

Letter to the Editor

Application of the Minkowski–Bouligand fractal dimension for the differential diagnosis of thyroid follicular neoplasias

Keywords: Thyroid, adenoma, minimally invasive carcinoma, complexity, texture analysis, fractality, karyometry, surgical pathology

To the Editor,

Nuclear characteristics are important for the differential diagnosis between benign or malignant neoplasias. Subjective interpretation by an observer, however, may cause diagnostic insecurity [5]. Quantitative morphologic analyses can be helpful in this situation [19]. There are several ways to perform karyometry, which may be done by basic morphometry [8,10,14,16,17,21–24,28], or by sophisticated texture analysis based on digitalized images [2–7,11–13,20,25–27,29–31]. Due to the fractal nature of chromatin organization in interphase nuclei [15], the scale-invariant self similarity is an important texture feature, which can be estimated by the fractal dimension (FD) [1,3,9,13,31]. For the analysis of nuclear chromatin in routinely stained slides it has been suggested that besides the FD, the goodness-of-fit (GOF) of the regression line in the log-log plots, which are essential for the FD estimation, could be important [3]. In this investigation we tried to find out whether this new parameter GOF could also be useful for differential diagnosis in surgical pathology. We compared the FD and GOF of nuclear chromatin in routinely HE stained paraffin sections of follicular adenomas and minimally invasive follicular carcinomas of the thyroid.

Our study consisted of 18 follicular adenomas and 24 microinvasive follicular carcinomas from our files. Tumors with a diameter up to 6 cm had been completely embedded in paraffin. For larger tumors the number of paraffin blocks taken had been $d + 4$ (d representing the largest diameter in cm). Diagnosis was based on criteria of the World Health Organization Histological Classification [18]. From each tumor 100 nuclei were randomly taken from routinely stained 5 μm HE stained paraffin sections using the Kontron Zeiss KS300 system (0.1 μm /pixel spatial resolution; 1.25 numerical aperture; 100 \times oil immersion objective) by one examiner blinded to the diagnosis. The

images were converted to grayscale format with levels of luminance ranging between 0 (absence of light) and 255 (very bright). Then a pseudo-three-dimensional “landscape-like” representation was created using the gray level (luminance) of each pixel (picture element) as the height of a z-axis. The fractal dimension (FD) was determined according to Minkowski–Bouligand extended to three dimensions, as described earlier [3]. The linear regression was estimated in a log-log plot composed of 30 points. The goodness-of-fit (GOF) was determined by the R^2 value of the regression between the real and the estimated values. For each nucleus the distribution of the residuals was compared with the normal distribution by the Kolmogorov–Smirnov test. Then for each patient the percentage (P) of cells with normally distributed residuals was calculated [3]. Group comparison was done by the Mann–Whitney test (WinStat software).

Patients of both groups were of similar age with a mean for adenomas of 40.3 years (range 17–72 years) and a mean for minimally invasive carcinomas of 49.5 years (range 19 to 88 years; $p > 0.05$). The same was true for the tumor size (mean for adenomas: 3.6 cm, range 1.0–7.0 cm; mean for minimally invasive carcinomas: 3.4 cm, range 1.3 to 8.5 cm; $p > 0.05$). There was no statistically significant difference ($p > 0.05$) of the FD values between both groups: adenoma mean value 2.424 (range 2.347 to 2.505); minimally invasive carcinomas: 2.432 (range 2.339 to 2.499). The R^2 values, however, were significantly different ($p < 0.05$), ranging between 0.922 and 0.953 (mean 0.939) in adenomas and between 0.929 and 0.952 (mean 0.943) in minimally invasive carcinomas. The percentage of cells with a Gaussian distribution of the residuals ranged from 58 to 97% (mean 79%) for adenomas and between 48 and 94% (mean 75%) for minimally invasive carcinomas.

The fractal dimension is derived from the slope of the curve in a log–log–diagram, assuming that this

curve can be well approximated by a straight line with an excellent GOF, equivalent to a high R^2 value and a normal distribution of the residuals. Since in our study many cells did not fulfill these criteria, we conclude that the calculated FDs should only be interpreted with caution. It is interesting to note that the GOF showed a statistically significant difference between both groups, whereas this was not the case for the FD. This situation is similar to an earlier study [3], where it had been shown that the GOF, but not the Minkowski–Bouligand FD, was an independent prognostic variable for patients suffering from B precursor acute lymphoid leukemia. Therefore we think that the GOF should be considered a new variable for texture analysis, especially for nuclear chromatin, where it was suggested to be a measure of “coarseness” [3]. According to international standards the differential diagnosis between adenoma and minimally invasive follicular carcinoma of the thyroid is only based on the criteria of capsular or vessel invasion but not on nuclear characteristics. Our study shows that there are subtle differences in the coarseness of the chromatin structure between the two entities, normally not visible to the human eye.

