Nutritional Intervention to Prevent Alzheimer’s Disease: Potential Benefits of Xanthophyll Carotenoids and Omega-3 Fatty Acids Combined

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Abstract.

Background: A growing body of scientific evidence suggests that enrichment of certain nutritional compounds in the brain may reduce the risk of Alzheimer’s disease (AD).

Objective: To investigate the impact of supplemental xanthophyll carotenoids plus omega-3 fatty acids on disease progression in patients with AD.

Methods: Three trial experiments were performed. In Trials 1 and 2 (performed on patients with AD over an 18-month period), 12 patients (AD status at baseline: 4 mild and 8 moderate) were supplemented with a xanthophyll carotenoid only formulation (Formulation 1; lutein:meso-zeaxanthin:zeaxanthin 10:10:2 mg/day) and 13 patients (AD status at baseline: 2 mild, 10 moderate, and 1 severe) were supplemented with a xanthophyll carotenoid and fish oil combination (Formulation 2; lutein:meso-zeaxanthin:zeaxanthin 10:10:2 mg/day plus 1 g/day of fish oil containing 430 mg docohexaenoic acid [DHA] and 90 mg eicopentaenoic acid [EPA]), respectively. In Trial 3, 15 subjects free of AD (the control group) were supplemented for 6 months with Formulation 1. Blood xanthophyll carotenoid response was measured in all trials by HPLC. Omega-3 fatty acids were profiled by direct infusion mass spectrometry.

Results: Xanthophyll carotenoid concentration increases were significantly greater for Formulation 2 compared to Formulation 1 (\(p < 0.05\)), and progression of AD was less for this group (\(p = 0.003\)), with carers reporting functional benefits in memory, sight, and mood.

Conclusion: This preliminary report suggests positive outcomes for patients with AD who consumed a combination of xanthophyll carotenoids plus fish oil, but further study is required to confirm this important observation.

Keywords: Alzheimer’s disease, dementia, DHA, fatty acids, lutein, meso-zeaxanthin, nutrition, omega-3, prevention, xanthophyll carotenoids

INTRODUCTION

Dementia is a neuropsychiatric syndrome characterized by a combination of cognitive decline, progressive behavioral and psychological symptoms...
Alzheimer’s disease (AD) is the most common type of dementia (accounting for approximately 60–70% of cases), followed by vascular dementia. Dementia represents a major cost burden for healthcare providers. In 2016, the World Alzheimer Report showed that the current annual societal and economic cost of dementia was US$818 billion, and that it is expected to become a trillion-dollar disease this year (2018) [2]. The cost of dementia is the result of direct (e.g., hospital care, community services, nursing homes) [3] and indirect costs (e.g., caregivers contribution) [4].

The exact cause of dementia is unknown, but established risk factors include age, family history of disease, and education [5], and putative risk factors include cigarette smoking, physical inactivity, and social isolation [6]. For AD, the effectiveness of current pharmacological treatments (medications) is limited and varies among individuals. The effect of current medications at best is only palliative, as they cannot halt disease progression. The UK NHS suggests, as preventative, the following: cessation of smoking, limiting alcohol, a healthy well-balanced diet, staying physically and mentally active. Unless new strategies for prevention and management are developed, this syndrome is expected to continue to place growing demands on healthcare and society as a whole.

There is now a growing body of scientific evidence which suggests that enrichment of certain nutritional compounds in the brain may play a role in reducing the risk of AD [7]. Several large prospective observational studies conclude that adherence to a healthy diet, such as the Japanese diet [8] or the Mediterranean diet [9] can reduce risk of AD. Of note, there have been several studies on the Mediterranean diet, characterized by a high intake of vegetables, olive oil, and a moderate intake of fish, dairy products, and wine, which show that adherence to such diets is associated with better cognitive performance [10–14] and a reduced risk of dementia, especially AD [9, 15–18], but no clear evidence of the exact foodstuffs responsible. It is our view that the challenge is therefore to identify the exact nutrients within these diets that are responsible for protecting the brain from degeneration. Achieving this goal would allow for targeted nutritional supplementation for patients with, or at risk of, AD, and thereby reduce the burden of the disease.

To date, the most promising leads have been with omega-3 fatty acids, which include docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA). These fatty acids are found in high concentrations in fish oil and in fatty fish such salmon and herring. There is a substantial concentration of DHA in the brain where it forms a structural component within this neural tissue. High consumption of omega-3 fatty acids is associated with better cognitive performance [19] and a reduced risk of dementia [20–24]. However, interventional studies have shown conflicting results, with some demonstrating improvements in cognition [25, 26] and others demonstrating no beneficial effect [27, 28].

