Cortical Thickness and Its Association with Clinical Cognitive and Neuroimaging Markers in Cerebral Amyloid Angiopathy

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Abstract

Background: Cerebral amyloid angiopathy (CAA) contributes to brain neurodegeneration and cognitive decline, but the relationship between these two processes is incompletely understood.

Objective: The purpose of this study is to examine cortical thickness and its association with cognition and neurodegenerative biomarkers in CAA.

Methods: Data were collected from the Functional Assessment of Vascular Reactivity study and the Calgary Normative Study. In total, 48 participants with probable CAA, 72 cognitively normal healthy controls, and 24 participants with mild dementia due to AD were included. Participants underwent an MRI scan, after which global and regional cortical thickness measurements were obtained using FreeSurfer. General linear models, adjusted for age and sex, were used to compare cortical thickness globally and in an AD signature region.

Results: Global cortical thickness was lower in CAA compared to healthy controls (mean difference (MD) –0.047 mm, 95% confidence interval (CI) –0.088, –0.005, \( p = 0.03 \)), and lower in AD compared to CAA (MD –0.104 mm, 95% CI –0.165, –0.043, \( p = 0.001 \)). In the AD signature region, cortical thickness was lower in CAA compared to healthy controls (MD –0.07 mm, 95% CI –0.13 to –0.01, \( p = 0.02 \)). Within the CAA group, lower cortical thickness was associated with lower memory scores (R² = 0.10; \( p = 0.05 \)) and higher white matter hyperintensity volume (R² = 0.09, \( p = 0.04 \)).

Conclusion: CAA contributes to neurodegeneration in the form of lower cortical thickness, and this could contribute to cognitive decline. Regional overlap with an AD cortical atrophy signature region suggests that co-existing AD pathology may contribute to lower cortical thickness observed in CAA.

Keywords: Alzheimer’s disease, cerebral amyloid angiopathy, cortical thickness, cognition, MRI, neurodegeneration

INTRODUCTION

Cerebral amyloid angiopathy (CAA) is a cerebral small vessel disease characterized by amyloid-β (Aβ) accumulation in the small vessels and leptomeninges in the brain [1]. It is a leading cause of intracerebral hemorrhage (ICH) and also contributes to cognitive decline [2]. The Boston Criteria are pathologically validated for the diagnosis of CAA during life, based on clinical [3] and magnetic resonance imaging (MRI) findings including the presence of ICH and cortical superficial siderosis (cSS) [4].
Previous studies suggest that lower brain volume, a hallmark of neurodegenerative diseases, is seen in CAA, and may contribute to the development of dementia for which CAA patients are at risk even without new ICH [5, 6]. Recent research has also shown that CAA is associated with neurodegeneration in the form of thinner cerebral cortex [7, 8], a biomarker of neurodegeneration in related diseases such as Alzheimer’s disease (AD) [9, 10]. However, regional distributions of cortical thinning, and the overall relationship of cortical thickness with cognition and other biomarkers in CAA is not yet fully understood. The aim of this study was therefore to: 1) examine global and regional cortical thickness in CAA; 2) determine how cortical thickness in CAA correlates with cognition; and 3) examine how cortical thickness in CAA relates to biomarkers of CAA pathology, primarily cerebral microbleed (CMB) count and white matter hyperintensity (WMH) volume.

METHODS

Study population

Data supporting the findings of this study are available for qualified researchers upon reasonable request. The data was collected from CAA, AD, and healthy control participants who consecutively enrolled in the prospective Functional Assessment of Vascular Reactivity in Small Vessel Disease (FAVR) study [11, 12], which had two study iterations. Healthy control data were collected from both FAVR and the Calgary Normative Study, which examines longitudinal changes in brain structure in healthy individuals [13, 14], CAA and AD participants were recruited from stroke, memory, and geriatric psychiatry clinics, while healthy controls were recruited from the community using print advertising and word of mouth. Testing occurred over a period from February 2010 to May 2018, with visits including a baseline MRI scan, clinical assessment, and neuropsychological testing.

