

EGFR, HLA-G, CD70, c-MET, and NY-ESO1 as potential biomarkers in high grade epithelial ovarian carcinoma

Duc Vo^a, Yan Liu^b, Anil K. Sood^c, Katy Rezvani^d, Amir A. Jazaeri^c and Jinsong Liu^{e,*}

^aMD Anderson Cancer Center, Department of Anatomical Pathology, Houston, TX, USA

ORCID: <https://orcid.org/0000-0002-2564-529X>

^bMD Anderson Cancer Center, Department of Anatomical Pathology, Houston, TX, USA

ORCID: <https://orcid.org/0000-0003-4242-1762>

^cMD Anderson Cancer Center, Department of Gynecologic Oncology & Reproductive Medicine, Houston, TX, USA

^dMD Anderson Cancer Center, Department of Stem Cell Transplantation, Houston, TX, USA

^eMD Anderson Cancer Center, Department of Anatomical Pathology, Houston, TX, USA

ORCID: <https://orcid.org/0000-0002-3239-6658>

Received 15 May 2023

Accepted 19 November 2023

Abstract. High grade epithelial ovarian carcinoma is an aggressive tumor. Treatment includes platinum therapy, however it recurs in most patients due to therapy resistance. In this project, we study the immunohistochemical (IHC) expression of five potential biomarkers/prognostic markers in high grade epithelial ovarian carcinoma: EGFR, HLA-G, CD70, c-MET, and NY-ESO1. A cohort of 274 patients is used. We compare the IHC expression with age, stage, ascites status, family history of cancer, disease free survival (DFS) and overall survival (OS). EGFR expression is significantly correlated with family history and worse OS. HLA-G is associated with worse OS. To confirm the results of EGFR and HLA-G, a second separated cohort of 248 patients is used. Positive EGFR expression again shows worse OS, while HLA-G expression has worse prognostic trend. CD70 has a worse OS trend. C-MET and NY-ESO1 do not have any clinical correlations. EGFR can potentially serve as target in future clinical immune therapy trials.

Keywords: EGFR, HLA-G, CD70, c-MET, NY-ESO1, survival

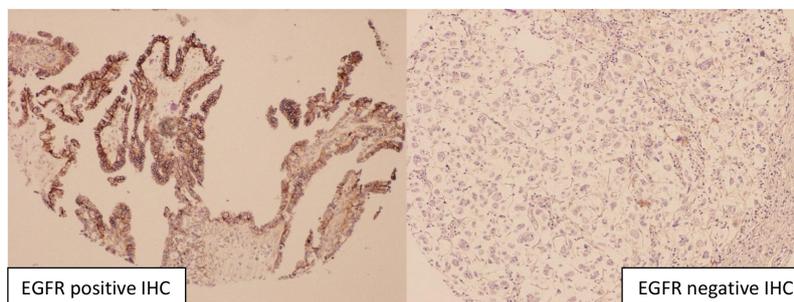
1. Background

Among all gynecological cancer types, ovarian carcinoma has the worst prognosis and the highest mortality rate [1]. Histologically, it consists of several distinct subtypes, including high grade serous carcinoma, endometrioid carcinoma, clear cell carcinoma, and mucinous carcinoma. Rare high grade subtypes include

malignant mixed Mullerian tumor (MMMT), undifferentiated carcinoma, and mixed type carcinoma [2]. Due to the aggressiveness of the disease and the absence of early symptoms, the five-year survival rate is less than 50% [3]. Current first-line treatment includes cytoreductive surgery and platinum-based chemotherapy. However, patients frequently experience recurrence due to platinum resistance [4,5].

Therefore, immunotherapy and targeted therapies can potentially serve as new treatments for ovarian carcinoma. Immunotherapy enhances the attack of immune system on neoplastic cells through different approaches: immunostimulatory cytokines, tumor antigen

*Corresponding author: Jinsong Liu, Department of Pathology, Unit G1.3728, The University of Texas MD Anderson Cancer Center, 1515 Holcombe Blvd, Houston, Texas 77030, USA. Tel.: +1 713 745 1102; E-mail: jlui@mdanderson.org.



EGFR IHC results			
	EGFR positive	EGFR negative	
Age <51	10	48	p = 0.58
Age ≥51	31	185	
High stage	40	214	p = 0.43
Low stage	1	12	
Ascites	19	165	p = 0.25
No ascites	3	12	
Family history of cancer	25	105	p = 0.01
No family history of cancer	10	111	

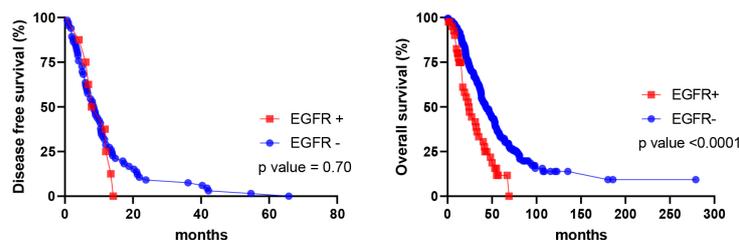


Fig. 1. EGFR results in cohort 1.

vaccines, antibodies targeting immunosuppressive ligands expressed by tumor cells and immune checkpoint inhibition. The immune system utilizes the immune checkpoints (e.g., CTLA-4, PD-1, HHLA2, B7-H4, and TIM-3) to recognize self-cells versus foreign-cells. The expression of immune checkpoints within the tumor cells helps evade the attack by T cells. The cytotoxicity of T cells toward tumor cells is restored by blocking the immune checkpoints [6,7].

