

Review

Immunogenetic Determinants of Parkinson's Disease Etiology

Pin-Jui Kung^{a,1}, Inas Elsayed^{b,c,1}, Paula Reyes-Pérez^{d,1} and Sara Bandres-Ciga^{e,1,*}

^a*Genome and Systems Biology Degree Program, National Taiwan University and Academia Sinica, Taipei, Taiwan*

^b*Faculty of Pharmacy, University of Gezira, Wad Medani, Sudan*

^c*International Parkinson Disease Genomics Consortium (IPDGC)-Africa, University of Gezira, Wad Medani, Sudan*

^d*Laboratorio Internacional de Investigacion sobre el Genoma Humano, Universidad Autonoma de Mexico, Queretaro, Mexico*

^e*Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, MD, USA*

Accepted 11 March 2022

Pre-press 31 March 2022

Abstract. Parkinson's disease (PD) is increasingly recognised as a systemic disorder in which inflammation might play a causative role rather than being a consequence or an epiphenomenon of the neurodegenerative process. Although growing genetic evidence links the central and peripheral immune system with both monogenic and sporadic PD, our understanding on how the immune system contributes to PD pathogenesis remains a daunting challenge. In this review, we discuss recent literature aimed at exploring the role of known genes and susceptibility loci to PD pathogenesis through immune system related mechanisms. Furthermore, we outline shared genetic etiologies and interrelations between PD and autoimmune diseases and underlining challenges and limitations faced in the translation of relevant allelic and regulatory risk loci to immune-pathological mechanisms. Lastly, with the field of immunogenetics expanding rapidly, we place these insights into a future context highlighting the prospect of immune modulation as a promising disease-modifying strategy.

Keywords: Parkinson's disease, immune system, immunogenetics, risk, onset, progression

INTRODUCTION

Heterogeneous and multifactorial in nature, Parkinson's disease (PD) follows a complex model of inheritance spanning the etiological spectrum ranging from monogenic disease (in a small proportion of affected individuals) to polygenic inheritance (in the vast majority of the cases) where environmental and genetic risk factors interact to induce PD pathology.

Emerging and compelling evidence supports chronic neuroinflammation, derived from impaired innate and/or adaptive immunity mechanisms, as among the main contributors to PD development, in which a pro-inflammatory state may trigger or promote neuronal loss [1]. A growing body of research recognises PD as a systemic disease characterised by the presence of central and peripheral inflammatory processes.

In recent years, extensive research in the PD genetics field has focused on unravelling both coding and non-coding genetic variation that could result in immune defects contributing to PD risk, onset, and progression. Indeed, the advent of

¹These authors contributed equally to this work.

*Correspondence to: Sara Bandres-Ciga, PhD, Molecular Genetics Section, Laboratory of Neurogenetics, NIA, NIH, Bethesda, MD 20892, USA. E-mail: sara.bandresciga@nih.gov.

high-throughput, high-resolution next-generation sequencing and genome-wide genotyping technologies has been instrumental in the identification of genes and regulatory genomic regions that play a prominent role in immune processes leading to the pathophysiology of the disease.

This review aims to provide an updated overview of the role of monogenic PD genes in the regulation of the immune system as well as our current knowledge on the contribution of recently identified immunogenetic risk factors leading to neurodegeneration in PD. We outline the challenges faced in the translation of relevant allelic and regulatory risk loci to immune-pathological mechanisms and discuss potential shared genetic etiologies between PD and autoimmune diseases. Furthermore, we illustrate the increasing need to generate harmonised large-scale omics, clinical and longitudinal data to enhance our understanding of the immune system involvement in PD etiology that could aid future development of immunomodulatory therapeutic interventions aimed to prevent or delay disease onset.

THE ROLE OF MONOGENIC PARKINSON'S DISEASE KNOWN GENES IN THE REGULATION OF THE IMMUNE SYSTEM

Monogenic PD—PD caused by a defined mutation in a single gene—constitutes about 5–10% of the total PD cases [2]. Mutations in several genes have been identified as causative for monogenic PD with different patterns of inheritance including autosomal dominant, autosomal recessive, and X-linked [3]. Scrutinising the molecular processes through which mutations in such genes culminate in the development of PD, modulation of immune-related pathways emerged as a potential pathogenic mechanism [4, 5]. This has stirred research to dissect the potential roles of monogenic PD-known genes in regulating the immune response [6].