CAA participants were required to have a diagnosis of probable CAA according to the validated Boston Criteria [3]. This included multiple hemorrhages restricted to lobar, cortical, or corticosubcortical regions, or a single lobar, cortical, or subcortical hemorrhage and evidence of focal disseminated superficial siderosis. CAA participants were also required to be free of symptomatic stroke within the previous year. Participants with mild dementia due to AD were diagnosed according to the National Institute on Aging and Alzheimer’s Association clinical criteria [15]. Exclusion criteria included: presence of other chronic neurological diseases, ongoing alcohol or drug abuse, not having a study partner that could provide corroborative information (not required for healthy control participants enrolled in the Calgary Normative Study), not sufficiently proficient in English, unable to undergo an MRI scan, and having a total score on the Montreal Cognitive Assessment of < 13. Healthy controls were required to have no prior history of stroke, no other central nervous system diseases, and have a Montreal Cognitive Assessment score of ≥ 25. The study was approved by the University of Calgary Conjoint Health Research Ethics Board, and participants gave written informed consent prior to study participation.

MRI acquisition and processing

Scanning for both FAVR and the Calgary Normative Study was conducted on a 3.0 Tesla MR Scanner (GE Signa VH/I or Discovery 750; GE Healthcare, Waukesha, Wisconsin) with a 12-channel head coil. The scanner was upgraded in January 2011; therefore, 27 participants underwent a scan on the GE Signa VH/I scanner while 117 had a scan on the Discovery 750 scanner. Detailed image parameters can be found in Supplementary Table 1. Participants in the Calgary Normative Study underwent the same imaging protocol as participants in the first (legacy) iteration of the FAVR study.

Cortical thickness measurements were obtained by processing 3D T1-weighed images with FreeSurfer (v6.0.0, surfer.nmr.mgh.harvard.edu) [16, 17]. Visualization of regions of thinning was achieved by aligning images to a common surface template and registering them to the Desikan-Killiany atlas, which parcellates the cortex into 34 gyral regions per hemisphere [18]. For twelve CAA participants with large residual lesions from past ICH we used results from the unaffected hemisphere only, similar to a previous study [7]. Freesurfer output was assessed visually (by AS) for quality. We excluded data from 5 CAA participants due to poor image quality (n = 4) and a missing baseline MRI scan (n = 1). Data was also excluded from one healthy control due to a missing baseline MRI scan. WMH volumes were measured on T2-weighted fluid attenuated inversion recovery images using the semi-automated software Quan
tomo (Version 1.0; Cybertrials, Inc, Calgary, Canada) [19], in accordance with consensus standards [20].
CMBs and cSS were assessed on susceptibility weighted images by a trained neurologist (EES) using previously published guidelines [21]. Images from participants with AD were reviewed for evidence of amyloid-related imaging abnormalities (ARIA) [22], while images from participants with CAA were reviewed for evidence of CAA-related inflammation [23]. Enlarged perivascular spaces (EPVS) were rated on T2-weighted images by a trained reviewer (EES) on a 4-point visual rating scale [24]. The CAA-Small Vessel Disease (CAA-SVD) score was calculated based on the number of CMBs, cSS, WMH, and EPVS, as previously described [25]. The amplitude of the blood oxygen level dependent (BOLD) signal in primary visual cortex in response to visual stimulation, a surrogate of vascular reactivity, was measured as described in a previous study [11].

Neuropsychological test battery

CAA participants completed a 1.25 h neuropsychological test battery based on harmonization standards for research on vascular cognitive impairment [26]. For each test, z-scores were derived from published norms stratified by age and then grouped into domains of memory, executive, function and processing speed. To harmonize individual tests with a national study [27], the test battery was changed in November 2016 after 37 participants were enrolled; the other 11 participants had the new battery. The memory domain score was derived from either the average of the delayed recall portion of the California Verbal Learning Test: Second Edition [28] and Rey-Osteireith Complex Figure Test [29] (first iteration of FAVR), or the delayed recall portion of the Rey Auditory Verbal Learning Test [30] and Brief Visuospatial Memory Test [31] (second iteration). The executive function domain score was based on the average of the Trail Making Test-B [32] and Control Oral Word Association Test-FAS [33], or the average of Trail Making Test-B and the Delis-Kaplan Executive Function [34] Letter Fluency tests. The processing speed domain score was based on the average of the Trail Making Test-A [32] and the Digit Symbol Coding test from the Wechsler Adult Intelligence Scale-Third Edition [35], or the Trail Making Test-A and the Digit Symbol Coding from Wechsler Adult Intelligence Scale-Fourth Edition [36].