Membrane proteins make up approximately 30% of total human proteins and they play roles in various physiological and pathological functions [8]. Tumor derived membrane proteins may serve as potential biomarkers in early diagnosis. They can also be used to assess disease progression and prediction for treatment response [8]. EGFR, HLA-G, CD70, c-MET and NY-ESO1 have been found to play certain roles in tumor genesis and immune evasion in different tumor types. Reports have shown membranous expression of these markers in ovarian carcinoma. However, the clinical significance is not fully understood.

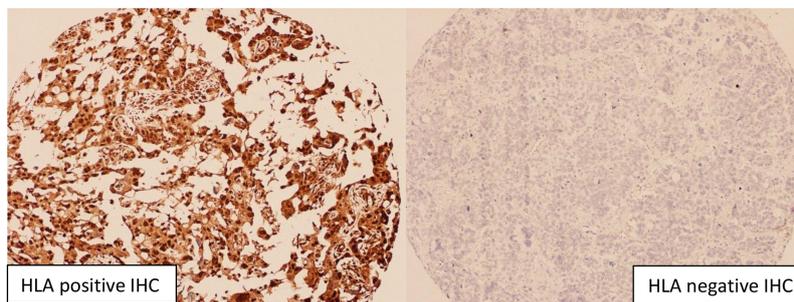
2. Objective

In our study, we conduct IHC studies of five previously mentioned markers with associated immunotherapy on two large cohorts of ovarian carcinoma. Subsequently, we correlate their expression with the survival rates.

3. Materials and methods

3.1. Study population

In this study, two cohorts of patients were utilized. The first cohort comprised a total of 418 female patients with various ovarian tumor types, with 274 of them diagnosed with high grade ovarian carcinoma. The second cohort included 453 female patients with different ovarian tumors, of which 248 were diagnosed with high grade ovarian carcinoma. Patient data was



HLA-G IHC results			
	HLA-G positive	HLA-G negative	
Age <51	4	54	p = 0.39
Age ≥51	9	207	
High stage	13	241	p = 0.40
Low stage	0	13	
Ascites	11	173	p = 0.91
No ascites	1	14	
Family history of cancer	9	121	p = 0.20
No family history of cancer	4	117	

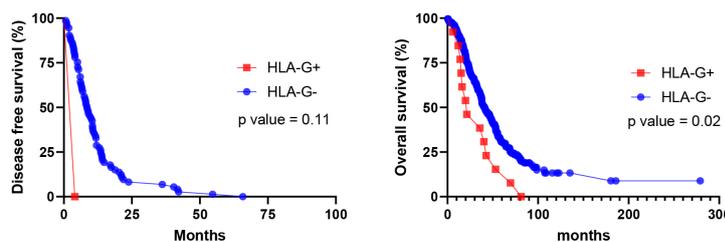


Fig. 2. HLA results in cohort 1.

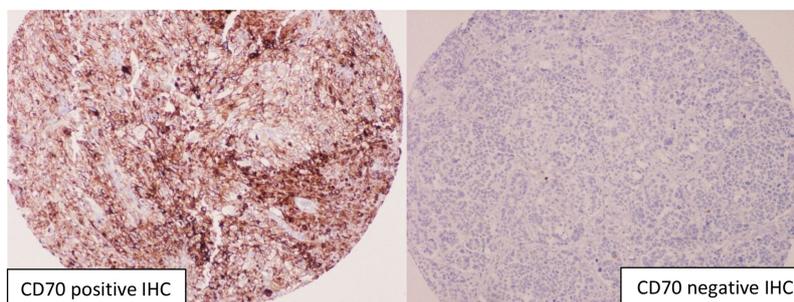
collected from electronic medical records and laboratory information systems spanning from 1987 to 2006. We extracted information such as ovarian cancer diagnoses, patient ages, AJCC stages, ascites presence, family history, DFS intervals, OS times, and survival statuses.