Other food items (nutrients) of interest include the xanthophyll carotenoids lutein (L), zeaxanthin (Z), and meso-zeaxanthin (MZ). L and Z are found in certain fruits and vegetables (e.g., spinach, broccoli, peppers, melon) [29]; whereas, MZ has been identified in fish [30] and is also believed to be generated from L at the retina [31]. In the human, these carotenoids are found in high concentrations at the macula (the center part of the retina, where they are referred to as macular pigment [MP]) and L and Z have been identified in the brain [32, 33]. High xanthophyll carotenoid intake has been found to result in a reduced risk of AD [34–36]. Indeed, interventional studies administering L and Z have shown improvement in different domains of cognition in patients free of AD [37–39]. However, a randomized clinical trial performed by our group in patients with AD who were supplemented with xanthophyll carotenoids found no benefit in measures of cognitive function performed in that trial [40].

In summary, there is general agreement that there are nutritional compounds in the brain which play a role in preventing AD [7], but attempts so far to identify these exact nutrients, and use them to prevent AD have been unsuccessful. We propose here that we have identified “a synergistic nutritional strategy” via a unique combination of xanthophyll carotenoids and omega-3 fatty acids to potentially slow the progression rate of AD and aid in the management of this disease. The preliminary trial data are presented below.

MATERIALS AND METHODS

Study sample

This research was conducted at the Nutrition Research Centre Ireland (NRCI; http://www.nrci.ie)
in collaboration with University Hospital Waterford (UHW), South East, Republic of Ireland. Patients with AD were recruited (for Trials 1 and 2, see below) from UHW via the Age-Related Care Unit department. Subjects for the control group (Trial 3, see below) were recruited via newspaper and radio advertisements, and by word of mouth in the local community. This study was conducted in accordance with full sensitivity to the ethical requirements of the patients recruited. The study objectives and methodology complied fully with the widely-recognized international text and codes of practice, such as the Declaration of Helsinki and Good Research Practice. A protocol for obtaining informed consent was developed specifically for this trial by the Principal Investigator (JMN) of the NRCI and Consultant Geriatrician (RM) of the UHW to ensure that the protocol was safe and appropriate, and in keeping with the ethical code germane to obtaining consent from vulnerable subjects (which includes patients with AD). The protocol for obtaining informed consent was assessed and approved by the local ethics committee prior to the study commencing [41]. The first ethical approval for the Carotenoids And Retinal Dementia Study (CARDS) was granted by local research ethics committee, Health Services Executive, South Eastern Area on 23 August 2012 and duration extension requests/amendments were granted on 14 January 2013, 30 October 2013, 6 May 2014, and the 27 May 2015.

Study trials

In brief, three trial experiments were performed. Two trials were conducted in patients with AD and one trial in age-matched control subjects (free of AD). In Trial 1, 12 patients with AD (4 mild and 8 moderate) were supplemented with a xanthophyll carotenoid only formulation (Formulation 1) over an 18-month period. For note, these 12 patients were followed from the original CARDS sample of 16 [40], as they were available and agreed to continue for the longitudinal 18-month intervention. The daily formulation in Trial 2 was 10 mg L, 10 mg MZ, and 2 mg of Z plus 1 gram of fish oil (430 mg DHA and 90 mg EPA). In Trial 3, 15 subjects free of AD (the control group) were supplemented for 6 months only using the same intervention as described above for Trial 1 (i.e., Formulation 1), and data from this trial has already been published [40]. Both study formulations were provided as a softgel capsules.

Xanthophyll carotenoids

Assessment performed at baseline and after 6 months of supplementation. Non-fasting blood samples were collected in 9 ml vacuette tubes containing a ‘Z Serum Sep Clot Activator’, and collection tubes were inverted circa 6 times. The blood samples were allowed to clot at room temperature for approximately 30 min and then centrifuged at 2700 rpm for 10 min in a Gruppe GC 12 centrifuge (Desaga Sarstedt) to separate the serum from the whole blood. The resulting serum samples were stored at circa –80°C until the time of batch analysis using high performance liquid chromatography (HPLC). First, the serum samples were analyzed for L and total Z (co-eluted Z and MZ) using a reversed-phase HPLC method (Assay 1) [41]. The mixed Z fraction was automatically collected from Assay 1 using an Agilent 1260 fraction collector. The eluent was dried under a solvent concentrator (MiVac, GeneVac, Mason Technologies, Dublin, Ireland) and analyzed on Assay 2 for quantification of Z and MZ (Assay 2) [42].

