EJ, Akers JC, Carter BS, Rockenstein EM, Galasko D, Masliah E, Rissman RA (2016) Prediction of conversion from mild cognitive impairment to dementia with neuronally derived blood exosome protein profile. *Alzheimers Dement (Amst)* **3**, 63-72.
- [254] Guo JL, and Lee VM-Y (2013) Neurofibrillary tangle-like tau pathology induced by synthetic tau fibrils in primary neurons over-expressing mutant tau. *FEBS Lett* **587**, 717-723.
- [255] Calafate S, Buist A, Miskiewicz K, Vijayan V, Daneels G, de Strooper B, de Wit J, Verstreken P, Moechars D (2015) Synaptic contacts enhance cell-to-cell tau pathology propagation. *Cell Rep* **11**, 1176-1183.
- [256] Clavaguera F, Hench J, Lavenir I, Schweighauser G, Frank S, Goedert M, Tolnay M (2014) Peripheral administration of tau aggregates triggers intracerebral tauopathy in transgenic mice. *Acta Neuropathol* **127**, 299-301.
- [257] Wu JW, Herman M, Liu L, Simoes S, Acker CM, Figueroa H, Steinberg JI, Margittai M, Kaye R, Zurzolo C, Di Paolo G, Duff KE (2013) Small misfolded tau species are internalized via bulk endocytosis and anterogradely and retrogradely transported in neurons. *J Biol Chem* **288**, 1856-1870.
- [258] Calafate S, Flavin W, Verstreken P, Moechars D (2016) Loss of Bin1 promotes the propagation of tau pathology. *Cell Rep* **17**, 931-940.
- [259] Flavin WP, Bousset L, Green ZC, Chu Y, Skarpathiotis S, Chaney MJ, Kordower JH, Melki R, Campbell EM (2017)

- Endocytic vesicle rupture is a conserved mechanism of cellular invasion by amyloid proteins. *Acta Neuropathol* **134**, 629-653.
- [260] Jucker M, Walker L (2013) Self-propagation of pathogenic protein aggregates in neurodegenerative diseases. *Nature* **501**, 45-51.
- [261] Congdon EE, Kim S, Bonchak J, Songrug T, Matzavinos A, Kuret J (2008) Nucleation-dependent tau filament formation: The importance of dimerization and an estimation of elementary rate constants. *J Biol Chem* **283**, 13806-13816.
- [262] Mirbaha H, Chen D, Morazova OA, Ruff KM, Sharma AM, Liu X, Goodarzi M, Pappu RV, Colby DW, Mirzaei H, Joachimiak LA, Diamond MI (2018) Inert and seed-competent tau monomers suggest structural origins of aggregation. *eLife* **7**, e36584.
- [263] Wu JW, Hussaini SA, Bastille IM, Rodriguez GA, Mrejeru A, Rilett K, Sanders DW, Cook C, Fu H, Boonen RACM, Herman M, Nahmani E, Emrani S, Figueroa YH, Diamond MI, Clelland CL, Wray S, Duff KE (2016) Neuronal activity enhances tau propagation and tau pathology *in vivo*. *Nat Neurosci* **19**, 1085-1092.
- [264] Walsh DM, Selkoe DJ (2016) A critical appraisal of the pathogenic protein spread hypothesis of neurodegeneration. *Nat Rev Neurosci* **17**, 251-260.
- [265] Lewis J, Dickson DW (2016) Propagation of tau pathology: Hypotheses, discoveries, and yet unresolved questions from experimental and human brain studies. *Acta Neuropathol* **131**, 27-48.
- [266] Wei Y, Liu M, Wang D (2022) The propagation mechanisms of extracellular tau in Alzheimer's disease. *J Neurol* **269**, 1164-1181.
- [267] Maphis N, Xu G, Kokiko-Cochran ON, Jiang S, Cardona A, Ransohoff RM, Lamb BT, Bhaskar K (2015) Reactive microglia drive tau pathology and contribute to the spreading of pathological tau in the brain. *Brain* **138**, 1738-1755.
- [268] Hopp SC, Lin Y, Oakley D, Roe AD, DeVos SL, Hanlon D, Hyman BT (2018) The role of microglia in processing and spreading of bioactive tau seeds in Alzheimer's disease. *J Neuroinflammation* **15**, 269.
- [269] Kovacs GG, Ferrer I, Grinberg LT, Alafuzoff I, Attems J, Budka H, Cairns NJ, Cray JF, Duyckaerts C, Ghetti B, Halliday GM, Ironside JW, Love S, Mackenzie IR, Munoz DG, Murray ME, Nelson PT, Takahashi H, Trojanowski JQ, Ansorge O, Arzberger T, Baborie A, Beach TG, Bieniek KF, Bigio EH, Bodi I, Dugger BN, Feany M, Gelpi E, Gentleman SM, Giaccone G, Hatanpaa KJ, Heale R, Hof PR, Hofer M, Hortobágyi T, Jellinger KA, Jicha GA, Ince P, Kofler J, Kövari E, Kril JJ, Mann DM, Matej R, McKee AC, McLean C, Milenkovic I, Montine TJ, Murayama S, Lee EB, Rahimi J, Rodriguez RD, Rozemüller A, Schneider JA, Schultz C, Seeley W, Seilhean D, Smith C, Tagliavini F, Takao M, Thal DR, Toledo JB, Tolnay M, Troncoso JC, Vinters HV, Weis S, Wharton SB, White III CL, Wisniewski T, Woulfe JM, Yamada M, Dickson DW (2015) Aging-related tau astrogliopathy (ARTAG): Harmonized evaluation strategy. *Acta Neuropathol* **2**, 87-102.
- [270] LoPresti P, Szuchet S, Pappasozomenos SC, Zinkowski RP, Binder LI (1995) Functional implications for the microtubule-associated protein tau: Localization in oligodendrocytes. *Proc Natl Acad Sci U S A* **92**, 10369-10373.
- [271] Müller R, Heinrich M, Heck S, Blohm D, Richters-Landsberg C (1997) Expression of microtubule-associated proteins MAP2 and tau in cultured rat brain oligodendrocytes. *Cell Tissue Res* **288**, 239-249.
- [272] Richetin K, Steullet P, Pachoud M, Perbet R, Parietti E, Maheswaran M, Eddarkaoui S, Bégard S, Pythoud C, Rey M, Caillierez R, Do KQ, Halliez S, Bezzi P, Buée L, Leuba G, Colin M, Toni N, Déglon N (2020) Tau accumulation in astrocytes of the dentate gyrus induces neuronal dysfunction and memory deficits in Alzheimer's disease. *Nat Neurosci* **23**, 1567-1579.
- [273] Boisvert MM, Erikson GA, Shokhirev MN, Allen NJ (2018) The aging astrocyte transcriptome from multiple regions of the mouse brain. *Cell Rep* **22**, 269-285.
- [274] Yamada K, Cirrito JR, Stewart FR, Jiang H, Finn MB, Holmes BB, Binder LI, Mandelkow E-M, Diamond MI, Lee VM-Y, Holtzman DM (2011) In vivo microdialysis reveals age-dependent decrease of brain interstitial fluid tau levels in P301S human tau transgenic mice. *J Neurosci* **31**, 13110-13117.
- [275] Perea JR, López E, Díez-Ballesteros JC, Ávila J, Hernández F, Bolós M (2019) Extracellular monomeric tau is internalized by astrocytes. *Front Neurosci* **13**, 442.
- [276] Andersson CR, Falsig J, Stavenhagen JB, Christensen S, Kartberg F, Rosenqvist N, Finsen B, Pedersen JT (2019) Antibody-mediated clearance of tau in primary mouse microglial cultures requires Fcγ-receptor binding and functional lysosomes. *Sci Rep* **9**, 4658.
- [277] Luo W, Liu W, Hu X, Hanna M, Caravaca A, Paul SM (2015) Microglial internalization and degradation of pathological tau is enhanced by an anti-tau monoclonal antibody. *Sci Rep* **5**, 11161.
- [278] Majerova P, Zilkova M, Kazmerova Z, Kovac A, Paholikova K, Kovacec B, Zilka N, Novak M (2014) Microglia display modest phagocytic capacity for extracellular tau oligomers. *J Neuroinflamm* **11**, 161.
- [279] Narasimhan S, Guo JL, Changolkar L, Stieber A, McBride JD, Silva LV, He Z, Zhang B, Gathagan RJ, Trojanowski JQ, Lee VMY (2017) Pathological tau strains from human brains recapitulate the diversity of tauopathies in nontransgenic mouse brain. *J Neurosci* **37**, 11406-11423.
- [280] Braak H, Del Tredici K (2016) Potential pathways of abnormal tau and α-synuclein dissemination in sporadic Alzheimer's and Parkinson's diseases. *Cold Spring Harb Perspect Biol* **8**, a023630.
- [281] Agnati LF, Bjekle B, Fuxe K (1995) Volume versus wiring transmission in the brain: A new theoretical frame of neuropsychopharmacology. *Med Res Rev* **15**, 33-45.
- [282] Nieuwenhuys R (1999) Structure and organisation of fibre systems. In *The Central Nervous System of Vertebrates*, Vol. 1, Nieuwenhuys R, Ten Donkelaar JH, Nicholson C, eds. Springer, Berlin, pp. 113-157.
- [283] Kalaria RN, Stockmeier CA, Harik SI (1989) Brain microvessels are innervated by locus ceruleus noradrenergic neurons. *Neurosci Lett* **97**, 203-208.
- [284] Fuxe K, Borroto-Escuela DO, Romero-Fernandez W, Diaz-Cabiale Z, Rivera A, Ferraro L, Tanganelli S, Tarakanov AO, Garriga P, Narvaez JA, Ciruela F, Guescini M, Agnati LF (2012) Extrasynaptic neurotransmission in the modulation of brain function. Focus on striatal neuronal-glial networks. *Front Physiol* **3**, 136.