3.2. Tissue microarray (TMA) construction and Immunohistochemistry

Formalin-fixed, paraffin-embedded (FFPE) tissue blocks from the two cohorts were retrieved from the MD Anderson Cancer Center storage room. The tissue blocks had been stored under ambient conditions at approximately 24°C. H&E stained sections from each patient were reviewed by a pathologist to identify representative tumor areas. TMAs were constructed by taking one to two core samples from the identified areas on the FFPE blocks and assembled on the TMA-paraffin blocks. The extraction of the tissue from FFPE

blocks and the mapping of the TMA blocks were performed using a precision instrument (Beecher Instruments, Silver Spring, MD, USA). For each case, one to two 1 mm wide and 5 mm deep samples were collected and mapped in the TMA blocks. The final TMAs consisted of seven blocks, each containing from 100 to 148 punches. Four-micrometer thick sections from each TMA block were stained with EGFR, CD70, c-MET, HLA-G, and NY-ESO1 IHC.

EGFR antibody clone 31G7 from Abnova with a 1:20 dilution, CD70 antibody clone E3Q1A from Cell Signaling with a 1:50 dilution, c-MET antibody clone SP44 from Ventana with a 1:1 dilution, HLA-G antibody clone 3H2678 from US Biological Life Sciences with a 1:100 dilution, and NY-ESO1 antibody clone E978 from Santa Cruz Biotechnology with a 1:100 dilution were used to perform immunohistochemical stains on the TMAs. HLA-G was incubated in Citrate buffer, while EGFR, NY-ESO1, c-Met, and CD70 were incubated in Tris-EDTA buffer. The staining process was



CD70 IHC results			
	CD70 positive	CD70 negative	
Age <51	4	54	p = 0.21
Age ≥51	7	209	
High stage	9	245	p = 0.44
Low stage	1	12	
Ascites	7	177	p = 0.44
No ascites	0	15	
Family history of cancer	3	127	p = 0.16
No family history of cancer	7	114	

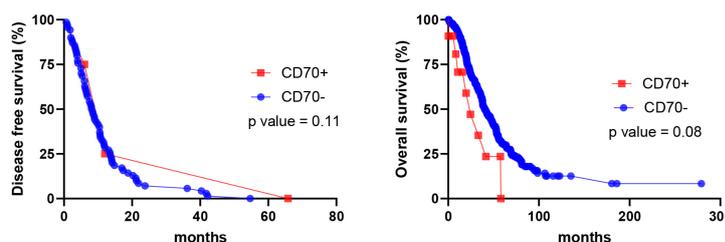


Fig. 3. CD70 results in cohort 1.

performed at MD Anderson Cancer Center immunohistochemical laboratory.

The results were stratified as either negative (no staining or non-specific staining) or positive (> 1% membranous staining for EGFR, CD70, and C-MET; > 1% cytoplasmic staining for HLA-G and NY-ESO1).

Descriptive statistics, Kaplan Meier curves, and survival analysis were performed using GraphPad Prism v.8.4.3 software (La Jolla, CA).

3.3. Institutional review board statement

The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Institutional Review Board of MD Anderson Cancer Center (protocol code: LAB11-0418, date of approval: 6/7/2011).

4. Results

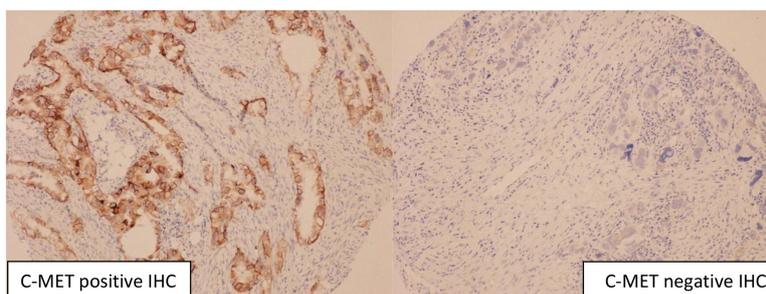
In the first cohort, 274 out of 418 patients were di-

agnosed with high grade ovarian carcinoma. The age ranged from 21.7 years to 92.4 years. The diagnoses were further categorized into 210 high grade serous carcinoma, 11 MMT, 44 mixed malignant carcinoma, and 9 undifferentiated carcinoma. Of the 274 patients, 254 had advanced AJCC stage at the time of diagnosis (stages III & IV) and 13 had low stage (stages I & II). Documentation of ascites was recorded for 199 patients: 184 with clinical symptoms of ascites versus 15 without. Detailed family cancer history was recorded for 251 patients: 130 with family history of some forms of cancer versus 121 without. The disease-free survival time (from treatment to recurrence of or death due to ovarian carcinoma) ranged from 2.4 months to 65.8 months. The overall survival time (from treatment to death due to ovarian carcinoma) ranged from 0.3 months to 279 months. 186 patients died due to high grade ovarian carcinoma, and 88 patients either survived or died due to other causes (Table 1).