Omega-3 fatty acid

Assessment performed at baseline and after 6 months of supplementation. For Trial 2, non-fasting venous blood samples were collected into 6 ml lithium heparin tubes for lipid analysis. Collection tubes were inverted 8–10 times to ensure thorough mixing and centrifuged at 4°C at 3000 rpm for 20 min to separate red blood cells and plasma from whole blood. The time of blood collection and time of separation did not exceed 2 h. Plasma was aliquoted to microtubes and stored at –80°C until further analysis. Lipids were profiled at the Medical Research Council Human
Nutrition Research, Cambridge, UK by direct infusion mass spectrometry as described previously [43].

**Macular pigment: Central nervous system xanthophyll carotenoid status**

MP was measured using the Heidelberg Spectralis® HRA+OCT Multicolor (Heidelberg Engineering GmbH, Heidelberg, Germany). This new technology utilizes confocal scanning laser ophthalmoscopy (cSLO) imaging with diode lasers and uses dual-wavelength autofluorescence (AF) for measuring MP. Dual-wavelength AF in this device uses two excitation wavelengths, one that is well-absorbed by MP (488 nm, blue), and one that is not well absorbed by the pigment (518 nm, green). Of note, the AF method utilized in this study has previously been tested and validated [44]. The Heidelberg Spectralis® provides an image of MP across its spatial profile, but here we report just central MP (at 0.23° eccentricity) and MP volume (calculated as MP average times the area under the curve out to 7° eccentricity).

**Dietary intake of xanthophyll carotenoids**

A subject’s weekly intake of carotenoid-rich foods (eggs, broccoli, corn, dark leafy vegetables) was inputted into the “L/Z screener” to provide a carotenoid-based diet score. The L and Z values used in the screener were those reported by Perry et al. [43]. This method of assessing and controlling for dietary intake of xanthophyll carotenoids has been used with success elsewhere [12]. Values are weighted for frequency of intake of the food and for bioavailability of L and Z within these foods. A ranking score reflecting the relative intakes (representing arbitrary units) was generated and used in analysis. For the AD subjects, dietary habits were confirmed by a family member or carer. The range of scores on the L/Z screener is 0 to 75. After adding foods with known concentrations of the L and Z into the screener, the following estimates can be made: a low dietary xanthophyll carotenoid intake score ranges from 0–15 (≤2 mg/day); a medium dietary xanthophyll carotenoid score ranges from 16–30 (3–13 mg/day); and a high dietary xanthophyll carotenoid intake score ranges from 31–75 (>13 mg/day).

**Clinical/functional assessment**

A clinical and functional assessment of each patient was performed at baseline and after 18-months of supplementation. Clinical assessment was performed by a qualified AD nurse (MB) under the supervision of a medical consultant (RM). Also, the patient carer (i.e., the primary person responsible for caring for the patient) was interviewed after 18 months of the patient taking the supplement. AD status at baseline was confirmed by the medical consultant. The Mini-Mental State Examination (MMSE) was used to guide the diagnosis of AD during patient recruitment (baseline). With MMSE, AD status is defined as follows: 21–25 = Mild AD; 11–20 = Moderate AD; 0–10 = Severe AD. Of note, MMSE data was not obtained at 18 months as this is not part of routine medical assessment at UHW. For our report, AD status was coded as follows: Mild = 0; Moderate = 1; Advanced = 2. AD status was provided by the research nurse (MB) under the guidance and supervision of the medical consultant and guided by all information available to the research team (i.e., MMSE; clinical story; mood; function). Health status and assessment of each patient was performed after 18 months, where possible. To achieve this, the research nurse interviewed the carer of the patient to assess health status and patient function. Notes were recorded from each assessment. Importantly, the research nurse confirmed that the carer was the same person at baseline and 18-month follow up.