Statistical analyses

Chi-square and Fisher’s exact test were used for categorical variables, while t-tests, one-way analyses of variance and their non-parametric equivalents (Wilcoxon rank-sum and Kruskal Wallis tests) were used for continuous variables. Pearson correlations were used to assess the relationship between cortical thickness and cognition, while Spearman rank correlations were used to assess the association between cortical thickness and WMH and CMB count. Cortical thickness was calculated as the average of the left and right hemisphere. WMH volume and hippocampal volume were expressed as a percentage of the total estimated intracranial volume to correct for intracranial size. Separate univariable analyses were conducted to determine which variables were correlated with cortical thickness, after which general linear models were built to compare thickness between the three groups. For continuous predictors, we computed adjusted partial correlation coefficients. We also compared cortical thickness in two pre-specified regions of interest: the occipital lobe (atlases regions lateral occipital, pericalcarine, cuneus, and lingual) and an AD signature region (consisting of the entorhinal cortex, fusiform, inferior parietal, inferior temporal, mid-temporal, and precuneus) as proposed by a prior study [37]. Data were analyzed using SAS version 9.4 (SAS Institute, Cary, NC). Statistical significance was set at $p \leq 0.05$, and all tests were 2-tailed.

Regional comparisons of thinning, adjusted for age and sex, were conducted using the command line group analysis stream in FreeSurfer, which utilizes a general linear model at each point on the inflated cortical surface. Data were smoothed using a 10 mm full-width at half-maximum smoothing kernel and corrected for multiple comparisons using 10,000 Monte Carlo Simulations [38]. Maps were thresholded using a liberal vertex-cluster forming threshold of $p < 0.05$ due to the exploratory nature of the study.

RESULTS

Participant characteristics

In total, the final dataset included 72 healthy controls (40 FAVR, 32 Calgary Normative Study), 48 CAA, and 24 AD participants. Participant characteristics are shown in Table 1. CAA participants were more likely to have hypertension and greater WMH volume, while AD participants had lower hippocampal volume. None of the participants with AD had ARIA. Cognitive scores across all domains were lowest in the AD group (Table 1). CAA participants presented with either ICH (19/48), transient focal
Table 1
Characteristics of the patient population

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Control (n = 72)</th>
<th>CAA (n = 48)</th>
<th>AD (n = 24)</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age</td>
<td>71.7 ± 8.2</td>
<td>74.1 ± 6.6</td>
<td>70.4 ± 7.5</td>
<td>0.10</td>
</tr>
<tr>
<td>Female, n (%)</td>
<td>50 (69%)</td>
<td>22 (46%)</td>
<td>11 (46%)</td>
<td>0.02</td>
</tr>
<tr>
<td>Education</td>
<td>15.1 ± 3.1</td>
<td>13.5 ± 2.9</td>
<td>15.8 ± 3.2</td>
<td>0.004</td>
</tr>
<tr>
<td>Hypertension, n</td>
<td>21</td>
<td>31</td>
<td>8</td>
<td>0.0004</td>
</tr>
<tr>
<td>Hypercholesterolemia, n</td>
<td>28</td>
<td>22</td>
<td>3</td>
<td>0.06</td>
</tr>
<tr>
<td>Diabetes, n</td>
<td>7</td>
<td>10</td>
<td>3</td>
<td>0.23</td>
</tr>
<tr>
<td>Smoker, n</td>
<td>2</td>
<td>2</td>
<td>1</td>
<td>0.90</td>
</tr>
<tr>
<td>WMH volume</td>
<td>0.18 (0.08–0.46)</td>
<td>1.47 (0.74–2.60)</td>
<td>0.31 (0.16–0.56)</td>
<td>&lt; 0.0001</td>
</tr>
<tr>
<td>Hippocampal volume</td>
<td>0.25 (0.22–0.27)</td>
<td>0.23 (0.22–0.27)</td>
<td>0.20 (0.19–0.23)</td>
<td>0.0007</td>
</tr>
<tr>
<td>BOLD visual response</td>
<td>2.66 (2.09–3.16)</td>
<td>1.96 (1.33–2.62)</td>
<td>2.18 (1.48–2.77)</td>
<td>0.007</td>
</tr>
<tr>
<td>Memory, z-score</td>
<td>0.61 ± 0.80</td>
<td>–0.81 ± 1.18</td>
<td>–2.07 ± 0.79</td>
<td>&lt; 0.0001</td>
</tr>
<tr>
<td>Executive function, z-score</td>
<td>0.14 ± 0.92</td>
<td>–1.30 ± 1.05</td>
<td>–1.49 ± 0.88</td>
<td>&lt; 0.0001</td>
</tr>
<tr>
<td>Processing speed, z-score</td>
<td>0.59 ± 0.87</td>
<td>–1.05 ± 1.04</td>
<td>–1.41 ± 1.06</td>
<td>&lt; 0.0001</td>
</tr>
<tr>
<td>Occipital lobe thickness</td>
<td>2.05 ± 0.10</td>
<td>2.04 ± 0.12</td>
<td>2.00 ± 0.15</td>
<td>0.29</td>
</tr>
<tr>
<td>AD signature thickness</td>
<td>2.72 ± 0.13</td>
<td>2.63 ± 0.15</td>
<td>2.44 ± 0.24</td>
<td>&lt; 0.0001</td>
</tr>
<tr>
<td>Global thickness</td>
<td>2.41 ± 0.10</td>
<td>2.35 ± 0.10</td>
<td>2.27 ± 0.16</td>
<td>&lt; 0.0001</td>
</tr>
</tbody>
</table>