Rita C. Ferreira^a, Patrícia S. de Matos^a,
Randall L. Adam^b, Neucimar J. Leite^b,
Konradin Metz^{a,*}

^a Department of Pathology, Faculty of Medicine,
State University of Campinas

^b Institute of Computing, State University
of Campinas, BR 13081–970 Campinas – SP, Brazil

* Corresponding author: Prof Dr Konradin Metz
Senior researcher
of the National Research Council CNPq
Department of Pathology, Faculty of Medicine
PO Box 6111, State University of Campinas
BR 13081–970 Campinas – SP, Brazil
E-mail: kmetze@fcm.unicamp.br
Tel./Fax: 55 19 32893897

References

- [1] R.L. Adam, T.C.G. Corsini, P.V. Silva, M.L. Cintra, N.J. Leite and K. Metz, Fractal dimensions applied to thick contour detection and residues – Comparison of keloids and hypertrophic scars, *Cytometry*, Part A **59A** (2004), 63–64.
- [2] R.L. Adam, E. Ribeiro, K. Metz, N.J. Leite and I. Lorand-Metze, Morphometric and granulometric features of erythroblasts as a diagnostic tool of hematologic diseases, *Cytometry*, Part A **59A** (2004), 46.
- [3] R.L. Adam, R.C. Silva, F.G. Pereira, N.J. Leite, I. Lorand-Metze and K. Metz, The fractal dimension of nuclear chromatin as a prognostic factor in acute precursor B lymphoblastic leukemia, *Cellular Oncology* **28** (2006), 55–59.
- [4] M.P. Auada, R.L. Adam, N.J. Leite, M.B. Puzzi, M.L. Cintra, W.B. Rizzo and K. Metz, Texture analysis of the epidermis based on fast Fourier transformation in Sjögren–Larsson syndrome, *Analytical and Quantitative Cytology and Histology* **28** (2006), 219–227.
- [5] J.P. Baak, Quantitative histopathological analysis of CIN sections, *Cellular Oncology* **28** (2006), 61–62.
- [6] A. Böcking, J. Stockhausen and D. Meyer-Ebrecht, Towards a single cell cancer diagnosis. Multimodal and monocellular measurements of markers and morphology (5M), *Cellular Oncology* **26** (2004), 73–79.
- [7] D. Chiu, M. Guillaud, D. Cox, M. Follen and C. Macaulay, Quality assurance system using statistical process control: an implementation for image cytometry, *Cellular Oncology* **26** (2004), 101.
- [8] E.M.M. Cia, M. Trevisan and K. Metz, Argyrophilic nucleolar organizer region (AgNOR) technique: a helpful tool for differential diagnosis in urinary cytology, *Cytopathology* **10** (1999), 30–39.
- [9] A. Gerger, P. Bergthaler and J. Smolle, An automated method for the quantification and fractal analysis of immunostaining, *Cellular Oncology* **26** (2004), 125–134.
- [10] M.F. Gilberti, K. Metz and I. Lorand-Metze, Changes of nucleolar organizer regions in granulopoietic precursors during the course of chronic myeloid leukemia, *Annals of Hematology* **71** (1995), 275–279.
- [11] M. Guillaud, D. Cox, A. Malpica, G. Staerkel, J. Maticic, D. van Niekirk, K. Adler-Storthz, N. Poulin, M. Follen and C. MacAulay, Quantitative histopathological analysis of cervical intra-epithelial neoplasia sections: Methodological issues, *Cellular Oncology* **26** (2004), 31–43.
- [12] M. Guillaud, M. Follen and C. MacAulay, Quantitative histopathological analysis of CIN sections, *Cellular Oncology* **28** (2006), 63–65.
- [13] A. Huisman, L.S. Ploeger, H.F. Dullens, N. Poulin, W.E. Grizzle and P.J. van Diest, Development of 3D chromatin texture analysis using confocal laser scanning microscopy, *Cellular Oncology* **27** (2005), 335–345.
- [14] S.P. Irazusta, J. Vassallo, L.A. Magna, K. Metz and M. Trevisan, The value of PCNA and AgNOR staining in endoscopic biopsies of gastric mucosa, *Pathology Research and Practice* **194** (1998), 33–39.
- [15] D.V. Lebedev, M.V. Filatov, A.I. Kuklin, A.Kh. Islamov, E. Kentzinger, R. Pantina, B.P. Toperverg and V.V. Isaev-Ivanov, Fractal nature of chromatin organization in interphase chicken erythrocyte nuclei: DNA structure exhibits biphasic fractal properties, *FEBS Letters* **579** (2005), 1465–1468.
- [16] I. Lorand-Metze, M.A. Carvalho and K. Metz, Relationship between morphometric analysis of nucleolar organizer regions and cell proliferation in acute leukemias, *Cytometry* **32** (1998), 51–56.