- [285] Morrison JH, Foote SL, O'Connor D, Bloom FE (1982) Laminar, tangential and regional organization of the noradrenergic innervation of monkey cortex: Dopamine-hydroxylase immunohistochemistry. *Brain Res Bull* **9**, 309-319.
- [286] Kaufman SK, Del Tredici K, Thomas TL, Braak H, Diamond MI (2018) Tau seeding activity begins in the transentorhinal/entorhinal regions and anticipates

- phospho-tau pathology in Alzheimer's disease and PART. *Acta Neuropathol* **136**, 57-67.
- [287] Morris GP, Clark IA, Vissel B (2018) Questions concerning the role of amyloid in the definition, aetiology and diagnosis of Alzheimer's disease. *Acta Neuropathol* **136**, 663-689.
- [288] Khan UA, Liu L, Provenzano FA, Diego E Berman DE, Caterina P, Profaci CP, Sloan R, Mayeux R, Duff KE, Small SA (2014) Molecular drivers and cortical spread of lateral entorhinal cortex dysfunction in preclinical Alzheimer's disease. *Nat Neurosci* **17**, 304-311.
- [289] Stranahan AM, Mattson MP, Alexis M (2010) Selective vulnerability of neurons in layer II of the entorhinal cortex during aging and Alzheimer's disease. *Neural Plast* **2010**, 108190.
- [290] van Groen T, Miettinen P, Kadish I (2003) The entorhinal cortex of the mouse: Organization of the projection to the hippocampal formation. *Hippocampus* **13**, 133-149.
- [291] Witter MP, Van Hoesen GW, Amaral DG (1989) Topographical organization of the entorhinal projection to the dentate gyrus of the monkey. *J Neurosci* **9**, 216-228.
- [292] Witter MP, Naber PA, Van Haeften T, Machielsen WCM, Rombouts SARB, Barkhof F, Scheltens P, Lopes Da Silva FH (2000) Cortico-hippocampal communication by way of parallel parahippocampal-subicular pathways. *Hippocampus* **10**, 398-410.
- [293] Vago DR, Kesner RP (2008) Disruption of the direct perforant path input to the CA1 subregion of the dorsal hippocampus interferes with spatial working memory and novelty detection. *Behav Brain Res* **189**, 273-283.
- [294] Da Insausti R, Amaral DG (2008) Entorhinal cortex of the monkey: IV. Topographical and laminar organization of cortical afferents. *J Comp Neurol* **509**, 608-641.
- [295] Suzuki WA, Amaral DG (1994) Topographic organization of the reciprocal connections between the monkey entorhinal cortex and the perirhinal and parahippocampal cortices. *J Neurosci* **14**, 1856-1877.
- [296] Suzuki WA, Amaral DG (1994) Perirhinal and parahippocampal cortices of the macaque monkey: Cortical afferents. *J Comp Neurol* **350**, 497-533.
- [297] Suzuki WA, Naya Y (2014) The Perirhinal cortex. *Annu Rev Neurosci* **37**, 39-53.
- [298] Šimić G, Leko MB, Wray S, Harrington CR, Delalle I, Jovanov-Milošević N, Bažadona D, Buée L, de Silva R, Di Giovanni G, Wischik CM, Hof PR (2017) Monoaminergic neuropathology in Alzheimer's disease. *Prog Neurobiol* **151**, 101-138.
- [299] Christen-Zaech S, Kraftsik R, Pillevuit O, Kiraly M, Martins R, Khalili K, Miklossy J (2003) Early olfactory involvement in Alzheimer's disease. *Can J Neurol Sci* **30**, 20-25.
- [300] Attems J, Lintner F, Jellinger K (2005) Olfactory involvement in aging and Alzheimer's disease: An autopsy study. *J Alzheimers Dis* **7**, 149-157.
- [301] Murphy C (2019) Olfactory and other sensory impairments in Alzheimer disease. *Nat Rev Neurol* **15**, 11-24.
- [302] Haberly LB, Price JL (1978) Association and commissural fiber systems of the olfactory cortex of the rat. *J Comp Neurol* **181**, 781-807.
- [303] Kosel KC, Van Hoesen GW, West JR (1981) Olfactory bulb projections to the parahippocampal area of the rat. *J Comp Neurol* **198**, 467-482.
- [304] Young JK (2020) Neurogenesis makes a crucial contribution to the neuropathology of Alzheimer's disease. *J Alzheimers Dis Rep* **4**, 365-371.
- [305] Braak H, Braak E (1985) On areas of transition between entorhinal allocortex and temporal isocortex in the human brain. Normal morphology and lamina-specific pathology in Alzheimer's disease. *Acta Neuropathol* **68**, 325-332.
- [306] Pencea V, Bingaman KD, Wiegand SJ, Luskin MB (2001) Infusion of brain-derived neurotrophic factor into the lateral ventricle of the adult rat leads to new neurons in the parenchyma of the striatum, septum, thalamus, and hypothalamus. *J Neurosci* **21**, 6706-6717.
- [307] Kokoeva MV, Yin H, Flier JS (2005) Neurogenesis in the hypothalamus of adult mice: Potential role in energy balance. *Science* **310**, 679-683.
- [308] Xu Y, Tamamaki N, Noda T, Kimura K, Itokazu Y, Matsumoto N, Dezawa M, Ide C (2005) Neurogenesis in the ependymal layer of the adult rat 3rd ventricle. *Exp Neurol* **192**, 251-264.
- [309] Matsuzaki K, Katakura M, Hara T, Li G, Hashimoto M, Shido O (2009) Proliferation of neuronal progenitor cells and neuronal differentiation in the hypothalamus are enhanced in heat-acclimated rats. *Pflugers Arch* **458**, 661-673.
- [310] Perez-Martin M, Cifuentes M, Grondona JM, Lopez-Avalos MD, Gomez-Pinedo U, Garcia-Verdugo JM, Fernandez-Llebrez P (2010) IGF-I stimulates neurogenesis in the hypothalamus of adult rats. *Eur J Neurosci* **31**, 1533-1548.
- [311] Pierce AA, Xu AW (2010) De novo neurogenesis in adult hypothalamus as a compensatory mechanism to regulate energy balance. *J Neurosci* **30**, 723-730.
- [312] Lee DA, Blackshaw S (2012) Functional implications of hypothalamic neurogenesis in the adult mammalian brain. *Int J Dev Neurosci* **30**, 615-621.
- [313] Li J, Tang Y, Cai D (2012) IKKb/NF-kB disrupts adult hypothalamic neural stem cells to mediate a neurodegenerative mechanism of dietary obesity and pre-diabetes. *Nat Cell Biol* **14**, 999-1012.
- [314] Werner L, Muller-Fielitz H, Ritzal M, Werner T, Rossner M, Schwaninger M (2012) Involvement of doublecortin-expressing cells in the arcuate nucleus in body weight regulation. *Endocrinology* **153**, 2655-2664.
- [315] Migaud M, Batailler M, Pillon D, Franceschini I, Malpoux B (2011) Seasonal changes in cell proliferation in the adult sheep brain and pars tuberalis. *J Biol Rhythms* **26**, 486-496.
- [316] Feliciano DM, Bordey A, Bonfanti L (2015) Noncanonical sites of adult neurogenesis in the mammalian brain. *Cold Spring Harb Perspect Biol* **7**, a018846.
- [317] Zheng J, Li H-L, Tian N, Liu F, Wang L, Yin Y, Yue L, Ma L, Wan Y, Wang J-Z (2020) Interneuron accumulation of phosphorylated tau impairs adult hippocampal neurogenesis by suppressing GABAergic transmission. *Cell Stem Cell* **26**, 331-345.
- [318] Dong J, Pan Y-B, Wu X-R, He L-N, Liu X-D, Feng D-F, Xu T-L, Sun S, Xu N-J (2019) A neuronal molecular switch through cell-cell contact that regulates quiescent neural stem cells. *Sci Adv* **5**, eaav4416.
- [319] Song J, Zhong C, Bonaguidi MA, Sun GJ, Hsu D, Gu Y, Meletis K, Huang ZJ, Ge S, Enikolopov G, Deisseroth K, Luscher B, Christian KM, Ming G-L, Song H (2012) Neuronal circuitry mechanism regulating adult quiescent neural stem-cell fate decision. *Nature* **489**, 150-154.
- [320] Andrews-Zwilling Y, Bien-Ly N, Xu Q, Li G, Bernardo A, Yoon SY, Zwilling D, Yan TX, Chen L, Huang Y (2010) Apolipoprotein E4 causes age- and tau-dependent impairment of GABAergic interneurons, leading to learning and memory deficits in mice. *J Neurosci* **30**, 13707-13717.

- [321] Li G, Bien-Ly N, Andrews-Zwilling Y, Xu Q, Bernardo A, Ring K, Halabisky B, Deng C, Mahley RW, Huang Y (2009) GABAergic interneuron dysfunction impairs hippocampal neurogenesis in adult apolipoprotein E4 knockin mice. *Cell Stem Cell* **5**, 634-645.
- [322] Najm R, Jones EA, Huang Y (2019) Apolipoprotein E4, inhibitory network dysfunction, and Alzheimer's disease. *Mol Neurodegener* **14**, 24.
- [323] Wang C, Najm R, Xu Q, Jeong DE, Walker D, Balestra ME, Yoon SY, Yuan H, Li G, Miller ZA, Miller BL, Malloy MJ, Huang Y (2018) Gain of toxic apolipoprotein E4 effects in human iPSC-derived neurons is ameliorated by a small-molecule structure corrector. *Nat Med* **24**, 647-657.