AD, Alzheimer’s disease; CAA, cerebral amyloid angiopathy; WMH, white matter hyperintensities; BOLD, blood oxygen level dependent. Values are represented as mean ± standard deviation, median (interquartile range), or number. WMH and hippocampal volume are expressed as a percent of the total estimated intracranial volume.

*Only available for the 79 participants in the first iteration of the FAVR study. †Neuropsychiatric data only available for healthy controls enrolled in FAVR. ‡Available for 45 CAA participants. §Available for 23 AD participants.

Table 2
CAA biomarker and CAA-SVD score distribution among CAA participants

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>N or median [25th percentile, 75th percentile]</th>
</tr>
</thead>
<tbody>
<tr>
<td>cSS</td>
<td>29</td>
</tr>
<tr>
<td>CMB</td>
<td>21 [5, 51]</td>
</tr>
<tr>
<td>CAA-SVD score (n = 40)*</td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>1</td>
<td>3</td>
</tr>
<tr>
<td>2</td>
<td>7</td>
</tr>
<tr>
<td>3</td>
<td>12</td>
</tr>
<tr>
<td>4</td>
<td>10</td>
</tr>
<tr>
<td>5</td>
<td>11</td>
</tr>
<tr>
<td>6</td>
<td>13</td>
</tr>
<tr>
<td>Centrum semiovale EPVS category (n = 44)*</td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>1</td>
<td>16</td>
</tr>
<tr>
<td>2</td>
<td>18</td>
</tr>
<tr>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>4</td>
<td>1</td>
</tr>
<tr>
<td>CAA Syndrome</td>
<td></td>
</tr>
<tr>
<td>ICH</td>
<td>19</td>
</tr>
<tr>
<td>Cognitive Symptoms</td>
<td>12</td>
</tr>
<tr>
<td>TFNE</td>
<td>17</td>
</tr>
</tbody>
</table>

cSS, cortical superficial siderosis; CMB, cerebral microbleeds; CAA-SVD, cerebral amyloid angiopathy-small vessel disease; EPVS, enlarged perivascular spaces; ICH, intracerebral hemorrhage; TFNE, transient focal neurological episodes. *Interpretable susceptibility weighted imaging was unavailable for some participants due to motion artifacts.