LRRK2

The leucine-rich repeat kinase 2 (*LRRK2*) gene has been linked to monogenic as well as sporadic PD. *LRRK2* mutations are the most frequently reported genetic factors in monogenic PD, representing 5–6% of familial PD cases and 1–2% of sporadic PD with variability in prevalence between different populations [7, 8]. The incomplete penetrance

of *LRRK2* indicates the involvement of other genetic/environmental components, which might act modifying the pathogenic effects of such mutations [9, 10]. *LRRK2* gene encodes a multi-function protein with kinase and GTPase activity that is found associated with PD pathogenesis. The structural homology of *LRRK2* with receptor-interacting protein kinases, a family of kinases with known implications in regulating the immune system, highlights the potential role of *LRRK2* in regulating immune-related pathways that might contribute to PD pathogenesis [11]. This has been further supported by the fact that *LRRK2* expression in peripheral immune cells (such as B-cell, T-cell, and monocytes) is markedly upregulated in PD with a coinciding upsurge in the activity of these immune cells [12]. Results from numerous studies advocate that *LRRK2* modulates immune responses to favor sustained inflammation and subsequent neurodegeneration in PD patients. For instance, mutated *LRRK2* was found to promote neuroinflammation through increasing microglia and macrophages chemotaxis to the brain tissues via modulating pathways controlling cell adhesion, polarisation, and directional motility. *LRRK2* is also implicated in modulating processes like phagocytosis, and the production of proinflammatory cytokines [13, 14]. Furthermore, *LRRK2* mediates such effects by stimulating receptors like TLR2 and TLR4, and through several downstream kinases, mediators, and cytokines [4, 15–17]. It is noteworthy that *LRRK2* effects on the immune response are genotype dependent. This has been observed in mouse models where a gain-of-function mutation (p.G2019S) in the *LRRK2* kinase protein resulted in enhanced inflammatory responses, while the kinase inactivating mutation (p.D1994S) produced an opposite effect.

Additionally, studies have shown that *LRRK2* mutants contribute to neurodegeneration through exacerbating neuroinflammation via peripheral pro-inflammatory cytokines. According to recent reports, *LRRK2* maintains neuroinflammation through stimulating peripheral immune cells type II interferon responses rather than effects on central microglia [18]. Within the same context, *LRRK2* was proved to influence myelopoiesis and peripheral myeloid cell differentiation [19]. Effects of *LRRK2* on peripheral immune responses and its implication in PD pathogenesis are currently under intense investigation to analyse the interplay between central and peripheral immune responses and decipher the link between PD and peripheral inflammatory conditions like Crohn's

disease (CD), ulcerative colitis (UC), and leprosy [20, 21].

SNCA

The *SNCA* gene encodes alpha-synuclein, the main histopathological hallmark of PD. Several mutations and genetic rearrangements in *SNCA* have been linked to autosomal dominant PD [22, 23]. Moreover, the dosage of *SNCA* has been found closely associated with PD onset, clinical phenotype, and patient's survival [24]. Given that earlier onset and more rapid disease progression is observed in patients with *SNCA* gene triplications as compared to those with duplications, a dosage effect of the *SNCA* gene is expected. These observations indicate a neurotoxic effect of increased alpha-synuclein, even when point mutations are not present. The expression of alpha-synuclein in different immune cells, and the reported colocalization of neuroinflammatory spots at the sites of alpha-synuclein aggregation (Lewy neurites/Lewy bodies) indicate a potential implication of immune-mediated mechanisms regulated by alpha-synuclein in the pathogenesis of PD [25]. Alpha-synuclein has been proved to impact both the innate and adaptive immune responses in a way that favours neurodegeneration and PD development. Acting as a ligand of TLR-2 and TLR4 receptors or via metabolic reprogramming, alpha-synuclein is responsible for launching a proinflammatory response of microglia leading to neuronal demise probably through the release of neurotoxic mediators or phagocytosis of the neuronal cells in the region [26, 27]. Degenerated neurons release intracellularly aggregated alpha-synuclein to the extracellular space leading to further immune stimulation and neurodegeneration, a cascade that eventually leads to PD development. The neuroinflammation can be further aggravated by the microglial and astrocytes major histocompatibility complex class-II (MHC-II) presentation of alpha-synuclein leading to the recruitment and activation of other peripheral phagocytes (such as monocytes, macrophages, and lymphocytes) [28, 29]. Similar inflammatory responses mediated by alpha-synuclein were reported in the enteric nervous system, a finding that implies a potential role of gastrointestinal (GI) infections or inflammatory conditions in PD pathogenesis [30].