- [324] Morrone CD, Bazzigaluppi P, Beckett TL, Hill ME, Koletar MM, Stefanovic B, McLaurin J (2020) Regional differences in Alzheimer's disease pathology confound behavioural rescue after Amyloid-attenuation. *Brain* **143**, 359-373.
- [325] Braak H, Del Tredici K (2009) Neuroanatomy and pathology of sporadic Parkinson's disease. *Adv Anat Embryol Cell Biol* **201**, 1-119.
- [326] Gorno-Tempini ML, Hillis AE, Weintraub S, Kertesz A, Mendez M, Cappa SF, Ogar JM, Rohrer JD, Black S, Boeve BF, Manes F, Dronkers NF, Vandenberghe R, Rascofsky K, Patterson K, Miller BL, Knopman DS, Hodges JR, Mesulam MM, Grossman M (2011) Classification of primary progressive aphasia and its variants. *Neurology* **76**, 1006-1014.
- [327] Mendez MF, Ghajarian M, Perryman KM (2002) Posterior cortical atrophy: Clinical characteristics and differences compared to Alzheimer's disease. *Dement Geriatr Cogn Disord* **14**, 33-40.
- [328] Ossenkoppele R, Pijnenburg YAL, Perry DC, Cohn-Sheehy BI, Scheltens NME, Vogel JW, Kramer JH, van der Vlies AE, La Joie R, Rosen HJ, van der Flier WM, Grinberg LT, Rozemuller AJ, Huang EJ, van Berckel BNM, Miller BL, Barkhof F, Jagust WJ, Scheltens P, Seeley WW, Rabinovici GD (2015) The behavioural/dysexecutive variant of Alzheimer's disease: Clinical, neuroimaging and pathological features. *Brain* **138**, 2732-2749.
- [329] Risacher SL, Anderson WH, Charil A, Castelluccio PF, Shcherbinin S, Saykin AJ, Schwarz AJ, For the Alzheimer's Disease Neuroimaging Initiative (2017) Alzheimer disease brain atrophy subtypes are associated with cognition and rate of decline. *Neurology* **89**, 2176-2186.
- [330] Koedam ELGE, Lauffer V, Van Der Vlies AE, Van Der Flier WM, Scheltens P, Pijnenburg YAL (2010) Early-versus late-onset Alzheimer's disease: More than age alone. *J Alzheimers Dis* **19**, 1401-1408.
- [331] Rabinovici GD, Jagust WJ, Furst AF, Ogar JM, Racine CA, Mormino EC, O'Neil JP, Lal RA, Dronkers NF, Miller BL, Gorno-Tempini ML (2008) A β amyloid and glucose metabolism in three variants of primary progressive aphasia. *Ann Neurol* **64**, 388-401.
- [332] de Souza LC, Corlier F, Habert M-O, Uspenskaya O, Maroy R, Lamari F, Chupin M, Lehericy S, Colliot O, Hahn-Barma V, Samri D, Dubois B, Bottilaender M, Sarazin M (2011) Similar amyloid- β burden in posterior cortical atrophy and Alzheimer's disease. *Brain* **134**, 2036-2043.
- [333] Lehmann M, Ghosh PM, Madison C, Laforce Jr R, Corbetta-Rastelli C, Weiner MW, Greicius MD, Seeley WW, Gorno-Tempini ML, Rosen HJ, Miller BL, Jagust WJ, Rabinovici GD (2013) Diverging patterns of amyloid deposition and hypometabolism in clinical variants of probable Alzheimer's disease. *Brain* **136**, 844-858.
- [334] Ossenkoppele R, Schonhaut DR, Schöll M, Lockhart SN, Ayakta N, Baker SL, O'Neil JP, Janabi M, Lazaris A, Cantwell A, Vogel J, Santos M, Miller ZA, Bettcher BM, Vessel KA, Kramer JH, Gorno-Tempini ML, Miller BL, Jagust WJ, Rabinovici GD (2016) Tau PET patterns mirror clinical and neuroanatomical variability in Alzheimer's disease. *Brain* **139**, 1551-1567.
- [335] Jagust W (2018) Imaging the evolution and pathophysiology of Alzheimer disease. *Nat Rev Neurosci* **19**, 687-700.
- [336] Buckner RL, Snyder AZ, Shannon BJ, LaRossa G, Sachs R, Fotenos AF, Sheline YI, Klunk WE, Mathis CA, Morris JC, Mintun MA (2005) Molecular, structural, and functional characterization of Alzheimer's disease: Evidence for a relationship between default activity, amyloid, and memory. *J Neurosci* **25**, 7709-7717.
- [337] Buckner RL, Sepulcre J, Talukdar T, Krienen FM, Liu H, Hedden T, Andrews-Hanna JR, Sperling RA, Johnson KA (2009) Cortical hubs revealed by intrinsic functional connectivity: Mapping, assessment of stability, and relation to Alzheimer's disease. *J Neurosci* **29**, 1860-1873.
- [338] Daianu M, Jahanshad N, Nir TM, Jack CR Jr., Weiner MW, Bernstein MA, Thompson PM, the Alzheimer's Disease Neuroimaging Initiative (2015) Rich club analysis in the Alzheimer's disease connectome reveals a relatively undisturbed structural core network. *Hum Brain Mapp* **36**, 3087-3103.
- [339] Elman JA, Madison CM, Baker SL, Vogel JW, Marks SM, Crowley S, O'Neil JP, Jagust WJ (2016) Effects of beta-amyloid on resting state functional connectivity within and between networks reflect known patterns of regional vulnerability. *Cereb Cortex* **26**, 695-707.
- [340] van der Flier WM, Pijnenburg YA, Fox NC, Scheltens P (2011) Early-onset versus late-onset Alzheimer's disease: The case of the missing APOE ϵ 4 allele. *Lancet Neurol* **10**, 280-288.
- [341] Ossenkoppele R, Lyoo CH, Sudre CH, van Westen D, Cho H, Ryu YH, Choi JY, Smith R, Strandberg O, Palmqvist S, Westman E, Tsai R, Kramer J, Boxer AL, Gorno-Tempini ML, La Joie R, Miller BL, Rabinovici GD, Hansson O, (2020) Distinct tau PET patterns in atrophy-defined subtypes of Alzheimer's disease. *Alzheimers Dement* **16**, 335-344.
- [342] Franzmeier N, Dewenter A, Frontzkowski L, Dichgans M, Rubinski A, Neitzel J, Smith R, Strandberg O, Ossenkoppele R, Buerger K, Duering M, Hansson O, Ewers M (2020) Patient-centered connectivity-based prediction of tau pathology spread in Alzheimer's disease. *Sci adv* **6**, eabd1327.
- [343] Ferreira D, Nordberg A, Westman E (2020) Biological subtypes of Alzheimer's disease: A systematic review and meta-analysis. *Neurology* **94**, 436-448.
- [344] Habes M, Grothe MJ, Tunc B, Mcmillan C, Wolk DA, Davatzikos C (2020) Disentangling heterogeneity in Alzheimer's disease and related dementias using data-driven methods. *Biol Psychiatry* **88**, 70-82.
- [345] Hanna Al-Shaikh FS, Duara R, Crook JE, Lesser ER, Schaefferbeke J, Hinkle KM, Ross OA, Ertekin-Taner N, Pedraza O, Dickson DW, Graff-Radford NR, Murray ME (2020) Selective vulnerability of the nucleus basalis of meynert among neuropathologic subtypes of Alzheimer disease. *JAMA Neurol* **77**, 225-233.

- [346] Jellinger KA (2021) Pathobiological subtypes of Alzheimer disease. *Dement Geriatr Cogn Disord* **49**, 321-333.
- [347] Murray ME, Graf-Radford NR, Ross OA, Petersen RC, Duara R, Dickson DW (2011) Neuropathologically defined subtypes of Alzheimer's disease with distinct clinical characteristics: A retrospective study. *Lancet Neurol* **10**, 785-796.
- [348] Ferreira D, Mohanty R, Murray ME, Nordberg A, Kantarci K, Westman E (2022) The hippocampal sparing subtype of Alzheimer's disease assessed in neuropathology and *in vivo* tau positron emission tomography: A systematic review. *Acta Neuropathol Commun* **10**, 166.
- [349] Charil A, Shcherbinin S, Southekal S, Devous MD, Mintun M, Murray ME, Miller BB, Schwarz AJ (2019) Tau subtypes of Alzheimer's disease determined *in vivo* using flortaucipir PET imaging. *J Alzheimers Dis* **71**, 1037-1048.
- [350] Mohanty R, Mårtensson G, Poulakis K, Muehlboeck J-S, Rodriguez-Vieitez E, Chiotis K, Grothe MJ, Nordberg A, Ferreira D (2020) Comparison of subtyping methods for neuroimaging studies in Alzheimer's disease: A call for harmonization. *Brain Commun* **2**, 192.
- [351] La Joie R, Visani AV, Lesman-Segev OH, Baker SL, Edwards L, Iaccarino L, Soleimani-Meigooni DN, Mellinger T, Janabi M, Miller ZA, Perry DC, Pham J, Strom A, Gorno-Tempini ML, Rosen HJ, Miller BL, Jagust WJ, Rabinovici GD (2021) Association of APOE4 and clinical variability in Alzheimer disease with the pattern of tau- and amyloid-PET. *Neurology* **96**, e650-e661.c.
- [352] Xia C, Makarets SJ, Caso C, McGinnis S, Gomperts SN, Sepulcre J, Gomez-Isla T, Hyman BT, Schultz A, Vasdev N, Johnson KA, Dickerson BC (2017) Association of *in vivo* [18F]AV-1451 tau PET imaging results with cortical atrophy and symptoms in typical and atypical Alzheimer disease. *JAMA Neurol* **74**, 427-436.