**Statistical analysis**

The statistical package IBM SPSS version 22 was used, and the 5% significance level applied, for all analyses. SigmaPlot version 8.0 was used to generate line graphics for presentation. Results were expressed as means ± standard deviations in tables and means ± standard errors for graphics. Between-group differences were analyzed using Independent Samples *t*-tests, One-way analysis of variance (ANOVA) or chi-square tests as appropriate. Repeated measures analyses of variance (rANOVA) was used to assess Time and/or Time-Group interactions between the treatment intervention groups for serum concentrations of L, Z and MZ. Time effects examine whether or not a response variable is different at the various time points of interest. Time-Group effects examine whether or not the time effect differs between the treatment intervention groups, therefore testing the impact of the intervention.
Table 1

Demographic, AD status and biochemical health variables for patients recruited into the Trials at baseline

<table>
<thead>
<tr>
<th>Demographic and health variables</th>
<th>Trial 1 (n = 12); AD</th>
<th>Trial 2 (n = 13); AD</th>
<th>Trial 3 (n = 15); Controls</th>
<th>Comparison AD groups</th>
<th>Comparison all groups</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>78.5 ± 8.754</td>
<td>78.77 ± 7.65</td>
<td>77 ± 7.4</td>
<td>p = 0.935</td>
<td>p = 0.444</td>
</tr>
<tr>
<td>Sex (%females)</td>
<td>6 (50%)</td>
<td>8 (62%)</td>
<td>6 (40%)</td>
<td>p = 0.561</td>
<td>p = 0.524</td>
</tr>
<tr>
<td>Dietary L/Z</td>
<td>14.9 ± 8.78</td>
<td>13.69 ± 10.1</td>
<td>21.92 ± 14.72</td>
<td>p = 0.751</td>
<td>p = 0.156</td>
</tr>
<tr>
<td>Serum L (µmol/L)</td>
<td>0.261 ± 0.142</td>
<td>0.154 ± 0.084</td>
<td>0.275 ± 0.175</td>
<td>p = 0.031</td>
<td>p = 0.068</td>
</tr>
<tr>
<td>Serum Z (µmol/L)</td>
<td>0.048 ± 0.035</td>
<td>0.062 ± 0.031</td>
<td>0.066 ± 0.035</td>
<td>p = 0.297</td>
<td>p = 0.409</td>
</tr>
<tr>
<td>BMI</td>
<td>24.6 ± 3</td>
<td>24.4 ± 3</td>
<td>27.17 ± 3</td>
<td>p = 0.863</td>
<td>p = 0.078</td>
</tr>
<tr>
<td>MMSE category</td>
<td>19 ± 2.89</td>
<td>16 ± 2.873</td>
<td>29 ± 1.311</td>
<td>p = 0.140</td>
<td>p = 0.000</td>
</tr>
<tr>
<td>Mild AD</td>
<td>4 (33%)</td>
<td>2 (15%)</td>
<td>na</td>
<td>p = 0.105</td>
<td>–</td>
</tr>
<tr>
<td>Severe AD</td>
<td>0 (0%)</td>
<td>1 (8%)</td>
<td>na</td>
<td>p = 0.105</td>
<td>–</td>
</tr>
<tr>
<td>MP 0.23</td>
<td>0.41 ± 0.24</td>
<td>–</td>
<td>0.55 ± 0.17</td>
<td>p = 0.097</td>
<td>–</td>
</tr>
<tr>
<td>MP Volume</td>
<td>4114 ± 2308</td>
<td>–</td>
<td>6092 ± 2421</td>
<td>p = 0.019</td>
<td>–</td>
</tr>
<tr>
<td>DHA</td>
<td>–</td>
<td>0.160 ± 0.118</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>EPA</td>
<td>–</td>
<td>0.153 ± 0.144</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
</tbody>
</table>

AD, Alzheimer’s disease; Trial 1, patients with AD supplemented with Formulation 1; Formulation 1, 10 mg lutein, 10 mg meso-zeaxanthin, and 2 mg of zeaxanthin (carotenoid only); Trial 2, patients with AD supplemented with Formulation 2; Formulation 2, 10 mg lutein, 10 mg meso-zeaxanthin, and 2 mg of zeaxanthin plus 1 gram of fish oil (430 mg docohexaenoic acid [DHA] and 90 mg eicopentaenoic acid [EPA]) (carotenoid plus fish oil); Trial 3, aged-matched control subjects supplemented with Formulation 1; Dietary L/Z, a dietary score for lutein and zeaxanthin intake (see methods); Serum L and Z, serum concentration of lutein and zeaxanthin measured in µmol/L; BMI, body mass index expressed in units of kg/m²; MMSE, Mini-Mental State Examination; MP 0.23, macular pigment at center (0.23 degrees of retinal eccentricity); MP volume, macular pigment average times the area under the curve; DHA, LPC 22:6 phospholipid; EPA, LPC 20:5 phospholipid.