Moreover, alpha-synuclein has evident roles in regulating the magnitude and quality of adaptive immune responses in many aspects [31]. For instance, alpha-synuclein was found essential for the development

and activation of humoral immunity, since *SNCA* knock-out mouse models demonstrated impaired B cells development and IgG production [32, 33]. In addition, alpha-synuclein activity is also important for T cell development and differentiation [32]. Deficiency of alpha-synuclein was accompanied by elevated IL2 and Th1 response and decreased IL4 and Th2 differentiation, a finding that further emphasises the role of alpha-synuclein in T cells differentiation and priming [32, 34]. Furthermore, in support of the notion that neurodegeneration in PD might be induced by autoimmune responses, it has been observed that alpha-synuclein induces substantia nigra (SN) neurons to display major histocompatibility complex class-I (MHC-1) presenting immunogenic alpha-synuclein epitopes that are recognized by T cells [35, 36]. Such alpha-synuclein reactive-T cells have been linked to the development of PD and are now proposed as early diagnostic markers of the disease [37].

VPS35

Mutations in the vacuolar protein sorting 35 ortholog (*VPS35*) gene have been reported in about 1% of monogenic PD and 0.2% of sporadic PD [38]. The *VPS35* gene that has been linked to late-onset, autosomal dominant familial PD, encodes a core component of the retromer complex responsible for protein transmembrane sorting through the endosomes-trans-Golgi network pathway [38, 39]. *VPS35* is expressed in neuronal immune cells, namely microglia, and astrocytes. Recent studies have reported the role of *VPS35* in regulating innate system responses through modulating microglia and astrocytes activation [40, 41]. *VPS35* regulates microglial functions and polarisation through modulating the trafficking and recycling of immunomodulating receptors/mediators. Scientific literature exploring the effects of *VPS35* on microglia have been contradicting, probably due to variation in the brain anatomical sites and the inducer of neuroinflammation [41, 42]. One report indicated that *VPS35* favours the priming of microglial response into a pro-inflammatory rather than an anti-inflammatory response following an ischemic brain injury in the brain cortex [41]. On the contrary, *in vitro* and murine model studies proved that *VPS35* deficiency is associated with an enhanced microglial activation and excessive neuroinflammation through induction of microglial inflammatory mediators in the hippocampus [42, 43]. Furthermore, recent

245 studies suggest that VPS35 interacts with LRRK2. It
246 has been demonstrated that LRRK2 phosphorylates
247 some RAB proteins, which are involved in vesicle
248 trafficking, [44] and the D620N VPS35 mutation is
249 implied in the hyper activation LRRK2 kinase path-
250 way [45].

251 *PRKN, PINK1, DJ1*

252 The Parkin RBR E3 ubiquitin-protein ligase
253 (*PRKN*), PTEN-induced kinase 1 (*PINK1*), and *DJ1*
254 (also known as PARK7) genes are widely known to
255 contribute to autosomal recessive PD and encode pro-
256 teins that are essential regulators of mitochondrial
257 homeostasis and quality control [46]. *PINK1* and
258 *PARKIN* control mitochondrial turnover via a selec-
259 tive autophagy process known as mitophagy [47].
260 They are also involved in regulating mitochondrial
261 fusion and fission, local repair of impaired mito-
262 chondria, and the genesis of new mitochondria [48].
263 *DJ1* works interactively with *PARKIN* and *PINK1*
264 in the mitochondrial quality control through regulat-
265 ing oxidative stress caused by ROS [49]. The three
266 genes, which are known for their neuroprotective
267 roles, are linked to recessive PD with their mutations
268 reported in 13% of early-onset PD. The similarity
269 in disease phenotype and clinicopathological fea-
270 tures suggests similar pathogenic pathways through
271 which the three genes culminate in PD development.
272 Among the reported pathogenic mechanisms through
273 which *PRKN*, *PINK1*, and *DJ-1* genes contribute
274 to PD is neuroinflammation. Neuroinflammation
275 caused by loss-of-function mutations of these pro-
276 teins can be mitochondrial-mediated, e.g., impaired
277 mitophagy caused by *PARKIN/PINK1* deficiency
278 results in the release of mitochondrial DNA and
279 other reactive species that can stimulate innate im-
280 munity and trigger inflammation [50, 51]. Furthermore,
281 *PRKN/PINK1* loss of function mutations impair the
282 proteins inhibitory effect on mitochondrial antigen
283 presentation through the alternative mitochondrial-
284 derived vesicles (MDV) pathway leading to excessive
285 mitochondrial antigen presentation and stimulation
286 of immune responses according to studies conducted
287 in *in vitro* and *in vivo* models [52]. The unleashed
288 mitochondrial antigen presentation through the MDV
289 pathway caused by the loss of *PRKN/PINK1* regu-
290 latory effect further supports the autoimmune basis
291 of PD. It also supports the role of the brain/gut
292 axis in PD pathogenesis, considering that *PINK1*-
293 *-/-* mouse models have demonstrated a propagated
294 inflammatory response after bacterial infection with