- [353] Sintini I, Schwarz CG, Martin PR, Graff-Radford J, Machulda MM, Senjem ML, Reid RI, Spychalla AJ, Drubach DA, Lowe VJ, Jack Jr CR, Josephs KA, Whitwell JL (2019) Regional multimodal relationships between tau, hypometabolism, atrophy, and fractional anisotropy in atypical Alzheimer's disease. *Hum Brain Mapp* **40**, 1618-1631.
- [354] Nasrallah IM, Chen YS, Hsieh M-K, Phillips JS, Ternes K, Stockbower GE, Sheline Y, McMillan CT, Grossman M, Wolk DA (2018) 18F-flortaucipir PET/MRI correlations in nonamnestic and amnestic variants of Alzheimer disease. *J Nucl Med* **59**, 299-306.
- [355] Day GS, Gordon BA, Jackson K, Christensen JJ, Rosana Ponisio M, Su Y, Ances BM, Benzinger TLS, Morris JC (2017) Tau-PET binding distinguishes patients with early-stage posterior cortical atrophy from amnestic Alzheimer disease dementia. *Alzheimer Dis Assoc Disord* **31**, 87-93.
- [356] Josephs KA, Martin PR, Botha H, Schwarz CG, Duffy JR, Clark HM, Machulda MM, Graff-Radford J, Weigand SD, Senjem ML, Utianski RL, Drubach DA, Boeve BF, Jones DT, Knopman DS, Petersen RC, Jack Jr CR, Lowe VJ, Whitwell JL (2018) [18F] AV-1451 tau-PET and primary progressive aphasia. *Ann Neurol* **83**, 599-611.
- [357] Smith R, Schöll M, Widner H, van Westen D, Svenningsson P, Hägerström D, Ohlsson T, Jögi J, Nilsson C, Hansson O (2017) *In vivo* retention of 18F-AV-1451 in corticobasal syndrome. *Neurology* **89**, 845-853.
- [358] Ali F, Whitwell J, Martin P, Senjem M, Knopman D, Jack C, Lowe VJ, Petersen RC, Boeve BF, Josephs KA (2018) [18F] AV-1451 uptake in corticobasal syndrome: The influence of beta-amyloid and clinical presentation. *J Neurol* **265**, 1079-1088.
- [359] Vogel JW, Young AL, Oxtoby NP, Smith R, Ossenkoppele R, Strandberg OT, La Joie R, Aksman LM, Grothe MJ, Turria-Medina Y, the Alzheimer's Disease Neuroimaging Initiative, Pontecorvo MJ, Devous MD, Rabinovici GD, Alexander DC, Lyoo CH, Evans AC, Hansson O (2021) Four distinct trajectories of tau deposition identified in Alzheimer's disease. *Nat Med* **27**, 871-881.
- [360] Sirkis DW, Bonham LW, Johnson TP, La Joie R, Yokoyama JS (2022) Dissecting the clinical heterogeneity of early-onset Alzheimer's disease. *Mol Psychiatry* **27**, 2674-2688.
- [361] Whitwell JL, Graf-Radford J, Tosakulwong N, Weigand SD, Machulda M, Senjem ML, Schwarz CG, Spychalla AJ, Jones DT, Drubach DA, Knopman DS, Boeve BF, Ertekin-Taner N, Petersen RC, Lowe VJ, Jack Jr CR, Josephs KA (2018) [18F]AV-1451 clustering of entorhinal and cortical uptake in Alzheimer's disease. *Ann Neurol* **83**, 248-257.
- [362] Schwarz AJ, Yu P, Miller BB, Shcherbinin S, Dickson J, Navitsky M, Joshi AD, Devous MD, Mintun MS (2016) Regional profiles of the candidate tau PET ligand 18F-AV-1451 recapitulate key features of Braak histopathological stages. *Brain* **139**, 1539-1550.
- [363] Alfano C, Studer M (2013) Neocortical arealization: Evolution, mechanisms, and open 200 questions. *Dev Neurobiol* **73**, 411-447.
- [364] Kanekar S, Gent M (2011) Malformations of cortical development. *Semin Ultrasound CT MR* **32**, 211-227.
- [365] Leventer RJ, Jansen A, Pilz DT, Stoodley N, Marini C, Dubeau F, Malone J, Mitchell LA, Mandelstam S, Scheffer IE, Berkovic SF, Andermann F, Andermann E, Guerrini R, Dobyns WB (2010) Clinical and imaging heterogeneity of polymicrogyria: A study of 328 patients. *Brain* **133**, 1415-1427.
- [366] Abbate C (2018) Topographic markers drive proteinopathies to selection of target brain areas at onset in neurodegenerative dementias. *Front Aging Neurosci* **10**, 308.
- [367] Khodanovich M, Nemirovich-Danchenko NM (2019) New neurons in the post-ischemic and injured brain: Migrating or resident? *Front Neurosci* **13**, 588.
- [368] Sun D (2016) Endogenous neurogenic cell response in the mature mammalian brain following traumatic injury. *Exp Neurol* **275**, 405-410.
- [369] Marques BL, Carvalho GA, Freitas EMM, Chiareli RA, Barbosa TG, Di Araújo AGP, Nogueira YL, Ribeiro RI, Parreira RC, Vieira MS, Resende RR, Gomez RS, Oliveira-Lima OC, Pinto MCX (2019) The role of neurogenesis in neurorepair after ischemic stroke. *Semin Cell Dev Biol* **95**, 98-110.
- [370] Schmidt W, Reymann KG (2002) Proliferating cells differentiate into neurons in the hippocampal CA1 region of gerbils after global cerebral ischemia. *Neurosci Lett* **334**, 153-156.
- [371] Bendel O, Bueters T, von Euler M, Ove Ogren S, Sandin J, von Euler G (2005) Reappearance of hippocampal CA1 neurons after ischemia is associated with recovery of learning and memory. *J Cereb Blood Flow Metab* **25**, 1586-1595.
- [372] Wojcik L, Sawicka A, Rivera S, Zaleska T (2009) Neurogenesis in gerbil hippocampus following brain ischemia: Focus on the involvement of metalloproteinases. *Acta Neurobiol Exp* **69**, 52-61.

- [373] Khodanovich MY, Kisel AA, Chernysheva GA, Smolyakova VI, Savchenko RR, Plotnikov MB (2016) Effect of fluoxetine on neurogenesis in hippocampal dentate gyrus after global transient cerebral ischemia in rats. *Bull Exp Biol Med* **161**, 351-354.
- [374] Khodanovich MY, Kisel AA, Kudabaeva MS, Chernysheva GA, Smolyakova VI, Glazacheva VY, Wasserlauf I, Pishchelko A, Plotnikov M, Yarnykh V (2018) Abnormal migration of immature neurons in the global cerebral ischemia model. *Am Neurol Assoc Ann Meet* **84**, 154-155.
- [375] Parent JM, Vexler ZS, Gong C, Derugin N, Ferriero DM (2002) Rat forebrain neurogenesis and striatal neuron replacement after focal stroke. *Ann Neurol* **52**, 802-813.
- [376] Jin K, Sun Y, Xie L, Peel A, Mao XO, Bateur S, Greenberg DA (2003) Directed migration of neuronal precursors into the ischemic cerebral cortex and striatum. *Mol Cell Neurosci* **24**, 171-189.
- [377] Yamashita T, Ninomiya M, Hernández Acosta P, García-Verdugo JM, Sunabori T, Sakaguchi M, Adachi K, Kojima T, Hirota Y, Kawase T, Araki N, Abe K, Okano H, Sawamoto K (2006) Subventricular zone-derived neuroblasts migrate and differentiate into mature neurons in the post-stroke adult striatum. *J Neurosci* **26**, 6627-6636.
- [378] Hayashi Y, Jinnou H, Sawamoto K, Hitoshi S (2018) Adult neurogenesis and its role in brain injury and psychiatric diseases. *J Neurochem* **147**, 584-594.
- [379] Thored P, Arvidsson A, Cacci E, Ahlenius H, Kallur T, Darsalia V, Ekdahl CT, Kokaia Z, Lindvall O (2006) Persistent production of neurons from adult brain stem cells during recovery after stroke. *Stem Cells* **24**, 739-747.
- [380] Fuentealba LC, Rompani SB, Parraguez JI, Obernier K, Romero R, Cepko CL, Alvarez-Buylla A (2015) Embryonic origin of postnatal neural stem cells. *Cell* **161**, 1644-1655.
- [381] Bielefeld P, Dura I, Danielewicz J, Lucassen PJ, Baekelandt V, Abrous DN, Encinas JM, Fitzsimons CP (2019) Insult-induced aberrant hippocampal neurogenesis: Functional consequences and possible therapeutic strategies. *Behav Brain Res* **372**, 112032.
- [382] Jessberger S, Nakashima K, Clemenson GD, Mejia E, Mathews E, Ure K, Ogawa S, Sinton CM, Gage FH, Hsieh J (2007) Epigenetic modulation of seizure induced neurogenesis and cognitive decline. *J Neurosci* **27**, 5967-5975.
- [383] Sierra A, Martín-Suárez S, Valcárcel-Martín R, Pascual-Brazo J, Aelvoet S-A, Abiega O, Deudero JJ, Brewster AL, Bernales I, Anderson AE, Baekelandt V, Maletić-Savatić M, Encinas JM (2015) Neuronal hyperactivity accelerates depletion of neural stem cells and impairs hippocampal neurogenesis. *Cell Stem Cell* **16**, 488-503.
- [384] Lugert S, Basak O, Knuckles P, Haussler U, Fabel K, Götz M, Haas CA, Kempermann G, Taylor V, Giachino C (2010) Quiescent and active hippocampal neural stem cells with distinct morphologies respond selectively to physiological and pathological stimuli and aging. *Cell Stem Cell* **6**, 445-456.