RESULTS

Baseline

Table 1 below presents baseline data for AD patients and control subjects enrolled into the three Trials. As seen in this table, the patients with AD in Trial 1 and Trial 2 were comparable in age, sex, BMI, dietary L/Z intake, serum Z concentrations and AD category. Patients in Trial 2 had significantly lower blood L concentrations than patients in Trial 1 (p = 0.031). Other observations of note at baseline were the positive and statistically significant relationships between dietary L and Z intake and serum concentrations of L and Z (r = 0.601, p = 0.000 and r = 0.330, p = 0.043, respectively).

Biochemical response: Xanthophyll carotenoids and omega-3 fatty acids

The increase observed in L and MZ concentrations was significantly greater in the AD patients that were supplemented with the combination of xanthophyll carotenoids plus omega-3 fatty acids (Formulation 2),...
When compared to the AD patients that were supplemented with the xanthophyll carotenoids only formulation (Formulation 1) and when compared to the control group that were supplemented with the xanthophyll carotenoid only formulation (Formulation 1) (serum L: $p=0.002$, Fig. 1A; serum MZ: $p=0.006$, Fig. 1B).

Also, for Trial 2, DHA and EPA concentrations increased in the blood of the AD patients (of note, DHA and EPA measurements were only performed in this trial because the other trials did not have DHA or EPA in the supplement formulation). The increase observed in DHA, the main component of the fish oil used in Formulation 2, was statistically significant ($p=0.041$, see Fig. 2).

**Clinical/functional response in patients with Alzheimer’s disease**

Figures 3A and 3B present the AD status for Trial 1 at baseline and 18-months, respectively. Figures 4A and 4B present the AD status for Trial 2 at baseline and 18-months, respectively. Of note, data at 18-months represents medical observations and patient health status (see Table 2). As seen from the pie charts (Fig. 3A, B) and Table 2, progression of AD was evident in this group (data from Trial 1, xanthophyll carotenoid only intervention) over the 18-month period, with 42% of patient’s health status dropping to the point that they could not continue in the trial. Reasons for dropout included: patient moved into a nursing home due to AD progressing and unable to take the supplement; patient became too unwell to continue; cognitive decline too severe; no longer able to follow instructions.

However, as seen in Fig. 4A and 4B (data from Trial 2, xanthophyll carotenoid and fish oil intervention) and Table 2, progression of AD appeared much less, with carers reporting functional benefits in memory, sight, and mood. Pearson Chi-Square analysis demonstrated statistically significant between-group difference for the changes seen in AD status from baseline to 18-months ($p=0.003$).

**DISCUSSION**

From this study, we propose that a combination of xanthophyll carotenoids and omega-3 fatty acids
Table 2
Biochemical/clinical/functional response in patients with Alzheimer’s disease