In the first cohort, 41 cases were positive for EGFR and 233 cases were negative. For HLA-G, 13 cases

Table 1
Clinical information of cohort 1

Number of patients	274			
Types of carcinoma	210 high grade serous	11 MMT	44 mixed type carcinoma	9 undifferentiated carcinoma
Age range	27.5–92.4 years old			
Stages	13 low stage (I & II)	254 high stage (III & IV)		
Ascites	184 with ascites	15 without ascites		
Family history of any cancer types	130 with family history	121 without family history		
Disease free survival range	2.4–65.8 months			
Overall survival time	0.3–279 months			
Survival status	186 died from ovarian carcinoma	88 alive/died from other causes		



C-MET IHC results			
	CD70 positive	CD70 negative	
Age <51	12	46	p = 0.11
Age ≥51	27	189	
High stage	34	220	p = 0.32
Low stage	3	10	
Ascites	18	166	p = 0.22
No ascites	3	12	
Family history of cancer	21	109	p = 0.14
No family history of cancer	12	109	

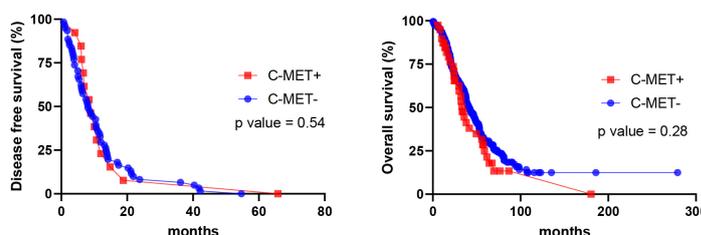


Fig. 4. C-MET results in cohort 1.

Table 2
IHC results for cohort 1

	Positive	Negative
EGFR	41	233
CD70	11	263
C-MET	39	235
HLA-G	13	261
NY-ESO1	23	251

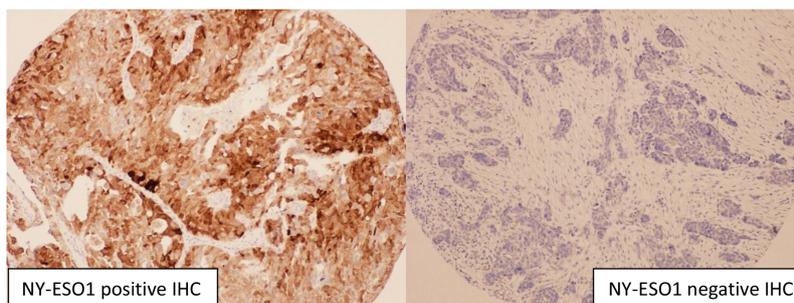
stained positive while 261 cases stained negative For CD70, 11 cases stained positive while 263 cases stained negative. For C-MET, 39 cases stained positive while

235 cases stained negative. For NY-ESO1, 23 cases stained positive while 251 cases stained negative (Table 2).

EGFR-positive and negative stains were stratified into different groups based on age (< 51 versus ≥ 51 years old), AJCC stage (low versus high), clinical symptoms of ascites and family cancer history. The only group that had clinical significance was the one related to family cancer history, with the *p*-value of 0.01. The other three groups did not demonstrate any clinical significance (*p*-value > 0.05). Additionally, Kaplan-Meier

Table 3
Clinical information of cohort 2

Number of patients	248			
Types of carcinoma	218 high grade serous	6 MMMT	24 mixed type carcinoma	1 undifferentiated carcinoma
Age range	39.7–85.4 years old			
Disease free survival range	1–152.7 months			
Overall survival time	0.3–227 months			
Survival status	72 died from ovarian carcinoma		176 alive/died from other causes	



NY-ESO1 IHC results in cohort 1			
	NY-ESO1 positive	NY-ESO1 negative	
Age <51	2	56	p = 0.13
Age ≥51	21	195	
High stage	21	233	p = 0.38
Low stage	2	11	
Ascites	16	168	p = 0.55
No ascites	2	13	
Family history of cancer	10	120	p = 0.53
No family history of cancer	12	109	

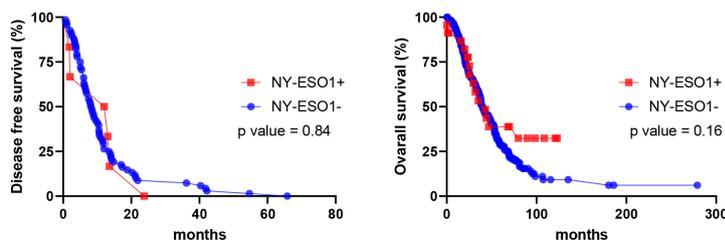


Fig. 5. NY-ESO1 results in cohort 1.

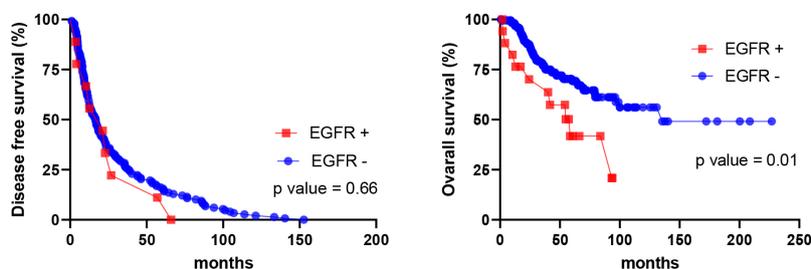


Fig. 6. EGFR expression vs DFS (left) and OS (right) in cohort 2.