Cortical thickness comparison

Global cortical thickness was lowest in AD, intermediate in CAA, and highest in controls (p < 0.0001, Table 1). Thickness in the AD signature region also differed across groups (p < 0.0001, Table 1). Age and sex were significantly associated with cortical thickness in univariable analyses (Table 3), therefore they were entered into the multivariable model. Adjusting for age and sex and compared to the control group, thickness was lower in CAA (mean difference [MD] –0.047 mm, 95% confidence interval [CI] –0.088 to –0.005, p = 0.03) and AD participants (MD –0.143 mm, 95% CI –0.195 to –0.091, p < 0.0001). Results were unchanged when we excluded the 12 CAA participants with ICH (MD –0.048 mm, 95% CI –0.094 to –0.002, p = 0.04). The addition of scanner type as an additional variable did not have an effect on the results (MD from controls: CAA –0.051 mm, 95% CI –0.101 to 0.106, p = 0.02; AD –0.162 mm, 95% CI –0.215 to –0.108, p < 0.0001). In a multivariable model comparing only AD and CAA participants, AD participants had thinner cortex (MD –0.104 mm, 95% CI –0.165 to –0.043, p = 0.001). In the AD signature region, after adjusting for age and sex, thickness was lower in AD versus controls (MD –0.28 mm, 95% CI –0.35 to –0.21, p < 0.0001) and in CAA versus controls (MD –0.07 mm, 95% CI –0.13 to –0.01, p = 0.02). A separate multivariable analysis showed thinner cortex in the AD signature region in the AD group compared to the CAA group (MD –0.22 mm,
Within the CAA group, thinner cortex was associated with lower z-scores in the memory domain ($r = 0.35$, $p = 0.02$), but not executive function ($r = 0.12$, $p = 0.43$) or processing speed ($r = 0.02$, $p = 0.88$). The association between thickness and memory remained significant after adjusting for sex and education ($R^2 = 0.10$, $p = 0.05$). Thinner cortex in the AD signature region in the CAA group was even more closely associated with lower z-scores ($r = 0.40$, $p = 0.006$), an association that remained significant after additionally adjusting for sex and education ($R^2 = 0.26$, $p = 0.002$). A similar association between thickness in the AD signature region and memory was also found in the AD group ($R^2 = 0.28$, $p = 0.01$), but not the healthy control group ($R^2 = 0.10$, $p = 0.16$), after adjusting for sex and education. As a sensitivity analysis, we tested whether the association of global thickness with memory in CAA differed between the old neuropsychological battery (37/48) and the new battery (11/48) but failed to find evidence of a difference (interaction $p$-value 0.51).

**Associations of cortical thickness with CAA biomarkers**

Higher WMH volume was associated with thinner cortex in univariable analysis ($r = -0.33$, $p = 0.02$) within CAA. This association remained after adjustment for age and sex ($R^2 = 0.09$, $p = 0.04$). No associations were found between thickness and other CAA imaging biomarkers and characteristics (Table 2). No associations were found between BOLD visual response and occipital lobe thickness ($r = 0.13$, $p = 0.44$) or global thickness ($r = 0.23$, $p = 0.17$) within CAA.

**DISCUSSION**

In this study we found that cortical thickness in CAA was intermediate between healthy controls and AD participants and that thinner cortex was associated with higher WMH volume and lower memory scores in CAA participants. In an AD signature region (comprising entorhinal, fusiform, inferior parietal, inferior temporal, mid-temporal, and precuneus), CAA as well as AD participants had thinner cortex compared to controls.

Areas of thinner cortex in CAA compared to controls were found in the frontal, left parietal, and right temporal lobes. We also found thinner cerebral cortex in CAA patients in an AD signature region derived and validated in a previous study, based on the regions that are affected early on in AD-related cognitive disorders. This pattern of cortical thinness is consistent with the predilection for vascular amyloid deposition in more posterior brain regions. However,
not all posterior regions were affected. We failed to find significantly thinner cortex in a pre-specified region of interest in the primary visual cortex, even though vascular amyloid is deposited there and we have previously found reduced BOLD activation in this region. Thus, there is an imperfect correlation between regions of amyloid deposition and regions of thinner cortex. The reasons for this are not entirely clear. The finding that cortex was thinner in the AD signature region suggests the possibility that co-morbid AD pathology [39], which is common in CAA even in persons without dementia, may be contributing in a synergistic manner to thinning of the cerebral cortex in persons with CAA.