<table>
<thead>
<tr>
<th>Study</th>
<th>Subject ID</th>
<th>Age (y)</th>
<th>Sex</th>
<th>Baseline MMSE score</th>
<th>Baseline AD Status</th>
<th>Baseline Serum L (μmol/L)</th>
<th>6-months Serum L (μmol/L)</th>
<th>Baseline Serum Z (μmol/L)</th>
<th>6-months Serum Z (μmol/L)</th>
<th>6-months Serum MZ (μmol/L)</th>
<th>Observations at 18 months</th>
</tr>
</thead>
<tbody>
<tr>
<td>ADCD2</td>
<td>78 M</td>
<td>16</td>
<td>Moderate</td>
<td>0.072</td>
<td>0.666</td>
<td>0.006</td>
<td>0.062</td>
<td>0.08</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADCD3</td>
<td>81 M</td>
<td>17</td>
<td>Moderate</td>
<td>0.432</td>
<td>0.933</td>
<td>0.046</td>
<td>0.088</td>
<td>0.08</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADCD4</td>
<td>84 M</td>
<td>18</td>
<td>Moderate</td>
<td>0.302</td>
<td>0.973</td>
<td>0.057</td>
<td>0.099</td>
<td>0.052</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADCD5</td>
<td>82 F</td>
<td>23</td>
<td>Mild</td>
<td>0.466</td>
<td>0.676</td>
<td>0.133</td>
<td>0.099</td>
<td>0.034</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADCD8</td>
<td>90 F</td>
<td>15</td>
<td>Moderate</td>
<td>0.149</td>
<td>0.564</td>
<td>0.032</td>
<td>0.056</td>
<td>0.03</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADCD9</td>
<td>76 M</td>
<td>22</td>
<td>Mild</td>
<td>0.325</td>
<td>1.631</td>
<td>0.076</td>
<td>0.189</td>
<td>0.03</td>
<td>Patient too sick to continue in study</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADCD11</td>
<td>68 F</td>
<td>19</td>
<td>Moderate</td>
<td>0.397</td>
<td>0.941</td>
<td>0.057</td>
<td>0.076</td>
<td>0.04</td>
<td>No information available</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADN24</td>
<td>68 M</td>
<td>23</td>
<td>Mild</td>
<td>0.149</td>
<td>0.285</td>
<td>0.054</td>
<td>0.041</td>
<td>0.01</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADN25</td>
<td>64 F</td>
<td>15</td>
<td>Moderate</td>
<td>0.202</td>
<td>0.771</td>
<td>0.035</td>
<td>0.078</td>
<td>0.02</td>
<td>Patient unable to continue in study</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADN26</td>
<td>74 F</td>
<td>21</td>
<td>Mild</td>
<td>0.364</td>
<td>1.972</td>
<td>0.059</td>
<td>0.248</td>
<td>0.294</td>
<td>Patient unable to follow instruction</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADN29</td>
<td>87 F</td>
<td>19</td>
<td>Moderate</td>
<td>0.238</td>
<td>2.415</td>
<td>0.02</td>
<td>0.245</td>
<td>0.234</td>
<td>Patient too sick to continue in study</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ADN31</td>
<td>90 M</td>
<td>20</td>
<td>Moderate</td>
<td>0.034</td>
<td>0.364</td>
<td>0.003</td>
<td>0.024</td>
<td>0.018</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A4</td>
<td>79 F</td>
<td>12</td>
<td>Moderate</td>
<td>0.051</td>
<td>0.548</td>
<td>0.027</td>
<td>0.057</td>
<td>0.02</td>
<td>Good improvement in memory</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A5</td>
<td>78 M</td>
<td>22</td>
<td>Mild</td>
<td>0.204</td>
<td>1.733</td>
<td>0.05</td>
<td>0.145</td>
<td>0.1</td>
<td>Good improvement in memory and sight</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A6</td>
<td>89 F</td>
<td>14</td>
<td>Moderate</td>
<td>0.273</td>
<td>3.599</td>
<td>0.059</td>
<td>0.312</td>
<td>0.28</td>
<td>Subject was improved and manageable</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A7</td>
<td>81 F</td>
<td>13</td>
<td>Moderate</td>
<td>0.095</td>
<td>2.515</td>
<td>0.041</td>
<td>0.226</td>
<td>0.15</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A8</td>
<td>87 M</td>
<td>17</td>
<td>Moderate</td>
<td>0.153</td>
<td>1.021</td>
<td>0.058</td>
<td>0.094</td>
<td>0.091</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A9</td>
<td>88 F</td>
<td>21</td>
<td>Mild</td>
<td>0.122</td>
<td>0.536</td>
<td>0.069</td>
<td>0.082</td>
<td>0.03</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A10</td>
<td>71 F</td>
<td>18</td>
<td>Moderate</td>
<td>0.107</td>
<td>2.722</td>
<td>0.026</td>
<td>0.244</td>
<td>0.186</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A12</td>
<td>79 F</td>
<td>17</td>
<td>Moderate</td>
<td>0.244</td>
<td>2.292</td>
<td>0.075</td>
<td>0.153</td>
<td>0.15</td>
<td>Good improvement in function</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A13</td>
<td>77 M</td>
<td>10</td>
<td>Severe</td>
<td>0.064</td>
<td>1.971</td>
<td>0.044</td>
<td>0.131</td>
<td>0.178</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A14</td>
<td>69 M</td>
<td>17</td>
<td>Moderate</td>
<td>0.074</td>
<td>0.342</td>
<td>0.048</td>
<td>0.039</td>
<td>0.1</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A16</td>
<td>87 M</td>
<td>18</td>
<td>Moderate</td>
<td>0.123</td>
<td>0.974</td>
<td>0.043</td>
<td>0.058</td>
<td>0.085</td>
<td>Good improvement in memory</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A17</td>
<td>74 F</td>
<td>18</td>
<td>Moderate</td>
<td>0.315</td>
<td>1.879</td>
<td>0.137</td>
<td>0.065</td>
<td>0.252</td>
<td>No difference reported</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C3A19</td>
<td>65 M</td>
<td>18</td>
<td>Moderate</td>
<td>0.182</td>
<td>1.97</td>
<td>0.105</td>
<td>0.148</td>
<td>0.226</td>
<td>Good improvement in function</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Trial 1, patients with AD supplemented with Formulation 1; Formulation 1, 10 mg lutein, 10 mg meso-zeaxanthin, and 2 mg of zeaxanthin (carotenoid only); Trial 2, patients with AD supplemented with Formulation 2; Formulation 2, 10 mg lutein, 10 mg meso-zeaxanthin, and 2 mg of zeaxanthin plus 1 gram of fish oil (430 mg docohexaenoic acid [DHA] and 90 mg eicopentaenoic acid [EPA]) (carotenoid plus fish oil); methods; Serum L and Z, serum concentration of lutein and zeaxanthin measured in μmol/L; MMSE, Mini-Mental State Examination; Observations at 18-months, data at 18-months represents clinical and carer observations of patient health status and changes in health status.
work synergistically to maintain function and quality of life in patients with AD. The findings from this work were unexpected, as the trials were not conducted to assess the impact of nutritional supplements on AD progression; rather, these trials were designed to measure biochemical response to xanthophyll carotenoid and omega-3 fatty acid supplementation [40, 41]. However, assessment of the patients (compiled from the research nurse reports, which included reports from the patient carers) presented very striking differences with regard to the health of the patients during the trial. Of importance, these differences were directly linked to the intervention group, where the positive outcomes were identified in patients consuming the combination of xanthophyll carotenoids and fish oil. Of note, this is the first scientific investigation reporting on the impact of a xanthophyll carotenoid and omega-3 intervention in patients with AD. Here, we examine the scientific rationale in support of our findings, and we do so below by discussing the role of nutrition in the brain under the following headings: AD pathogenesis; presence of xanthophyll carotenoids and omega-3 fatty acids in the brain; mechanisms of action; synergistic effect of nutrition.