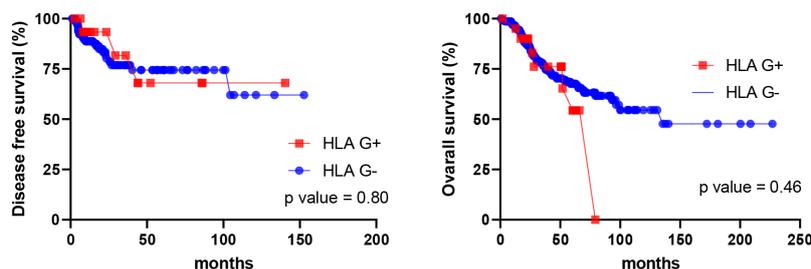


Fig. 7. HLA-G expression vs DFS (left) and OS (right) in cohort 2.

curves were used to compare the DFS and OS in EGFR positive and EGFR-negative cases. No clinical significance was observed with respect to DFS intervals (p -value = 0.70; EGFR-positive median of 9.8 months versus EGFR-negative median of 8.5 months). However, there was clinical significance in term of OS rate (p -value < 0.0001; EGFR-positive median of 24 months versus EGFR-negative median of 43.1 months) (Fig. 1).

The same statistical analysis was performed for HLA-G. There was no significant difference between HLA-G status versus age, AJCC stage, clinical symptoms of ascites, and family history of cancer. In addition, HLA-G staining pattern is not associated with DFS. However, with the OS, HLA-G-positive expression is associated with shorter survival compared to HLA-G negative expression (p -value = 0.027; HLA-G-positive median of 21.4 months versus HLA-G-negative median of 41 months) (Fig. 2).

For CD70, C-MET, and NY-ESO1 immunohistochemical stains, there was no significant difference between each individual stain versus age, AJCC stage, clinical symptoms of ascites, and family history of cancer. Furthermore, the Kaplan-Meier curve showed that there was no correlation between each stain versus DFS and OS (Figs 3–5).

A second separate cohort was used to confirm the association between DFS and OS versus EGFR and HLA-G staining patterns. In this cohort, 248 out of 453 females were diagnosed with high grade ovarian carcinoma. The age ranged from 39.7 years to 85.4 years. The diagnoses were categorized into 218 high grade serous carcinoma, 5 MMT, 24 mixed malignant carcinoma, and 1 undifferentiated carcinoma. The DFS intervals ranged from 1 to 152.7 months, and the OS ranged from 0.6 to 227 months. 72 patients died from ovarian carcinoma, and 176 patients died from other causes or are still alive (Table 3). For EGFR, Kaplan-Meier curve showed no clinical significance with the DFS intervals (p -value = 0.66; EGFR-positive median of 21.3 months versus EGFR-negative median of 16.65

months). However, there was clinical significance with the OS rate (p -value = 0.01; EGFR-positive median of 58.5 months versus EGFR-negative median of 135.5 months) (Fig. 6). For HLA-G analysis there was no clinical significance in both DFS (p -value = 0.8) and OS rates (p -value = 0.46) (Fig. 7).

5. Discussion/conclusion

Ovarian epithelial tumors are generally separated into two major categories: type I versus type II. Type I carcinoma follows a step-wise progression from benign histology to borderline tumors to low grade malignancy. Molecularly, these tumors are associated with KRAS, BRAF, PTEN, PIK3CA or ARID1A mutations. Clinically, they grow locally, metastasize late, and behave in a less aggressive manner [9]. Type II carcinoma, on the other hand, is frequently discovered at an advanced stage and is more aggressive. Molecularly, it is associated with TP53 mutations, hypermethylation, or BRCA1/2 mutations [10]. Histologically, it consists of high grade serous carcinoma, mixed type carcinoma, malignant mixed Mullerian tumor, and undifferentiated carcinoma. In this study, we compare the clinical data of patients with high grade carcinoma against the immunohistochemical expression of EGFR, HLA-G, CD70, NY-ESO1, and C-Met, in the hope that they can be used as biomarkers or part of the therapeutic regime in the future.

5.1. EGFR

Epidermal growth factor receptor (EGFR) is a receptor tyrosine kinase protein that functions in cell development, autophagy and metabolism regulation. Mutations in EGFR have been associated with different cancer types, such as lung, colon, and pancreas [11, 12]. In our analysis, we find an association between EGFR staining and family history of different cancer

types. This supports the idea that EGFR plays a vital role in neoplastic development, not limited to a specific type. Furthermore, with the association of family history, mutations in EGFR may be part of the syndromic cause for carcinoma. We also performed an analysis to associate the staining pattern of EGFR to other clinical data (age, clinical symptoms of ascites, and staging), but we found no correlation.