295 concomitant development of peripheral and neural
296 cytotoxic mitochondrial-specific CD8 + T cells. The
297 consequent decline in dopaminergic neuron density
298 and development of motor impairment in the mice
299 indicate that the *PINK1*-/- associated autoimmune
300 response likely contributes to dopaminergic neu-
301 rodegeneration and PD development [52]. Moreover,
302 *PRKN/PINK1* deficiency can lead to an exacerbated
303 inflammatory response through inhibition on the
304 NLRP3 inflammasome signalling [53, 54]. Similar
305 to *PRKN/PINK1* mutations, *DJ-1* dysfunction was
306 observed in CD4 + T cells to enhance neuroinflam-
307 mation due to the loss of *DJ-1* antioxidant effect and
308 regulatory ROS mediated inflammation [55].

309 Furthermore, these proteins can also induce neu-
310 roinflammation through other mitochondrial-ind-
311 ependent mechanisms, e.g., *PINK1* deficiency was
312 reported to promote innate immune responses
313 through nitric oxide production by glia/astrocytes
314 cells leading to inflammation-induced neuronal death
315 [56]. Additionally, *DJ-1* exerts anti-inflammatory
316 effects independently from the ROS regulatory
317 pathways, e.g., through inducing the synthesis of
318 anti-inflammatory mediators like prostaglandin D2,
319 stimulating the migration of CD3 + T cell, and
320 CD4 + T cells differentiation [57, 58].

321 *GBA*

322 Mutations of the glucocerebrosidase gene (*GBA*),
323 encoding for the lysosomal enzyme β -gluco-
324 cerebrosidase, are the most prominent genetic risk
325 factor recognized for PD. According to the records,
326 *GBA* mutations, which are detected in 8–12% of
327 total sporadic PD in the world, increase the risk of
328 developing PD by 5–10 folds in a carrier compared
329 to the general population [59]. *GBA* is expressed
330 in immune cells including monocytes/macrophages
331 and lymphocytes, and it has been associated with an
332 aberrant inflammatory response mediated by mono-
333 cytes/macrophages and B cells [60, 61]. In addition,
334 the glucocerebrosidase enzyme activity is generally
335 reduced in monocytes from PD patients compared to
336 control cases according to published reports. Such
337 reduction was found correlated with the disease's
338 clinical characteristics, which highlights the potential
339 of peripheral monocytes glucocerebrosidase activity
340 as an early marker of PD diagnosis [62, 63].

341 Moreover, *GBA* mutations demonstrated associa-
342 tion with marked astrocytes and microglial activation
343 indicating that *GBA* mutations are implicated in
344 launching and/or aggravating neuroinflammatory

345 responses [64, 65]. Such effect is detected early
346 enough before neurodegeneration, a time where ther-
347 apeutic intervention can be maximally beneficial.
348 Hence, investigating the mechanisms and mediators
349 through which *GBA* mutations initiate and propa-
350 gate neuroinflammation is now a hot area of research
351 [66]. Despite the mentioned association of *GBA*
352 mutations with central and peripheral inflammatory
353 responses, initial investigations of central and periph-
354 eral inflammatory cytokines as early markers of PD
355 have returned with negative outcomes [67].

356 COMMON GENETIC RISK FACTORS 357 CONTRIBUTING TO THE IMMUNE 358 SYSTEM RESPONSE IN PARKINSON'S 359 DISEASE

360 Since the development of advanced genotyping,
361 sequencing technologies and complex analysis tools,
362 large-scale genetic studies have expanded rapidly and
363 have provided opportunities for a mechanistic elu-
364 cidation of PD etiology. Since 2009, genome-wide
365 association studies (GWAS) and meta-analyses have
366 shown evidence for an implication of genetic con-
367 tributors conferring moderate and low risk to PD
368 susceptibility. The largest and latest GWAS meta-
369 analyses conducted in Europeans, Asians, and Latino
370 populations, have nominated a total of 92 loci pre-
371 disposing to PD as well as several other suggestive
372 genomic regions linked to age at disease onset and
373 progression that warrant further study [68, 69]. PD
374 risk loci such as *LRRK2*, *MAPT*, *BST1*, and *HLA*
375 among others have been recognized as key players to
376 the immune-mediated response involved in PD devel-
377 opment (Table 1) [21], strengthening the hypothesis
378 that common variation plays a crucial role in disease
379 etiology. Furthermore, expression and methylation
380 quantitative trait loci analyses have further nominated
381 possible functional mechanisms by which genetic
382 variants may be contributing to the immune system
383 regulation [21].