There is currently no known cure for AD. Current management of AD involves drugs which aim to slow disease progression and improve cognition and behavior. Of note, only two drugs are in widespread clinical use. These include cholinesterase inhibitors and NMDA-receptor antagonists (memantine) [45, 46]. However, these drugs are associated with numerous adverse effects including gastrointestinal disturbance, rhinorrhea, confusion and sedation. Cholinesterase inhibitor therapy is associated with a doubling of risk of hospitalization for bradycardia [47]. Also, cholinesterase inhibitors and memantine exert only a modest and temporary beneficial effect on cognitive function, mood, and behavior [45]. The existing pharmacological approach for the management of established AD remains unsatisfactory, thereby prompting increasing attention being directed toward preventative strategies for this condition [48]. Well-recognized modifiable risk factors for AD include smoking, diabetes, ischemic heart disease, hypertension, obesity, and dyslipidemia [49]. Also, there is a growing body of evidence that good nutrition is important for optimal cognition [50], maintenance of cognition [51], and is also associated with reduced risk of AD [7, 52].

Oxidative stress is believed to play a major role in the development of AD, and represents one of the earliest events of this condition [53]. The brain is particularly susceptible to oxidative damage because of the high oxygen demand and high proportion of polyunsaturated fatty acids in this neural tissue. In particular, the oxidation-reduction reactions necessary for the generation of Adenosine triphosphate (ATP) produce reactive oxygen species (ROS) as electrons are transferred from one molecule to another [54]. The generation of ROS (or free radicals) and associated cellular damage contributes to the pathology seen in susceptible neurons in AD. It is for this reason that antioxidants offer real potential for preventing AD.