Additionally, we demonstrate that EGFR expression is significantly associated with a worse OS rate, with a median of 24 months versus 43.1 months in non-EGFR patients. To confirm the result, a separate cohort is used, and a similar outcome is concluded: EGFR expression is associated with a shorter median survival of 58.5 months, while the non-EGFR population is associated with a longer median survival of 135.5 months. Our result is inconsistent with Mehner's study, which has a sample size of 488. Her study concludes that there is no association between EGFR staining patterns and survival outcomes. However, Mehner's study incorporates different types of ovarian epithelial carcinoma (low grade serous, mucinous, endometrioid, clear cell, high grade serous, and mixed type) [13]. Our study, on the other hand, is more specific for high grade carcinoma (high grade serous, MMT, mixed type, and undifferentiated). Thus, the difference in tumor type selection could explain the disparity between the two studies.

Preclinical study of anti-EGFR antibody shows promising results, as tumor cells exhibit increased sensitivity to NK cell-mediated antibody-dependent cellular cytotoxicity [14]. With the correlation of worse prognosis, EGFR can serve as a potential biomarker and therapeutic marker in the future.

5.2. HLA-G

Human leukocyte antigen-G (HLA-G) is expressed within the placental and embryonic tissues. It plays a role in the tolerance of the maternal immune system by inhibiting the differentiation, proliferation, and cytokine release from different immune cell types. It is postulated that tumor cells use HLA-G to evade immune surveillance [15]. HLA-G mRNA and protein levels, as well as immunohistochemical expression, have been found in greater quantity in high stage and high grade epithelial ovarian cancer [16,17]. In our analysis, we do not find any correlation between HLA-G expression and age, clinical symptoms of ascites, advanced AJCC stage, or DFS intervals. When analyzing the OS rate, a correlation between positive HLA-G expression

and worse OS rate is observed (p -value = 0.02; 21.4 months versus 41 months). However, the analysis in the second cohort yielded a contradicting result, with no correlation (p -value = 0.46). Nevertheless, the trend is promising, as the HLA-G negative expression seems to have a better survival trend.

Literature review of HLA-G associated survival rates has yielded mixed results. The conclusion from Jung's study suggests an unfavorable prognosis, while Rutten's study shows the contrary. The discrepancy might be due to different sample selections. Jung's cohort includes both type I and II ovarian carcinoma, while Rutten's cohort includes only type II ovarian carcinoma [16,18]. In addition, both our study and Rutten's study use tissue microarrays, but with different methods of scoring. Rutten's study stratifies the staining results by adding the score of staining percentage (0-5) and intensity (0-3). Positive HLA-G staining has a score of ≥ 7 . We think that this method might decrease the sensitivity of the HLA-G immunohistochemical stain. In our study, we consider positive staining as any cytoplasmic staining. Irrespective of the scoring method, the conclusion regarding the prognostic significance of HLA-G is still undetermined.

5.3. CD70

CD70 is a membranous protein that belongs to the tumor necrosis factor family, its activation leads to cellular proliferation. CD70 expression is associated with cisplatin resistance and decreased survival rates in ovarian carcinoma [19]. Additionally, CD70 antibody-drug conjugate has shown to inhibit the proliferation of ovarian carcinoma cells that express CD70 both *in vitro* and *in vivo* [20]. Therefore, CD70 has the potential to be a biomarker and therapeutic marker. However, our study only has 11 cases with CD70 positivity. Analysis of CD70 expression does not correlate with age, AJCC stage, clinical symptoms of ascites, DFS, or OS rate (all p -values > 0.05). Nevertheless, the OS curve aligns with the literature studies, with positive CD70 expression showing a worse OS trend than negative CD70. The lack of a significant p -value in our analysis might be due to the selection bias. During the construction of tissue microarray, only a minute portion of the entire ovarian carcinoma is randomly selected for each case. Because of tumor heterogeneity, the representation on the tissue microarray might not accurately represent the IHC expression of the remaining carcinomatous cells. A further study with more comprehensive tissue selection is recommended to assess the clinical significance of CD70.

5.4. C-MET

C-mesenchymal-epithelial transition factor (c-MET) is a tyrosine kinase receptor that plays a role in embryogenesis and tissue regeneration [21]. Normally, hormone regulation promotes c-MET and HGF to enhance the proliferation of the ovarian surface to repair the area damaged by ovum expulsion. In ovarian carcinoma, hormonal regulation is lost, leading to constant activation from c-MET and HGF signaling. This results in proliferation, stromal invasion, and metastasis of the carcinoma. Thus, c-MET could serve as a potential biomarker [22]. In our study, we do not find any correlation of c-MET expression and advanced age (indirectly related to hormonal regulation), clinical symptoms of ascites, or advanced stages (indirectly related to metastasis). There is also no association of c-MET with DFS or OS rate. Therefore, c-MET might not be an ideal candidate for ovarian carcinoma biomarker.