384 While GWAS meta-analyses have identified
385 increasing numbers of novel genetic risk loci in myr-
386 iad datasets, recent research in the PD genetics field
387 has focused on understanding how genetic risk vari-
388 ants may disrupt biological processes and drive the
389 underlying pathobiology of the disease. In this con-
390 text, an exciting era for PD research has arisen with
391 large-scale omics analyses supporting the role of the
392 immune response in the pathophysiology of PD [21,
393 70, 71]. Based on an unbiased approach applied to

394 large genetic and genomics datasets available to date,
395 pathway-specific polygenic risk scores and transcrip-
396 tomics analyses have identified that a cumulative
397 effect of common genetic risk variants contribute to
398 the innate and adaptive immune responses, as critical
399 pathways in PD etiology [70]. In concordance with
400 this study, heritability and gene-set enrichment meth-
401 ods aimed at exploring particular functional marks for
402 regulatory activity and gene-set lists have supported
403 the implication of the adaptive and innate immune
404 system in PD etiology and an enrichment for sporadic
405 PD genetic heritability [72].

406 Interestingly, a more recent study has shown a
407 significant enrichment of PD risk heritability in
408 microglia and monocytes [73]. As an effort to under-
409 stand the contribution of the immune system in PD
410 pathogenesis, the authors showed that the microglial
411 signature gene, *P2RY12*, located near a PD GWAS
412 signal colocalizes to a microglia open chromatin
413 region and enhancer region [73]. *P2RY12* has been
414 found to be downregulated in AD brain sections
415 of microglia as a marker of inflammation, hence
416 supporting a possible similar role in PD as a tar-
417 getable microglial gene candidate and pathogenic
418 player [74]. Additionally, a recent genetic analysis
419 of the human microglial transcriptome across brain
420 regions, aging and disease pathologies has nominated
421 microglia-specific enhancers, finding associations
422 with microglial expression of *USP6NL* for AD and
423 *P2RY12* for PD [75].

424 Furthermore, much attention has been paid to
425 explore the role of immune cells in PD pathogene-
426 sis from a genetics perspective. Interestingly, recent
427 studies showed that genes within PD GWAS loci are
428 specifically expressed in T-cells [72]. Of note, GWAS
429 have nominated a transcription factor named *SATB1*,
430 which is associated with T-cell function and the estab-
431 lishment of immune tolerance [76, 77]. Specifically,
432 the number of infiltrated CD8 positive T-cells is ele-
433 vated in the PD substantia nigra pars compacta which
434 correlates to neuronal cell loss. In addition, half of
435 CD8 positive T-cells express an immune activation
436 marker named IFN γ [78].

437 Moreover, PD-associated variants, such as
438 rs76904798 which regulates the expression of
439 *LRRK2*, had been shown to colocalize with periph-
440 eral monocyte expression quantitative trait loci
441 (eQTL) [71]. Recent publications supported the
442 notion that *LRRK2* levels were elevated in PD
443 patient monocytes which tended to secrete more
444 inflammatory factors and cytokines than healthy
445 subjects [12]. Although *LRRK2* plays a regulatory