As noted previously, the xanthophyll carotenoids L, Z, and MZ are found in their highest concentration at the macula (retina), which is part of the central nervous system. To date, L and Z have been identified in the hippocampus, cerebellum and...
frontal, occipital and temporal cortices [32, 33], and in amounts that correlate positively and significantly with their respective retinal concentrations [55]. Therefore, the retinal concentrations of these xanthophyll carotenoids (known as MP) can be used as a biomarker for these carotenoids in the brain. Interestingly, MP is positively related to cognitive performance in cognitively healthy individuals [56] and cognitively impaired individuals [57]. Indeed, data from our group has shown that xanthophyll carotenoid intervention (with a formulation containing L, MZ and Z; 10:10:2) in cognitively healthy individuals results in improvements in memory and function [39]. In another paper, we have shown that patients with AD have significantly lower serum xanthophyll carotenoid concentrations and MP levels compared to controls [41]. However, data on the impact of xanthophyll carotenoid intervention for patients with AD is limited. The emerging view that these xanthophyll carotenoids are neuro-protective is premised upon their chemical composition (i.e., amount of conjugated double-bonds and the presence of polar hydroxyl groups) and their localization within the lipid bi-layer of the cell membrane, thus bestowing antioxidant and anti-inflammatory activity at loci where they are needed [58, 59].

It has also been suggested that these xanthophyll carotenoids can enhance the structural integrity of membranes [60] and positively impact neural efficiency [61, 62].

Omega-3 fatty acids (especially DHA) are present in high concentrations in the human brain, but most of the studies to date have been conducted on the impact of fatty acids on brain development in infants [63]. DHA participates in a number of important neuronal processes including enhancement of membrane integrity and fluidity, neurogenesis, neuroplasticity, neuron differentiation and survival. Of interest, over the past 150 years, the Western diet has changed such that the ratio of omega-3 to omega-6 fatty acids has altered from 1:1 to 1:20, signifying that the Western diet is now deficient in omega-3 fatty acids [64]. This recent nutrient deficiency of DHA has most likely contributed to the increased prevalence and incidence AD, given the above outlined functions of DHA for the brain.

The results from the current study suggest that the improvements we identified in patients with AD were uniquely achieved only when a combination of the xanthophyll carotenoids and omega-3 fatty acids were provided to the patients. In this work, we observed noticeable differences between patients supplemented with xanthophyll carotenoids plus omega-3 fatty acids compared to patients supplemented with the xanthophyll carotenoids only, in terms of their health and function. In Trial 2, the “clinical story” for the patients was more positive than patients in Trial 1. In fact, the carers for patients in Trial 2 reported functional benefits in memory, sight and mood. Following completion of the trial, families and carers of the patients in Trial 2 have contacted our research center requesting continued access to the supplement, as they are adamant that the intervention with xanthophyll carotenoids and fish oil has had a very positive impact for the patient. We acknowledge that these reports are anecdotal. Also, our data demonstrates a superior biochemical (blood) response to the xanthophyll carotenoids when supplemented in combination with the omega-3 fatty acid (fish oil) compared to providing the exact same formulation of xanthophyll carotenoids (MZ:L:Z: 10:10:2) in the absence of fish oil. The improved delivery of these important brain nutrients to the target (neural) tissue is likely to have contributed to the positive outcomes linked to this formulation (i.e. via enhanced absorption and distribution of the carotenoids). Of note, this finding is consistent with a previous report by Johnson et al who showed that supplementation with DHA and lutein improve serum and MP levels in healthy elderly female subjects [65]. This finding may be explained by the hydrophobic nature of the xanthophyll carotenoids, as it is known that consuming carotenoid-rich foods in the presence of oils or cholesterol will increase carotenoid uptake and bio-accessibility [66].

It is important to point out that while the results of this study are very promising and offer hope for the management of AD, there are several limitations that need to be acknowledged. Firstly, the number of subjects in each Trial for this report was small and therefore our findings must only be viewed as preliminary. The findings presented here will need to be confirmed by a larger powered sample, and with appropriately definite outcome measures and study design. In the current trials, MMSE was only performed at baseline and we did not have the opportunity to capture specific outcome measures of functional or cognitive abilities. We acknowledge that such investigation is merited and will greatly aid our understanding of the positive outcomes reported here. Also, MP was not measured in Trial 2 patients, which represents another limitation of the current study.

In conclusion, this preliminary report, which is supported by a biologically plausible scientific ratio-
nale, suggests positive outcomes for patients with AD who consumed a combination of xanthophyll carotenoids and fish oil over an 18-month period. These patients demonstrated maintenance of cognitive and visual function that was not achieved when a xanthophyll carotenoid only intervention was used. We believe that the results of this study should guide a focused nutritional interventional strategy for patients with AD, but we acknowledge that these positive findings will need to be tested and confirmed with further study.

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CONFLICT OF INTEREST

Authors’ disclosures available online (https://www.j-alz.com/manuscript-disclosures/18-0160r2).

REFERENCES