5.5. NY-ESO1

New York esophageal squamous carcinoma 1 (NY-ESO1) is expressed at the fetal level within the germ cells. As the germ cells undergo spermatid differentiation, NY-ESO1 expression is physiologically lost. Its expression is affected by tumor grade, stage, or therapeutic intervention [23]. NY-ESO1 expression is also associated with poor clinical outcome in ovarian carcinoma [24]. However, our study does not find any association between NY-ESO1 expression and age, clinical symptoms of ascites, or advanced stages (indirectly related to metastasis). NY-ESO1 also does not correlate with DFS or OS rates. Therefore, NY-ESO1 might not be an ideal candidate for tumor marker.

With the total sample size of 522 from two separate cohorts, our study is the largest clinicopathological study of EGFR, HLA-G, CD70, c-MET, and NY-ESO1 expressions in high grade ovarian carcinoma. The sample size strongly supports the clinical significance of EGFR and HLA-G expression in predicting a worse prognosis. CD70, c-MET, and NY-ESO1, on the other hand, do not correlate with prognosis.

Unlike Her-2, the five immunohistochemical stains that we analyze do not have consensus interpretation. In different EGFR studies, methods that have been used include low (< 10% positivity) versus high (> 10%); membranous versus cytoplasmic; 1+ to 3+ score; and any > 1% staining [20,25,26,27]. HLA-G interpretations include positive membranous or combined membranous and cytoplasmic, and low-middle-strong [15, 17]. CD70 also has similar interpretations [19,28]. C-

Met has a standard score for gastric tumors, from 0 (< 50% with weak membranous staining) to 3+ (> 50% with strong membranous staining) [29]. However, there is currently no standard score for ovarian tumors. For NY-ESO1 interpretation, one study from Roswell Park Cancer Institute stratifies membranous or cytoplasmic staining as negative (< 5%) to 4+ (> 75%) [24]. Our study stratified the result into negative versus positive. Positive expression includes > 1% membranous staining for EGFR, CD70, c-MET; or > 1% cytoplasmic staining for HLA-G and NY-ESO1. Since staining is performed on tissue microarrays, only a small portion of the tumor from each case is presented. Due to the heterogeneity of ovarian epithelial tumors, using a higher percentage of stained tumor cells to be considered positive might decrease the sensitivity of the study. Therefore, we decide to use > 1% membranous or cytoplasmic staining to be positive.

We analyzed a panel of membrane markers including EGFR, HLA-G, CD70, c-MET, and NY-ESO1 using the largest cohort of ovarian cancer. We found that immunohistochemical expressions of EGFR are associated with worse prognosis. The expression of HLA-G also correlates with poor prognosis, either statistically significant or trend. Although CD70 expression has no statistically significant association with prognosis, it does follow the trend of poor prognosis. C-MET and NY-ESO1 do not have any correlation with OS rate or other clinical data.

Targeted therapy, such as EGFR inhibitors (erlotinib, cetuximab or lapatinib), has proven to be non-effective in ovarian carcinoma treatment [30]. However, the results from this study can be helpful for designing future clinical immunotherapy trials, especially for EGFR and HLA-G and their roles in immune regulation. They can be used as a prerequisite study prior to placing patients with disease recurrence on immunotherapy. In addition, if there are fruitful results in clinical trials, these biomarkers can be incorporated into routine clinical pathology practice. They are as affordable as other diagnostic immunohistochemical markers.

Acknowledgments

This research is supported in part by NIH/NIC to MD Anderson SPORT in Ovarian Cancer, Award Number 1P50CA281701-01. The authors declare no conflict of interest.

Author contributions

Conceptualization: Duc Vo and Jinsong Liu.

Interpretation or analysis of data: Duc Vo and Jinsong Liu.

Preparation of the manuscript: Duc Vo.

Revision for important intellectual content: Yan Liu, Anil K Sood, Katy Rezvani, Amir A Jazaeri.

Supervision: Jinsong Liu.