Table 1
The implication of PD risk loci in the immune system regulation

Chr	Nearest gene	Immune system implication	Reference
1	<i>DJ-1</i>	By regulating oxidative stress caused by ROS, DJ1 interacts with PARKIN and PINK1 in the mitochondrial quality control pathway. As a result of DJ-1 loss-of-function, ROS are released, which can activate innate immunity and trigger inflammation.	PMID: 22403686, PMID: 19276172
1	<i>FCGR2A</i>	Highly expressed in microglia and macrophages. Prominent role in phagocytosis and debris cleaning (microglia-specific enhancer)	PMID: 34617105
1	<i>GBA</i>	Several immune cells express GBA, including monocytes, macrophages, and lymphocytes. GBA deficiency has several immunological effects, such as multi-system inflammation, B-lymphocyte hyperproliferation, increased levels of proinflammatory cytokines, microglial activation, and astrogliosis.	PMID: 33935104, PMID: 26376862
1	<i>PINK1</i>	An analysis of gene expression profiles in PINK1-deficient mice indicated that the loss of PINK1 function modified the expression of immunomodulatory genes in the striatum.	PMID: 21249202
2	<i>IL-1β</i>	Proinflammatory cytokine that increases the degeneration of dopamine neurons in animal models of PD. <i>IL-1β</i> has been also identified as a mediator of microglia activation.	PMID: 18304357
2	<i>IL1R2</i>	The <i>IL1R2</i> gene is implicated in neurodegenerative diseases. IL-1R2 expression is suppressed by pro-inflammatory agents, such as LPS and IFN- γ .	PMID: 23195532
2	<i>STK39</i>	The association of <i>STK39</i> variants with PD points to a potential of this gene on inflammation and oxidative stress. These functional consequences should be further investigated.	PMID: 26469904
3	<i>SATB1</i>	SATB1, a T-lymphocyte-enriched transcription factor and chromatin organiser, is crucial to the regulation of many genes involved in the T-lymphocyte development and activation.	PMID: 10716941, PMID: 17057718
3	<i>TLR9</i>	TLR9 is part of the toll-like receptor family which activates an inflammatory cascade by recognizing mitochondrial DNA as an endogenous danger-associated molecular pattern. The TLR9-mediated inflammatory response has been reported to increase the expression of pro-inflammatory cytokines, such as IL-1 β .	PMID: 20347818
4	<i>BST1</i>	The <i>BST1</i> gene encodes for the leukocyte surface protein CD157, which is highly expressed in bone marrow cells of patients with rheumatoid arthritis, suggesting that it may promote the growth of pre-B lymphocytes.	PMID: 12415565
4	<i>SCARB2/LIMP2</i>	<i>SCARB2</i> , which is highly expressed in plasmacytoid dendritic cells, plays a role in mediating GBA trafficking from endoplasmic reticulum to lysosome. <i>SCARB2</i> expression regulates the production of IFN and the TLR9-mediated activation of IFN regulatory factor 7 (IRF7).	PMID: 24485911; PMID: 25862818
4	<i>SNCA</i>	<i>SNCA</i> encodes alpha-synuclein which acts as a ligand for toll-like receptor 2 (TLR2) on microglia, linking it with the innate immune system. Moreover, TLR2 has been identified as being present on T lymphocytes, B lymphocytes, monocytes, and macrophages. alpha-synuclein epitopes can be presented on MHC molecules and can activate both helper and cytotoxic T-cells.	PMID: 23463005, PMID: 27358579
6	<i>ERβ</i>	Estrogen has been suggested to have a neuroprotective role in PD, and regulate neuroinflammatory genes, including <i>IL-6</i> .	PMID: 11102464, PMID: 15635591

6	<i>HLA-DQA1</i>	HLA-DQA, is considered to be a key immune system player, and it is found to be increased in PD patients.	PMID: 27148593
6	<i>HLA-DRB6</i>	HLA-DRB6, a component of the major histocompatibility complex, is decreased in PD patients. The change of <i>HLA</i> alleles influences the expression of MHC which leads to the abnormal T cell immunity in PD patients.	PMID: 28892059; PMID: 34548497
6	<i>PARK2</i>	Parkin knockout mice show mitochondrial dysfunction and oxidative stress, which induces inflammatory factors production. Parkin deficient individuals show decreased lymphocyte mitochondrial complex I activity, providing additional evidence that loss of Parkin function causes mitochondrial dysfunction.	PMID: 27345367, PMID: 29665074
6	<i>TNF</i>		PMID: 29710331; PMID: 29710331
7	<i>GPNMB</i>	GPNMB is found to be highly expressed in microglia. Inhibition of GPNMB by siRNA significantly reduces the expression of TNF- α , IL-1 β and inducible nitric oxide synthase (iNOS) in activated mouse BV2 cells, suggesting that GPNMB is involved in microglia activation and the production of pro-inflammatory cytokines.	PMID: 24682924
7	<i>IL-6</i>	<i>IL-6</i> is a multifunctional cytokine involved in immune response and inflammation and plays a crucial role in the central nervous system. Previous reports showed that <i>IL-6</i> is induced by alpha-synuclein and PD-causing mutants in astrocytes and microglia cells.	PMID: 17012252
8	<i>PDLIM2</i>	PDLIM2 has been shown to inhibit the development of T-helper 17 (TH17) cells through the activator of transcription 3 (STAT3) pathway, known to have a pathogenic role in inflammatory disease.	PMID: 22155789
9	<i>SH3GL2</i>	SH3GL2 plays a role in endocytosis and it has been hypothesized that this protein acts downstream of LRRK2 to induce synaptic autophagosome formation and may be deregulated in PD.	PMID: 28282269
12	<i>IFN-γ</i>		PMID: 17376993; PMID: 21472005
12	<i>LRRK2</i>	The rs76904798 variant, which regulates the expression of LRRK2, colocalizes with a peripheral monocyte eQTL. Accordingly, LRRK2 is strongly expressed in B-lymphocytes, T-lymphocytes, and monocytes from PD patients, and it is positively related to cytokine production in T-lymphocytes.	PMID: 30824768
13	<i>MBNL2</i>	The function of <i>MBNL2</i> is related to postnatal splicing patterns in brains. A previous study showed that knockout <i>MBNL2</i> related to the proinflammatory process by increasing microglia expression in specific brain regions, such as medial prefrontal cortex and hippocampus.	PMID: 30060068; PMID: 34617105
17	<i>MAPT</i>	<i>MAPT</i> , which encodes the tau protein, has been shown to induce morphological transformation of microglia which results in the generation of NO, and proinflammatory cytokines (IL-1 β , IL-6, TNF- α).	PMID: 21813771
17	<i>ATP6V0A1</i>	Microglia and their precursors express the <i>ATP6V0A1</i> gene, involved in the acidification of intracellular compartments and in phagosomal fusion, which is essential for phagocytosis.	PMID: 18510934
20	<i>DDRGK1</i>	DDRGK1 depletion inhibits the expression of NF- κ B target genes, suggesting that DDRGK1 is involved in the regulation of the NF- κ B pathway through interaction with I κ B α .	PMID: 23675531
21	<i>DYRK1A</i>	Key role in the phosphorylation of several immune response mediators (neuron-specific enhancer)	PMID: 34617105