- [385] Bielefeld P, Schouten M, Meijer GM, Breuk MJ, Geijtenbeek K, Karayel S, Tiaglik A, Vuuregge AH, Willems RAL, Witkamp D, Lucassen PJ, Encinas JM, Fitzsimons CP (2019) Co-administration of anti microRNA-124 and -137 oligonucleotides prevents hippocampal neural stem cell loss upon non-convulsive seizures. *Front Mol Neurosci* **12**, 31.
- [386] Parent JM, Yu TW, Leibowitz RT, Geschwind DH, Sloviter RS, Lowenstein DH (1997) Dentate granule cell neurogenesis is increased by seizures and contributes to aberrant network reorganization in the adult rat hippocampus. *J Neurosci* **17**, 3727-3738.
- [387] Jessberger S, Römer B, Babu H, Kempermann G (2005) Seizures induce proliferation and dispersion of doublecortin-positive hippocampal progenitor cells. *Exp Neurol* **196**, 342-351.
- [388] Overstreet-Wadiche LS, Bromberg DA, Bensen AL, Westbrook GL (2006) Seizures accelerate functional integration of adult-generated granule cells. *J Neurosci* **26**, 4095-4103.
- [389] LaSarge CL, Pun RYK, Muntifering MB, Danzer SC (2016) Disrupted hippocampal network physiology following PTEN deletion from newborn dentate granule cells. *Neurobiol Dis* **96**, 105-114.
- [390] McCloskey DP, Hintz TM, Pierce JP, Scharfman HE (2006) Stereological methods reveal the robust size and stability of ectopic hilar granule cells after pilocarpine induced status epilepticus in the adult rat. *Eur J Neurosci* **24**, 2203-2210.
- [391] Kaneko N, Sawamoto K (2009) Adult neurogenesis and its alteration under pathological conditions. *Neurosci Res* **63**, 155-164.
- [392] Lazarov O, Marr RA (2010) Neurogenesis and Alzheimer's disease: At the crossroads. *Exp Neurol* **223**, 267-281.
- [393] Lazarov O, Marr RA (2013) Of mice and men: Neurogenesis, cognition and Alzheimer's disease. *Front Aging Neurosci* **5**, 43.
- [394] Mu Y, Gage FH (2011) Adult hippocampal neurogenesis and its role in Alzheimer's disease. *Mol Neurodegener* **6**, 85.
- [395] Winner B, Kohl Z, Gage FH (2011) Neurodegenerative disease and adult neurogenesis. *Eur J Neurosci* **33**, 1139-1151.
- [396] Bolos M, Pallas-Bazarra N, Terreros-Roncal J, Perea JR, Jurado-Arjona J, Avila J, Llorens-Martín M (2017) Soluble tau has devastating effects on the structural plasticity of hippocampal granule neurons. *Transl Psychiatry* **7**, 1267.
- [397] Pierfelice T, Alberi L, Gaiano N (2011) Notch in the vertebrate nervous system: An old dog with new tricks. *Neuron* **69**, 840-855.
- [398] Kandel ER (2012) The molecular biology of memory: cAMP, PKA, CRE, CREB-1, CREB-2, and CPEB. *Mol Brain* **5**, 14.
- [399] Ge S, Goh EL, Sailor KA, Kitabatake Y, Ming GL, Song H (2006) GABA regulates synaptic integration of newly generated neurons in the adult brain. *Nature* **439**, 589-593.
- [400] Jagasia R, Steib K, Englberger E, Herold S, Faus-Kessler T, Saxe M, Gage FH, Song H, Lie DC (2009) GABA-cAMP response element-binding protein signaling regulates maturation and survival of newly generated neurons in the adult hippocampus. *J Neurosci* **29**, 7966-7977.
- [401] Herold S, Jagasia R, Merz K, Wassmer K, Lie DC (2011) CREB signalling regulates early survival, neuronal gene expression and morphological development in adult subventricular zone neurogenesis. *Mol Cell Neurosci* **46**, 79-88.
- [402] Merz K, Herold S, Lie DC (2011) CREB in adult neurogenesis master and partner in the development of adult-born neurons? *Eur J Neurosci* **33**, 1078-1086.
- [403] Vitolo OV, Sant'Angelo A, Costanzo V, Battaglia F, Arancio O, Shelanski M (2002) Amyloid beta peptide inhibition of the PKA/CREB pathway and long-term potentiation:

- Reversibility by drugs that enhance cAMP signaling. *Proc Natl Acad Sci U S A* **99**, 13217-13221.
- [404] Ma QL, Harris-White ME, Ubeda OJ, Simmons M, Beech W, Lim GP, Teter B, Frautschy SA, Cole GM (2007) Evidence of Abeta- and transgene-dependent defects in ERK-CREB signaling in Alzheimer's models. *J Neurochem* **103**, 1594-1607.
- [405] Caccamo A, Maldonado MA, Bokov AF, Majumder S, Oddo S (2010) CBP gene transfer increases BDNF levels and ameliorates learning and memory deficits in a mouse model of Alzheimer's disease. *Proc Natl Acad Sci U S A* **107**, 22687-22692.
- [406] Bartolotti N, Segura L, Lazarov O (2015) Diminished CRE-induced plasticity is linked to memory deficits in familial Alzheimer's disease mice. *J Alzheimers Dis* **50**, 477-489.
- [407] Chenn A, Walsh CA (2003) Increased neuronal production, enlarged forebrains and cytoarchitectural distortions in beta-catenin overexpressing transgenic mice. *Cereb Cortex* **13**, 599-606.
- [408] Sato N, Meijer L, Skaltsounis L, Greengard P, Brivanlou AH (2004) Maintenance of pluripotency in human and mouse embryonic stem cells through activation of Wnt signaling by a pharmacological GSK-3-specific inhibitor. *Nat Med* **10**, 55-63.
- [409] Lie DC, Colamarino SA, Song HJ, Désiré L, Mira H, Consiglio A, Lein ES, Jessberger S, Lansford H, Dearie AR, Gage FH (2005) Wnt signalling regulates adult hippocampal neurogenesis. *Nature* **437**, 1370-1375.
- [410] Shimizu T, Kagawa T, Inoue T, Nonaka A, Takada S, Aburatani H, Taga T (2008) Stabilized beta-catenin functions through TCF/LEF proteins and the Notch/RBP-Jkappa complex to promote proliferation and suppress differentiation of neural precursor cells. *Mol Cell Biol* **28**, 7427-7441.
- [411] Choi SH, Bylykbashie E, Chatila ZK, Lee SW, Pulli B, Clemenson GD, Kim E, Rompala A, Oram MK, Asselin C, Aronson J, Zhang C, Miller SJ, Lesinski A, Chen JW, Kim DY, Van Praag H, Spiegelman BM, Gage FH, Tanzi RE (2018) Combined adult neurogenesis and BDNF mimic exercise effects on cognition in an Alzheimer's mouse model. *Science* **361**, eaan8821.
- [412] Briley D, Ghirardi V, Woltjer R, Renck A, Zolochovska O, Tagliatalata G, Micci MA (2016) Preserved neurogenesis in non-demented individuals with AD neuropathology. *Sci Rep* **6**, 27812.
- [413] Horgusluoglu E, Nudelman K, Nho K, Saykin AJ (2017) Adult neurogenesis and neurodegenerative diseases: A systems biology perspective. *Am J Med Genet B Neuropsychiatr Genet* **174**, 93-112.
- [414] Hamilton LK, Joppe SE, L MC, Fernandes KJ (2013) Aging and neurogenesis in the adult forebrain: What we have learned and where we should go from here. *Eur J Neurosci* **37**, 1978-1986.
- [415] Kuhn HG, Cooper-Kuhn CM, Boekhoorn K, Lucassen PJ (2007) Changes in neurogenesis in dementia and Alzheimer mouse models: Are they functionally relevant? *Eur Arch Psychiatry Clin Neurosci* **257**, 281-289.
- [416] Boekhoorn K, Joels M, Lucassen PJ (2006) Increased proliferation reflects glial and vascular-associated changes, but not neurogenesis in the presenile Alzheimer hippocampus. *Neurobiol Dis* **24**, 1-14.
- [417] Ziabreva I, Perry E, Perry R, Minger SL, Ekonomou A, Przyborski S, Ballard C (2006) Altered neurogenesis in Alzheimer's disease. *J Psychosom Res* **61**, 311-316.
- [418] Wang R, Dineley KT, Sweatt JD, Zheng H (2004) Presenilin 1 familial Alzheimer's disease mutation leads to defective associative learning and impaired adult neurogenesis. *Neuroscience* **126**, 305-312.
- [419] Zhang C, McNeil E, Dressler L, Siman R (2006) Long-lasting impairment in hippocampal neurogenesis associated with amyloid deposition in a knock-in mouse model of familial Alzheimer's disease. *Exp Neurol* **204**, 77-87.
- [420] Rodríguez JJ, Verkhratsky A (2011) Neurogenesis in Alzheimer's disease. *J Anat* **219**, 78-89.
- [421] Jin K, Galvan V, Xie L, Mao XO, Gorostiza OF, Bredesen DE, Greenberg DA (2004) Enhanced neurogenesis in Alzheimer's disease transgenic (PDGF-APPsw,Ind) mice. *Proc Natl Acad Sci U S A* **101**, 13363-13367.
- [422] Crary JF, Trojanowski JQ, Schneider JA, Abisambra JF, Abner EL, Alafuzoff I, Arnold SE, Attems J, Beach TG, Bigio EH, Cairns NJ, Dickson DW, Gearing M, Grinberg LT, Hof PR, Hyman BT, Jellinger KA, Jicha GA, Kovacs GG, Knopman DS, Kofler J, Kukull WA, Mackenzie IR, Masliah E, McKee A, Montine TJ, Murray ME, Neltner JH, Santa-Maria I, Seeley WW, Serrano-Pozo A, Shelanski ML, Stein T, Takao M, Thal DR, Toledo JB, Troncoso JC, Vonsattel JP, White CL 3rd, Wisniewski T, Woltjer RL, Yamada M, Nelson PT (2014) Primary age-related tauopathy (PART): A common pathology associated with human aging. *Acta Neuropathol* **128**, 755-766.