References

- [1] Z. Momenimovahed et al., Ovarian cancer in the world: Epidemiology and risk factors, *Int J Womens Health* **11** (2019), 287–299.
- [2] R.J. Kurman and L.M. Shih, The dualistic model of ovarian carcinogenesis: Revisited, revised, and expanded, *Am J Pathol* **186**(4) (2016), 733–747.
- [3] L.A. Torre et al., Ovarian cancer statistics, 2018, *CA Cancer J Clin* **68**(4) (2018), 284–296.
- [4] A. Gockley et al., Outcomes of women with high-grade and low-grade advanced-stage serous epithelial ovarian cancer, *Obstet Gynecol* **129**(3) (2017), 439–447.
- [5] L.Y. Guan and Y. Lu, New developments in molecular targeted therapy of ovarian cancer, *Discov Med* **26**(144) (2018), 219–229.
- [6] X. Jiang et al., Immune checkpoint: The novel target for anti-tumor therapy, *Genes Dis* **8**(1) (2021), 25–37.
- [7] S. Morand et al., Ovarian cancer immunotherapy and personalized medicine, *Int J Mol Sci* **22**(12) (2021).
- [8] G. Varady et al., Cell surface membrane proteins as personalized biomarkers: Where we stand and where we are headed, *Biomark Med* **7**(5) (2013), 803–819.
- [9] K.Y. Terada, H.J. Ahn and B. Kessel, Differences in risk for type 1 and type 2 ovarian cancer in a large cancer screening trial, *J Gynecol Oncol* **27**(3) (2016), e25.
- [10] M. Koshiyama, N. Matsumura and I. Konishi, Recent concepts of ovarian carcinogenesis: Type I and type II, *Biomed Res Int* **2014** (2014), 934261.
- [11] S. Sigismund, D. Avanzato and L. Lanzetti, Emerging functions of the EGFR in cancer, *Mol Oncol* **12**(1) (2018), 3–20.
- [12] W.L. Cheng et al., The Role of EREG/EGFR Pathway in Tumor Progression, *Int J Mol Sci* **22**(23) (2021).
- [13] C. Mehner et al., EGFR as a prognostic biomarker and therapeutic target in ovarian cancer: Evaluation of patient cohort and literature review, *Genes Cancer* **8**(5-6) (2017), 589–599.
- [14] N. Mallmann-Gottschalk et al., EGFR-Specific Tyrosine Kinase Inhibitor Modifies NK Cell-Mediated Antitumoral Activity against Ovarian Cancer Cells, *Int J Mol Sci* **20**(19) (2019).
- [15] A. Lin and W.H. Yan, Human Leukocyte Antigen-G (HLA-G) Expression in Cancers: Roles in immune evasion, metastasis and target for therapy, *Mol Med* **21**(1) (2015), 782–791.
- [16] Y.W. Jung et al., Correlation of human leukocyte antigen-G (HLA-G) expression and disease progression in epithelial ovarian cancer, *Reprod Sci* **16**(11) (2009), 1103–1111.
- [17] C. Menier et al., Human leukocyte antigen-G is expressed in advanced-stage ovarian carcinoma of high-grade histology, *Hum Immunol* **70**(12) (2009), 1006–1009.
- [18] M.J. Rutten et al., HLA-G expression is an independent predictor for improved survival in high grade ovarian carcinoma, *J Immunol Res* **2014** (2014), 274584.
- [19] N. Liu et al., Increased CD70 expression is associated with clinical resistance to cisplatin-based chemotherapy and poor survival in advanced ovarian carcinoma, *Onco Targets Ther* **6** (2013), 615–619.
- [20] M. Shiomi et al., CD70 antibody-drug conjugate: A potential novel therapeutic agent for ovarian cancer, *Cancer Sci* **112**(9) (2021), 3655–3668.
- [21] Y. Zhang et al., Function of the c-Met receptor tyrosine kinase in carcinogenesis and associated therapeutic opportunities, *Mol Cancer* **17**(1) (2018), 45.
- [22] Y. Kwon and A.K. Godwin, Regulation of HGF and c-MET Interaction in Normal Ovary and Ovarian Cancer, *Reprod Sci* **24**(4) (2017), 494–501.
- [23] A. Raza et al., Unleashing the immune response to NY-ESO-1 cancer testis antigen as a potential target for cancer immunotherapy, *J Transl Med* **18**(1) (2020), 140.
- [24] J.B. Szender et al., NY-ESO-1 expression predicts an aggressive phenotype of ovarian cancer, *Gynecol Oncol* **145**(3) (2017), 420–425.
- [25] P. de Graeff et al., The ErbB signalling pathway: protein expression and prognostic value in epithelial ovarian cancer, *Br J Cancer* **99**(2) (2008), 341–349.
- [26] C.K. Lin et al., The expression of six biomarkers in the four most common ovarian cancers: Correlation with clinicopathological parameters, *APMIS* **117**(3) (2009), 162–175.
- [27] A. Noske et al., An intracellular targeted antibody detects EGFR as an independent prognostic factor in ovarian carcinoma, *BMC Cancer* **11** (2011), 294.
- [28] A. De Meulenaere et al., CD70 Expression and Its Correlation with Clinicopathological Variables in Squamous Cell Carcinoma of the Head and Neck, *Pathobiology* **83**(6) (2016), 327–333.
- [29] N. Fuse et al., Prognostic impact of HER2, EGFR, and c-MET status on overall survival of advanced gastric cancer patients, *Gastric Cancer* **19**(1) (2016), 183–191.
- [30] A. Cortez et al., Advances in ovarian cancer therapy, *Cancer Chemotherapy Pharmacology* **81**(1) (2018), 17–38.