We found that thinner cortex was associated with the memory cognitive domain but not with executive function or processing speed. This association with memory was even stronger when considering only the AD signature region. In our prior work [40], we showed that all three cognitive domains were affected in CAA, although compared to AD there was relatively greater impairment in executive function and processing speed in CAA. Therefore, the lack of association between cerebral cortex thinness and lower executive function and processing speed was somewhat unexpected. It is possible that the dysfunction in executive function and processing speed may be more related to subcortical white matter disconnection, microinfarction, the effects of prior ICH, and altered reactivity rather than cortical neurodegeneration.

Of the other CAA markers, we found that thinner cortex was associated with higher WMH volume. However, we failed to find associations with cerebral microbleed (CMB) count, CAA SVD score, or occipital BOLD activation. In another recently pub-
lished study [12], we failed to find an association between cortical thinness and mean white matter diffusivity in a skeletonized white matter region of interest. The association with WMH is plausible because WMH has been associated with cerebral atrophy in general population studies. Mechanisms by which WMH could cause cerebral atrophy include axonal loss with secondary apoptosis of cerebral neurons [41], or it may be a surrogate for more widespread ischemic changes or low cerebral perfusion. Because WMH are much more widespread in CAA than AD, it also suggests that there are CAA-specific pathways of neurodegeneration and that not all neurodegeneration in persons with CAA is due to accompanying AD pathology.

Previous studies have found that thinning of the cerebral cortex is seen in most neurodegenerative diseases as well as in cerebral small disease in the general population [42]. Histopathologically, cortical thinning associated with advancing age corresponds to neuronal and dendritic shrinkage, as well as neuronal loss [43]. Alterations in genes expressed by pyramidal cells, astrocytes, and microglia underlie these changes, and also account for some of the cortical thinning seen in AD [44]. However, only a few studies have previously looked at cerebral cortical thickness or brain volumes in persons with CAA. A study of sporadic and hereditary Dutch-type CAA also found intermediate global cortical thinness in CAA between AD and controls, along with regional overlaps, similar to this study [7]. These findings, along with our own, are in contrast to a prior study with a higher number of patients with CAA with dementia, which found no average difference in thickness between CAA and AD [8]. These findings suggest that cortical thickness in CAA could be related to the stage of cognitive dysfunction. It is also possible that patients with CAA and dementia have a higher burden of co-morbid AD pathology, such that their cortical thickness is similar to patients with pure AD [7]. Additional factors that could contribute to cortical thinning and associated cognitive impairment in CAA, warranting additional future study, include the effects of cerebral microinfarcts [5], hemorrhagic lesions including cSS [45], impaired vascular reactivity [7], and neuronal apoptosis as a consequence of axonal damage from white matter lesions [41].

Strengths and limitations

A strength of our study is that we included a diverse and clinically representative cohort of people with CAA and AD, along with using a neuroimaging processing pipeline that is reliable across different scanner manufacturers [46]. Limitations included the use of different cognitive tests for CAA participants enrolled in the two different study iterations of FAVR, although we did not find an interaction effect or difference between the two different cognitive batteries. We also did not include measurements of Aβ distribution by positron emission tomography, which could have been used to determine whether amyloid deposits colocalized with thinner cortex. However, a limitation of using amyloid PET is that it does not discriminate well between vascular and parenchymal Aβ. In the CAA group, thinning was found in regions such as the medial frontal and temporal lobes, areas that have been found in amyloid imaging studies to contain high loads of vascular Aβ [47, 48]. Because a validated highly specific in vivo molecular biomarker of CAA does not exist, it is difficult to rule out the role of parenchymal AD pathology in the cortical thinness observed in CAA. The sample size was relatively small; however, the sample size was sufficient to reproduce known differences between AD and controls and the association of AD-associated atrophy with memory impairment.

CONCLUSIONS

This study suggests that CAA contributes to neurodegeneration in the form of thinner cerebral cortex, causing some of the memory impairment seen in CAA. This provides a potential mechanism by which CAA may cause cognitive decline in the absence of hemorrhagic stroke, as has been observed in community-based autopsy studies [49]. Some of the thinning may be attributable to WMH, but substantial overlap with regions affected by AD suggests that concomitant AD pathology may also be playing a role. Further research should further examine mechanisms of thinning of the cerebral cortex, and the prognostic significance of thinner cortex including whether it marks patients with a higher risk of future cognitive decline.

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SUPPLEMENTARY MATERIAL

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