- [423] Bell WR, An Y, Kageyama Y, English C, Rudow GL, Pletnikova O, Thambisetty M, O'Brien R, Moghekar AR, Albert MS, Rabins PV, Resnick SM, Troncoso JG (2019) Neuropathologic, genetic, and longitudinal cognitive profiles in primary age-related tauopathy (PART) and Alzheimer's disease. *Alzheimers Dement* **15**, 8-16.
- [424] Jellinger KA, Alafuzoff I, Attems J, Beach TG, Cairns NJ, Crary JF, Dickson DW, Hof PR, Hyman BT, Jack CR Jr., Jicha GA, Knopman DS, Kovacs GG, Mackenzie IR, Masliah E, Montine TJ, Nelson PT, Schmitt F, Schneider JA, Serrano-Pozo A, Thal DR, Toledo JB, Trojanowski JQ, Troncoso JC, Vonsattel JP, Wisniewski T (2015) PART, a distinct tauopathy, different from classical sporadic Alzheimer disease. *Acta Neuropathol* **129**, 757-762.
- [425] Tomlinson BE, Blessed G, Roth M (1968) Observations on the brains of non-demented old people. *J Neurol Sci* **7**, 331-356.
- [426] Jellinger KA, Bancher C (1998) Senile dementia with tangles (tangle predominant form of senile dementia). *Brain Pathol* **8**, 367-376.
- [427] Neltner JH, Abner EL, Jicha GA, Schmitt FA, Patel E, Poon LW, Marla G, Green RC, Davey A, Johnson MA, Jazwinski SM, Kim S, Davis D, Woodard JL, Kryscio RJ, Van Eldik LJ, Nelson PT (2016) Brain pathologies in extreme old age. *Neurobiol Aging* **37**, 1-11.
- [428] Santa-Maria I, Haggiagi A, Liu X, Wasserscheid J, Nelson PT, Dewar K, Clark LN, Crary JF (2012) The MAPT H1 haplotype is associated with tangle-predominant dementia. *Acta Neuropathol* **124**, 693-704.
- [429] Braak H, Braak E (1995) Staging of Alzheimer's disease-related neurofibrillary changes. *Neurobiol Aging* **16**, 271-278.
- [430] McKee AC, Stern RA, Nowinski CJ, Stein TD, Alvarez VE, Daneshvar DH, Lee HS, Wojtowicz SM, Hall G, Baugh CM, Riley DO, Kubilus CA, Cormier KA, Jacobs MA, Martin BR, Abraham CR, Ikezu T, Reichard RR, Wolozin BL, Budson AE, Goldstein LE, Kowall NW,

- Cantu RC (2013) The spectrum of disease in chronic traumatic encephalopathy. *Brain* **136**, 43-64.
- [431] Corsellis JA, Brierley JB (1959) Observations on the pathology of insidious dementia following head injury. *J Ment Sci* **105**, 714-720.
- [432] Corsellis JA, Bruton CJ, Freeman-Browne D (1973) The aftermath of boxing. *Psychol Med* **3**, 270-303.
- [433] Hof PR, Knabe R, Bovier P, Bouras C (1991) Neuropathological observations in a case of autism presenting with self-injury behaviour. *Acta Neuropathol* **82**, 321-326.
- [434] Geddes JF, Vowles GH, Nicoll JA, Revesz T (1999) Neuronal cytoskeletal changes are an early consequence of repetitive head injury. *Acta Neuropathol* **98**, 171-178.
- [435] Omalu BI, DeKosky ST, Hamilton RL, Minster RL, Kamboh MI, Shakir AM, Wecht CH (2006) Chronic traumatic encephalopathy in a national football league player: Part II. *Neurosurgery* **59**, 1086-1092.
- [436] Omalu BI, DeKosky ST, Minster RL, Kamboh MI, Hamilton RL, Wecht CH (2005) Chronic traumatic encephalopathy in a national football league player. *Neurosurgery* **57**, 128-134.
- [437] Omalu BI, Bailes J, Hammers JL, Fitzsimmons RP (2010) Chronic traumatic encephalopathy, suicides and parasuicides in professional American athletes: The role of the forensic pathologist. *Am J Forensic Med Pathol* **31**, 130-132.
- [438] McKee AC, Cantu RC, Nowinski CJ, Hedley-Whyte ET, Gavett BE, Budson AE, Santini VE, Lee H-S, Kubilus CA, Stern RA (2009) Chronic traumatic encephalopathy in athletes: Progressive tauopathy after repetitive head injury. *J Neuropathol Exp Neurol* **68**, 709-735.
- [439] McKee AC, Gavett BE, Stern RA, Nowinski CJ, Cantu RC, Kowall NW, Perl DP, Hedley-Whyte ET, Price B, Sullivan C, Morin P, Lee H-S, Kubilus CA, Daneshvar DH, Wulff M, Budson AE (2010) TDP-43 proteinopathy and motor neuron disease in chronic traumatic encephalopathy. *J Neuropathol Exp Neurol* **69**, 918-929.
- [440] Gavett BE, Stern RA, Cantu RC, Nowinski CJ, McKee AC (2010) Mild traumatic brain injury: A risk factor for neurodegeneration. *Alzheimers Res Ther* **2**, 18.
- [441] Gavett BE, Stern RA, McKee AC (2011) Chronic traumatic encephalopathy: A potential late effect of sport-related concussive and subconcussive head trauma. *Clin Sports Med* **30**, 179-188.
- [442] Daneshvar DH, Baugh CM, Nowinski CJ, McKee AC, Stern RA, Cantu RC (2011) Helmets and mouth guards: The role of personal equipment in preventing sport-related concussions. *Clin Sports Med* **30**, 145-163.
- [443] Daneshvar DH, Nowinski CJ, McKee AC, Cantu RC (2011) The epidemiology of sport-related concussion. *Clin Sports Med* **30**, 1-17.
- [444] Costanza A, Weber K, Gandy S, Bouras C, Hof PR, Giannakopoulos G, Canuto A (2011) Contact sport-related chronic traumatic encephalopathy in the elderly: Clinical expression and structural substrates. *Neuropathol Appl Neurobiol* **37**, 570-584.
- [445] Stern RA, Riley DO, Daneshvar DH, Nowinski CJ, Cantu RC, McKee AC (2011) Long-term consequences of repetitive brain trauma: Chronic traumatic encephalopathy. *PM R* **3**, S460-S467.
- [446] Goldstein LE, Fisher AM, Tagge CA, Zhang XL, Velisek L, Sullivan JA, Upreti C, Kracht JM, Ericsson M, Wojnarowicz MW, Golettiani CJ, Maglakelidze GM, Casey N, Moncaster JA, Minaeva O, Moir RD, Nowinski CJ, Stern RA, Cantu RC, Geiling J, Blusztajn JK, Wolozin BL, Ikezu T, Stein TD, Budson AE, Kowall NW, Chargin D, Sharon A, Saman S, Hall GF, Moss WC, Cleveland RO, Tanzi RE, Stanton PK, Mckee AC (2012) Chronic traumatic encephalopathy in blast-exposed military veterans and a blast neurotrauma mouse model. *Sci Transl Med* **4**, 134ra60.
- [447] Saing T, Dick M, Nelson PT, Kim RC, Cribbs DH, Head E (2012) Frontal cortex neuropathology in dementia pugilistica. *J Neurotrauma* **29**, 1054-1070.
- [448] Smith DH, Johnson VE, Stewart W (2013) Chronic neuropathologies of single and repetitive TBI: Substrates of dementia? *Nat Rev Neurol* **9**, 211-221.
- [449] Sponheim SR, McGuire KA, Kang SS, Davenport ND, Aviyente S, Bernat EM, Lim KO (2011) Evidence of disrupted functional connectivity in the brain after combat-related blast injury. *Neuroimage* **54**(Suppl 1), S21-S29.
- [450] McKee AC, Cairns NJ, Dickson DW, Folkerth RD, Keene CD, Litvan I, Perl DP, Stein TD, Vonsattel JP, Stewart W, Tripodis Y, Cray JF, Bieniek KF, Dams-O'Connor K, Alvarez VE, Gordon WA, group TC (2016) The first NINDS/NIBIB consensus meeting to define neuropathological criteria for the diagnosis of chronic traumatic encephalopathy. *Acta Neuropathol* **131**, 75-86.
- [451] Vile AR, Atkinson L (2017) Chronic traumatic encephalopathy: The cellular sequela to repetitive brain injury. *J Clin Neurosci* **41**, 24-29.
- [452] Kernie SG, Parent JM (2010) Forebrain neurogenesis after focal Ischemic and traumatic brain injury. *Neurobiol Dis* **37**, 267-274.
- [453] Schmidt ML, Zhukareva V, Newell KL, Lee V, Trojanowski J (2001) Tau isoform profile and phosphorylation state in dementia pugilistica recapitulate Alzheimer's disease. *Acta Neuropathol* **101**, 518-524.
- [454] McKee AC, Daneshvar DH, Alvarez VE, Stein TD (2014) The neuropathology of sport. *Acta Neuropathol* **127**, 29-51.
- [455] Moir RD, Lathe R, Tanzi RE (2018) The antimicrobial protection hypothesis of Alzheimer's disease. *Alzheimers Dement* **14**, 1602-1614.
- [456] Lathe R, Sapronova A, Kotelevtsev Y (2014) Atherosclerosis and Alzheimer's diseases with a common cause? Inflammation, oxysterols, vasculature. *BMC Geriatr* **14**, 36.