- [17] I. Lorand-Metze, F.G. Pereira, F.P. Costa and K. Metze, Proliferation in non-Hodgkin's lymphomas and its prognostic value related to staging parameters, *Cellular Oncology* **26** (2004), 63–71.
- [18] P.S. de Matos, A.P. Ferreira, F.O. Facuri, L.V. Assumpcao, K. Metze and L.S. Ward, Usefulness of HBME-1, cytokeratin 19 and galectin-3 immunostaining in the diagnosis of thyroid malignancy, *Histopathology* **47** (2005), 391–401.
- [19] K. Metze and R.A. Adam, Quantification in histopathology – some pitfalls, *Brazilian Journal of Medical and Biological Research* **38** (2005), 141–143.
- [20] K. Metze, V. Bedin, R.L. Adam, M.I. Cintra, E.M. de Souza and N.J. Leite, Parameters derived from the fast Fourier Transform are predictive for the recurrence of basal cell carcinoma, *Cellular Oncology* **27** (2005), 137.
- [21] K. Metze, A.C. Chiari, F.L. Andrade and I. Lorand-Metze, Changes in AgNOR configurations during the evolution and treatment of chronic lymphocytic leukemia, *Hematology and Cell Therapy* **41** (1999), 205–210.
- [22] K. Metze, A.M. Lobo and I. Lorand-Metze, Nucleolus organizer regions (AgNORs) and total tumor mass are independent prognostic parameters for treatment-free period in chronic lymphocytic leukemia, *International Journal of Cancer* **89** (2000), 440–443.
- [23] K. Metze and I. Lorand-Metze, Interpretation of the AgNOR pattern in hematologic cytology, *Acta Haematologica* **89** (1993), 110.
- [24] K. Metze, G.B. Oliveira, F.G. Pereira, R.L. Adam and I. Lorand-Metze, Spontaneous apoptosis in chronic lymphocytic leukemia is not an independent prognostic factor for stability of disease when compared with combined AGNOR and TTM scores, *Cellular Oncology* **27** (2005), 199–201.
- [25] K. Metze, A.C.S. Piazza, A.A. Piazza, R.L. Adam and N.J. Leite, Texture analysis of agnor stained nuclei in lung cancer, *Cellular Oncology* **27** (2005), 137–138.
- [26] K. Metze, R.C. Silva, R.L. Adam, N.J. Leite, F.G. Pereira and I. Lorand-Metze, Relation between chromatin texture and phenotype in acute leukemias, *Cell. Oncol.* **27** (2005), 112–113.
- [27] B. Nielsen and H.E. Danielsen, Prognostic value of adaptive textural features – the effect of standardizing nuclear first-order gray level statistics and mixing information from nuclei having different area, *Cellular Oncology* **28** (2006), 85.
- [28] G.B. Oliveira, F.G. Pereira, K. Metze and I. Lorand-Metze, Spontaneous apoptosis in chronic lymphocytic leukemia and its relationship to clinical and cell kinetic parameters, *Cytometry* **46** (2001), 329–335.
- [29] L.S. Ploeger, Computer science meets medical science, *Cellular Oncology* **28** (2006), 69–70.
- [30] L.S. Ploeger, J.A. Belien, N.M. Poulin et al., Confocal 3D DNA cytometry: assessment of required coefficient of variation by computer simulation, *Cellular Oncology* **26** (2004), 93–99.
- [31] L.B. Rocha, R.L. Adam, N.J. Leite, K. Metze and M.A. Rossi, Biomineralization of polyanionic collagen–elastin matrices during cavariar bone repair, *Journal of Biomedical Material Research A* **79** (2006), 237–245.