446 role in immune cells and PD, the mechanism of
 447 LRRK2 regulating PD patient monocyte or T-cell
 448 gene expression has not been yet unravelled.

449 Besides immune responses nominated through
 450 polygenic risk of biological processes, additional
 451 pathways have also been shown to induce immune
 452 activation in neurodegenerative disease through com-
 453 mon genetic variation, such as the alpha-synuclein
 454 pathway and the MAPK signalling pathway [70].
 455 Previous studies demonstrated that microglia acti-
 456 vation was induced by aggregated alpha-synuclein,
 457 and this phenomenon raised dopaminergic neurotox-
 458 icity [79]. An additional supportive study has shown
 459 that alpha-synuclein induced the inflammatory factor
 460 interleukin-6 (IL-6) through MAPK MEK1/2, JNK
 461 and p38 MAPK in human astrocytes [80]. There-
 462 fore, genetic variation in the *SNCA* locus may also
 463 be responsible for inducing innate and adaptive immu-
 464 nity [81, 82]. MAPKs are a group of serine/threonine
 465 protein kinases participating in the regulation of
 466 inflammatory mediator production, stress response
 467 and maintenance of the immune system, which con-
 468 tribute to PD pathology [83, 84]. In addition, MAPKs
 469 contribute to T-cell differentiation and maturation, as
 470 well as immune cell activation [85, 86].

471 The majority of nominated risk variants are located
 472 in intronic and intergenic regions, regulating gene
 473 expression. For example, rs1990622, a genetic vari-
 474 ant thought to affect cognition in PD, has the ability
 475 to enhance long-range chromatin looping by promot-
 476 ing the interaction between the chromatin organising
 477 protein CCCTC-binding factor (CTCF) downstream
 478 of *TMEM106B* and distal regulatory elements that
 479 in turn act increasing the expression of *TMEM106B*
 480 [87, 88]. *TMEM106B* has been found to modulate
 481 inflammation in the central nervous system (CNS)

482 of post-mortem aged brains [89, 90]. Whole-genome
 483 co-expression network analysis nominated five gene
 484 clusters derived from frontal cortex data analyses
 485 conducted in elder individuals and categorised CNS
 486 cell types where these clusters were more differen-
 487 tially expressed, including microglia, astroglia and
 488 other neuron-associated groups. Interestingly, the
 489 rs1990622 genotype showed a significant elevation
 490 in expression with increased risk allele load on the
 491 microglia-associated genes cluster [89]. Further anal-
 492 yses confirmed the relationship between microglia
 493 gene set expression levels and the rs1990622 risk
 494 variant [89]. In this study, authors demonstrated that
 495 *TMEM106B* modulates the polarisation of immune
 496 cells towards pro-inflammatory stage of gene expres-
 497 sion signature in the risk allele (Thr185) compared
 498 with the protective allele (Ser185) of rs1990622 in
 499 human monocyte-derived dendritic cells [89]. This
 500 suggests that PD-associated SNPs participating in
 501 immune response or immune-associated genes might
 502 interact with CTCF binding sites, regulating gene
 503 expression in harmful neuronal cells, and increas-
 504 ing the risk of neurodegeneration. However, further
 505 verification and discussion are needed in PD studies.