- [457] Jamieson GA, Maitland NJ, Wilcock GK, Craske J, Itzhaki RF (1991) Latent herpes simplex virus type 1 in normal and Alzheimer's disease brains. *J Med Virol* **33**, 224-227.
- [458] De Chiara G, Marcocci ME, Sgarbanti R, Civitelli L, Ripoli C, Piacentini R, Garaci E, Grassi C, Palamara AT (2012) Infectious agents and neurodegeneration. *Mol Neurobiol* **46**, 614-638.
- [459] Itzhaki RF (2014) Herpes simplex virus type 1 and Alzheimer's disease: Increasing evidence for a major role of the virus. *Front Aging Neurosci* **6**, 202.
- [460] Balin BJ, Hudson AP (2014) Etiology and pathogenesis of late-onset Alzheimer's disease. *Curr Allergy Asthma Rep* **14**, 417.
- [461] Miklossy J (2015) Historic evidence to support a causal relationship between spirochetal infections and Alzheimer's disease. *Front Aging Neurosci* **7**, 46.
- [462] Soscia SJ, Kirby JE, Washicosky KJ, Tucker SM, Ingelsson M, Hyman B, Burton MA, Goldstein LE, Duong S, Tanzi RE, Moir RD (2010) The Alzheimer's disease-associated amyloid β -protein is an antimicrobial peptide. *PLoS One* **5**, e9505.

- [463] Kumar DKV, Choi HS, Washicosky KJ, Eimer WA, Tucker S, Ghofrani J, Lefkowitz A, Mccoll G, Goldstein LE, Tanzi RE, Moir RD (2016) Amyloid- β peptide protects against microbial infection in mouse and worm models of Alzheimer's disease. *Sci Transl Med* **8**, 340ra72.
- [464] White MR, Kandel R, Tripathi S, Condon D, Qi L, Taubenberger J, Hartshorn KL (2014) Alzheimer's associated β -amyloid protein inhibits influenza A virus and modulates viral interactions with phagocytes. *PLoS One* **9**, e101364.
- [465] Bourgade K, Garneau H, Giroux G, Le Page AY, Bocti C, Dupuis G, Frost EH, Fülöp Jr. T (2015) β -Amyloid peptides display protective activity against the human Alzheimer's disease-associated herpes simplex virus-1. *Biogerontology* **16**, 85-98.
- [466] Bourgade K, Le Page A, Bocti C, Witkowski JM, Dupuis G, Frost EH, Fülöp Jr T (2016) Protective effect of amyloid- β peptides against herpes simplex virus-1 infection in a neuronal cell culture model. *J Alzheimers Dis* **50**, 1227-1241.
- [467] Eimer WA, Vijaya Kumar DK, Navalpur Shanmugam NK, Rodriguez AS, Mitchell T, Washicosky KJ, György B, Breakefield XO, Tanzi RE, Moir RD (2018) Alzheimer's disease-associated β -amyloid is rapidly seeded by herpes viridae to protect against brain infection. *Neuron* **99**, 56.e3-63.e3.
- [468] Piacentini R, De Chiara G, Li Puma DD, Ripoli C, Marcocci ME, Garaci E, Palamara AT, Grassi C (2014) HSV-1 and Alzheimer's disease: More than a hypothesis. *Front Pharmacol* **5**, 97.
- [469] Duarte LF, Fariás MA, Álvarez DM, Bueno SM, Riedel CA, González PA (2019) Herpes simplex virus type 1 infection of the central nervous system: Insights into proposed interrelationships with neurodegenerative disorders. *Front Cell Neurosci* **13**, 46.
- [470] Itzhaki RF, Lathe R, Balin BJ, Ball MJ, Bearer EL, Braak H, Bullido MJ, Carter C, Clerici M, Cosby SL, Del Tredici K, Field H, Fulop T, Grassi C, Griffin WST, Haas J, Hudson AP, Kamer AR, Kell DB, Licastro F, Letenneur L, Lövhheim H, Mancuso R, Miklossy J, Otth C, Palamara AT, Perry G, Preston C, Pretorius E, Strandberg T, Tabet N, Taylor-Robinson SD, Whittum-Hudson JA (2016) Microbes and Alzheimer's disease. *J Alzheimers Dis* **51**, 979.
- [471] Wozniak MA, Mee AP, Itzhaki RF (2009) Herpes simplex virus type 1 DNA is located within Alzheimer's disease amyloid plaques. *J Pathol* **217**, 131-138.
- [472] Lövhheim H, Gilthorpe J, Adolfsson R, Elgh F (2014) Reactivated herpes simplex infection increases the risk of Alzheimer's disease. *Alzheimers Dement* **11**, 593-599.
- [473] Letenneur L, Pérès K, Fleury H, Garrigue I, Barberger-Gateau P, Helmer C, Orgogozo J-M, Gauthier S, Dartigues J-F (2008) Seropositivity to herpes simplex virus antibodies and risk of Alzheimer's disease: A population-based cohort study. *PLoS One* **3**, e3637.
- [474] Kobayashi N, Nagata T, Shinagawa S, Oka N, Shimada K, Shimizu A, Tatebayashi Y, Yamada H, Nakayama K, Kondo K, Tatebayashi Y, Yamada H, Nakayama K, Kondo K (2013) Increase in the IgG avidity index due to herpes simplex virus type 1 reactivation and its relationship with cognitive function in amnesic mild cognitive impairment and Alzheimer's disease. *Biochem Biophys Res Commun* **430**, 907-911.
- [475] Mancuso R, Baglio F, Cabinio M, Calabrese E, Hernis A, Nemni R, Clerici M (2014) Titers of herpes simplex virus type 1 antibodies positively correlate with grey matter volumes in Alzheimer's disease. *J Alzheimers Dis* **38**, 741-745.
- [476] Itzhaki RF, Lin WR, Shang D, Wilcock GK, Faragher B, Jamieson GA (1997) Herpes simplex virus type 1 in brain and risk of Alzheimer's disease. *Lancet* **349**, 241-244.
- [477] Itzhaki RF, Lin WR (1998) Herpes simplex virus type 1 in brain and the type 4 allele of the apolipoprotein E gene are a combined risk factor for Alzheimer's disease. *Biochem Soc Trans* **26**, 273-277.
- [478] Zambrano A, Solis L, Salvadores N, Cortés M, Lerchundi R, Otth C (2008) Neuronal cytoskeletal dynamic modification and neurodegeneration induced by infection with herpes simplex virus type 1. *J Alzheimers Dis* **14**, 259-269.
- [479] Wozniak MA, Frost AL, Itzhaki RF (2009) Alzheimer's disease specific tau phosphorylation is induced by herpes simplex virus type 1. *J Alzheimers Dis* **16**, 341-350.
- [480] Martin C, Aguila B, Araya P, Vio K, Valdivia S, Zambrano A, Concha MI, Otth C (2013) Inflammatory and neurodegeneration markers during asymptomatic HSV-1 reactivation. *J Alzheimers Dis* **39**, 849-859.
- [481] Gnann JW, Whitley RJ (2017) Herpes simplex encephalitis: An update. *Curr Infect Dis Rep* **19**, 13.
- [482] Ball MJ (1982) Limbic predilection in Alzheimer dementia: Is reactivated herpesvirus involved? *Can J Neurol Sci* **9**, 303-306.
- [483] Damasio AR, Van Hoesen GW (1985) The limbic system and the localisation of herpes simplex encephalitis. *J Neurol Neurosurg Psychiatry* **48**, 297-301.
- [484] Caparros-Lefebvre D, Girard-Buttaz I, Reboul S, Lebert F, Cabaret M, Verier A, Steinling M, Pruvo JP, Petit H (1996) Cognitive and psychiatric impairment in herpes simplex virus encephalitis suggest involvement of the amygdalo-frontal pathways. *J Neurol* **243**, 248-256.
- [485] Beffert U, Bertrand P, Champagne D, Gauthier S, Poirier J (1998) HSV-1 in brain and risk of Alzheimer's disease. *Lancet* **351**, 1330-1331.
- [486] Wu HM, Huang CC, Chen SH, Liang YC, Tsai JJ, Hsieh CL, Hsu K-S (2003) Herpes simplex virus type 1 inoculation enhances hippocampal excitability and seizure susceptibility in mice. *Eur J Neurosci* **18**, 3294-3304.
- [487] Taylor SW, Lee DH, Jackson AC (2007) Herpes simplex encephalitis presenting with exclusively frontal lobe involvement. *J Neurovirol* **13**, 477-481.
- [488] Ando Y, Kitayama H, Kawaguchi Y, Koyanagi Y (2008) Primary target cells of herpes simplex virus type 1 in the hippocampus. *Microbes Infect* **10**, 1514-1523.
- [489] Jamieson GA, Maitland NJ, Wilcock GK, Yates CM, Itzhaki RF (1992) Herpes simplex virus type 1 DNA is present in specific regions of brain from aged people with and without senile dementia of the Alzheimer type. *J Pathol* **167**, 365-368.
- [490] De Chiara G, Piacentini R, Fabiani M, Mastrodonato A, Marcocci ME, Limongi D, Napolitano G, Protto V, Coluccio P, Celestino I, Li Puma DD, Grassi C, Palamara AT (2019) Recurrent herpes simplex virus-1 infection induces hallmarks of neurodegeneration and cognitive deficits in mice. *PLoS Pathog* **15**, e1007617.
- [491] Menendez CM, Jinkins JK, Carr DJJ (2016) Resident T cells are unable to control herpes simplex virus-1 activity in the brain ependymal region during latency. *J Immunol* **197**, 1262-1275.
- [492] Yao H-W, Ling P, Tung Y-Y, Hsu S-M, Chen S-H (2014) In vivo reactivation of latent herpes simplex virus 1 in mice

- can occur in the brain before occurring in the trigeminal ganglion. *J Virol* **88**, 11264-11270.
- [493] Mori I, Nishiyama Y, Yokochi T, Kimura Y (2005) Olfactory transmission of neurotropic viruses. *J Neurovirol* **11**, 129-137.
- [494] Gillet L, Frederico B, Stevenson PG (2015) Host entry by gamma-herpesviruses - lessons from animal viruses? *Curr Opin Virol* **15**, 34-40.
- [495] Little CS, Bowe A, Lin R, Litsky J, Fogel RM, Balin BJ, Fresa-Dillon KL (2005) Age alterations in extent and severity of experimental intranasal infection with Chlamydia pneumoniae in BALB/c mice. *Infect Immun* **73**, 1723-1734.
- [496] Conrady CD, Zheng M, van Rooijen N, Drevets DA, Royer D, Alleman A, Carr DJ (2013) Microglia and a functional type I IFN pathway are required to counter HSV-1-driven brain lateral ventricle enlargement and encephalitis. *J Immunol* **190**, 2807-2817.
- [497] Braun E, Zimmerman T, Hur TB, Reinhartz E, Fellig Y, Panet A, Steiner I (2006) Neurotropism of herpes simplex virus type 1 in brain organ cultures. *J Gen Virol* **87**, 2827-2837.
- [498] Chucair-Elliott AJ, Conrady C, Zheng M, Kroll CM, Lane TE, Carr DJ (2014) Microglia-induced IL-6 protects against neuronal loss following HSV-1 infection of neural progenitor cells. *Glia* **62**, 1418-1434.
- [499] Rotschafer JH, Hu S, Little M, Erickson M, Low WC, Cheeran MC (2013) Modulation of neural stem/progenitor cell proliferation during experimental herpes simplex encephalitis is mediated by differential FGF-2 expression in the adult brain. *Neurobiol Dis* **58**, 144-155.
- [500] Li Puma DD, Piacentini R, Leone L, Gironi K, Marcocci ME, De Chiara G, Palamara AT, Grassi C (2019) Herpes simplex virus type-1 infection impairs adult hippocampal neurogenesis via amyloid- β protein accumulation. *Stem Cells* **37**, 1467-1480.
- [501] Li Puma DD, Piacentini R, Grassi C (2021) Does impairment of adult neurogenesis contribute to pathophysiology of Alzheimer's disease? A still open question. *Front Mol Neurosci* **13**, 578211.
- [502] Quiroz YT, Sperling RA, Norton DJ, Baena A, Arboleda-Velasquez JF, Cosio D, Schultz A, Lapoint M, Guzman-Velez E, Miller JB, Kim LA, Chen K, Tariot PN, Lopera F, Reiman EM, Johnson KA (2018) Association between amyloid and tau accumulation in young adults with autosomal dominant Alzheimer disease. *JAMA Neurol* **75**, 548-556.
- [503] Wang L, Benzinger TL, Su Y, Christensen J, Friedrichsen K, Aldea P, McConathy J, Cairns NJ, Fagan AM, Morris JC, Ances BM (2016) Evaluation of tau imaging in staging Alzheimer disease and revealing interactions between β -amyloid and tauopathy. *JAMA Neurol* **73**, 1070-1077.
- [504] Bischof GN, Jessen F, Fliessbach K, Dronse J, Hammes J, Neumaier B, Onur O, Fink GR, Kukulja J, Drzezga A, van Eimeren T, for the Alzheimer's Disease Neuroimaging Initiative (2016) Impact of tau and amyloid burden on glucose metabolism in Alzheimer's disease. *Ann Clin Transl Neurol* **3**, 934-939.
- [505] Hanseeuw BJ, Betensky RA, Schultz AP, Papp KV, Mormino EC, Sepulcre J, Bark JS, Cosio DM, LaPoint M, Chhatwal JP, Rentz DM, Sperling RA, Johnson KA (2017) Fluorodeoxyglucose metabolism associated with tau-amyloid interaction predicts memory decline. *Ann Neurol* **81**, 583-596.
- [506] Adams JN, Lockhart SN, Li L, Jagust WJ (2018) Relationships between tau and glucose metabolism reflect Alzheimer's disease pathology in cognitively normal older adults. *Cereb Cortex* **29**, 1997-2009.
- [507] Jacobs HIL, Hedden T, Schultz AP, Sepulcre J, Perea RD, Amariglio RE, Papp KV, Rentz DM, Sperling RA, Johnson KA (2018) Structural tract alterations predict downstream tau accumulation in amyloid positive older individuals. *Nat Neurosci* **21**, 424-431.
- [508] Villemagne VL, Doré V, Burnham SC, Masters CL, Rowe CC (2018) Imaging tau and amyloid- β proteinopathies in Alzheimer disease and other conditions. *Nat Rev Neurol* **14**, 225-236.
- [509] Sepulcre J, Schultz AP, Sabuncu M, Gomez-Isla T, Chhatwal J, Becker A, Sperling R, Johnson KA (2016) *In vivo* tau, amyloid, and gray matter profiles in the aging brain. *J Neurosci* **36**, 7364-7374.
- [510] Lockhart SL, Schöll M, Baker SL, Ayakta N, Swinerton KN, Bell RK, Mellinger TJ, Shah VD, O'Neil JP, Janabi M, Jagust WJ (2017) Amyloid and tau PET demonstrate region-specific associations in normal older people. *Neuroimage* **150**, 191-199.
- [511] Demars MP, Hollands C, Zhao KDT, Lazarov O (2013) Soluble amyloid precursor protein-a rescues age-linked decline in neural progenitor cell proliferation. *Neurobiol Aging* **34**, 2431-2440.
- [512] Aizawa K, Ageyama N, Terao K, Hisatsune T (2011) Primate-specific alterations in neural stem/progenitor cells in the aged hippocampus. *Neurobiol Aging* **32**, 140-150.
- [513] Leuner B, Kozorovitskiy Y, Gross CG, Gould E (2007) Diminished adult neurogenesis in the marmoset brain precedes old age. *Proc Natl Acad Sci U S A* **104**, 17169-17173.
- [514] Bergmann O, Spalding KL, Frisen J (2015) Adult neurogenesis in humans. *Cold Spring Harb Perspect Biol* **7**, a018994.
- [515] Fu H, Hardy J, Duff KE (2018) Selective vulnerability in neurodegenerative diseases. *Nat Neurosci* **21**, 1350-1358.
- [516] Mattsson N, Schott JM, Hardy J, Turner MR, Zetterberg H (2016) Selective vulnerability in neurodegeneration: Insights from clinical variants of Alzheimer's disease. *J Neurol Neurosurg Psychiatry* **87**, 1000-1004.
- [517] Mrdjen D, Fox EJ, Bukhari SA, Montine KS, Bendall SC, Montine TJ (2019) The basis of cellular and regional vulnerability in Alzheimer's disease. *Acta Neuropathol* **138**, 729-749.
- [518] Brothers HM, Gosztyla ML, Robinson SR (2018) The physiological roles of amyloid- β peptide hint at new ways to treat Alzheimer's disease. *Front Aging Neurosci* **10**, 118.
- [519] Takuma H, Arawaka S, Mori H (2003) Isoforms changes of tau protein during development in various species. *Dev Brain Res* **142**, 121-127.
- [520] Arendt T, Stieler J, Ueberham U (2017) Is sporadic Alzheimer's disease a developmental disorder? *J Neurochem* **143**, 396-408.
- [521] Garcia-Verdugo JM, Ferron S, Flames N, Collado L, Desfilis E, Font E (2002) The proliferative ventricular zone in adult vertebrates: A comparative study using reptiles, birds, and mammals. *Brain Res Bull* **57**, 765-775.
- [522] Sawada M, Sawamoto K (2013) Mechanisms of neurogenesis in the normal and injured adult brain. *Keio J Med* **62**, 13-28.
- [523] Paredes MF, Sorrells SF, Garcia-Verdugo JM, Alvarez-Buylla A (2016) Brain size and limits to adult neurogenesis. *J Comp Neurol* **524**, 646-664.

- [524] Christensen K, Doblhammer G, Rau R, Vaupel JW (2009) Ageing populations: The challenges ahead. *Lancet* **374**, 1196-208.
- [525] Sanai N, Tramontin AD, Quiñones-Hinojosa A, Barbaro NM, Gupta N, Kunwar S, Lawton MT, McDermott MW, Parsa AT, Verdugo JM-G, Berger MS, Alvarez-Buylla A (2004) Unique astrocyte ribbon in adult human brain contains neural stem cells but lacks chain migration. *Nature* **427**, 740-744.
- [526] Palmer TD, Schwartz PH, Taupin P, Kaspar B, Stein SA, Gage FH (2001) Cell culture. Progenitor cells from human brain after death. *Nature* **411**, 42-43.
- [527] Nunes MC, Singh Roy N, Keyoung HM, Goodman RR, McKhann II G, Jiang L, Kang J, Nedergaard M, Goldman SA (2003) Identification and isolation of multipotential neural progenitor cells from the subcortical white matter of the adult human brain. *Nat Med* **9**, 439-447.
- [528] Eriksson PS, Perfilieva E, Björk-Eriksson T, Alborn A-M, Nordborg C, Peterson DA, Gage FH (1998) Neurogenesis in the adult human hippocampus. *Nat Med* **4**, 1313-1317.
- [529] Choi SH, Tanzi RE (2019) Is Alzheimer's disease a neurogenesis disorder? *Cell Stem Cell* **25**, 7-8.
- [530] Kempermann G, Gage FH, Aigner L, Song H, Curtis MA, Thuret S, Kuhn HG, Jessberger S, Frankland PW, Cameron HA, Gould E, Hen R, Abrous DN, Toni N, Schinder AF, Zhao X, Lucassen PJ, Frisén J (2018) Human adult neurogenesis: Evidence and remaining questions. *Cell Stem Cell* **23**, 25-30.