506 Heritable alterations contributing to regulatory
 507 gene expression such as DNA methylation and
 508 histone modifications, which alter chromatin acces-
 509 sibility and gene regulation have been linked to
 510 PD etiology. Large-scale analysis also indicated that
 511 chromatin remodelling and organisation which is
 512 highly integrated in maintaining and controlling chro-
 513 matin landscape and genome stability [70] play a
 514 crucial role in PD pathogenesis. A recent study
 515 revealed that *BIN1*, *SORL1* and *MEF2C* show differ-
 516 entially methylated enhancers in the prefrontal cortex
 517 of Alzheimer's patients [91]. In addition, evidence

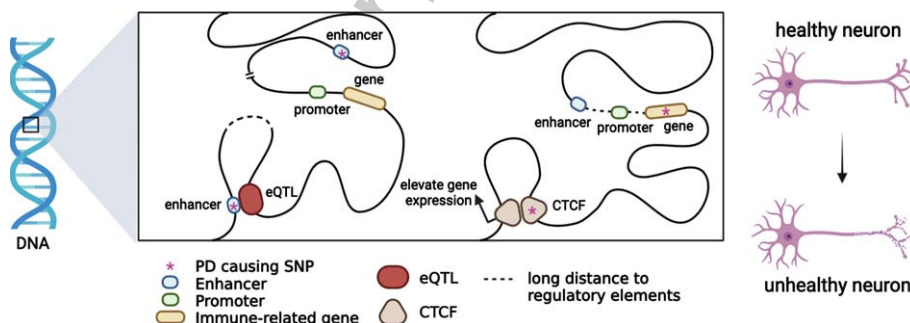


Fig. 1. Putative functional mechanisms for which PD genetic variants might contribute to the immune system regulation. Most PD risk loci span non-coding regions. When a PD risk SNP with an eQTL mechanism colocalizes with immune-related genes in topologically associating domains, immune-gene regulation will be affected by chromatin interaction. PD causing SNPs may be located in enhancers or promoters, and immune-gene regulation will be affected by trans-acting or cis-acting regulatory mechanisms.

518 supports that a portion of genetic variants contributes
519 to active and repressive chromatin states, such as
520 heterochromatin and euchromatin [92]. Epigenetic
521 regulation contributing to the immune system reg-
522 ulation affords an opportunity to develop potential
523 therapeutic strategies in PD as neuroprotective targets
524 in disease onset and progression (Fig. 1).

525 GENETIC COMORBIDITIES BETWEEN 526 PARKINSON'S DISEASE AND 527 AUTOIMMUNE DISEASES

528 Several genetic factors have been found to be
529 common between PD and autoimmune disorders.
530 For instance, genome-wide analysis identified 17
531 shared loci adjacent to *GAK*, *HLA-DRB5*, *LRRK2*,
532 and *MAPT* genes for PD and 7 autoimmune diseases,
533 including rheumatoid arthritis, UCs, and CD, sug-
534 gesting that moderate and low risk loci contribute to
535 pleiotropy among these diseases [21].

536 The Human leukocyte antigen (*HLA*), also known
537 as major histocompatibility complex (MHC) is a fam-
538 ily of genes encoding membrane proteins responsible
539 for antigen presentation to T cells, hence play-
540 ing a crucial role in the adaptive immune response
541 [93] is the most reported genetic risk factor asso-
542 ciated with autoimmune diseases and has been
543 widely linked to several neurological conditions [94].
544 The *HLA* region was first identified to be associ-
545 ated with late onset PD in 2010 and commonly
546 referred to as the *PARK18* locus [95]. More recently,
547 fine-mapping analyses have nominated four *HLA*
548 types as associated with PD: *HLA-DQA1*03:01*,
549 *HLA-DQB1*03:02*, *HLA-DRB1*04:01*, and *HLA-*
550 *DRB1*04:04* [96]. Furthermore, identification of new
551 SNPs raised the possibility of PD having an asso-
552 ciation with regulatory elements effecting the *HLA*
553 class II gene expression and classical *HLA* class II
554 allele [97]. Since then, different approaches have
555 been undertaken to further dissect the role of *HLA*
556 in PD and autoimmune conditions.

557 Genetic characterization of the *HLA-DRB1* locus
558 have revealed shared genetic risk factors associated
559 with rheumatoid arthritis (RA) and PD. Such asso-
560 ciation remains controversial since the directionality
561 of effect for both diseases is sometimes inconsistent,
562 suggesting either a predisposing (*HLA-DRB1*01:01*
563 allele) [98] or protective effect (*HLA-DRB1*04*
564 allele) [98, 99]. Therefore, there is an ongoing debate
565 about the direction of the association between PD
566 and RA [100]. In an attempt to further clarify such

567 an association, research aimed at exploring genetic
568 correlations between these diseases has been con-
569 ducted in European and non-European populations.
570 In a Taiwanese case-study, PD incidence was observ-
571 ably higher in RA patients than in controls [101] and
572 a significant genetic correlation between PD and RA
573 has been reported [102]. More recently, a Mendelian
574 randomization study showed a genome-wide nega-
575 tive correlation, suggesting that RA has a protective
576 effect on PD [103], a conclusion supported by other
577 epidemiological case studies [104].

578 Likewise, the etiology of CD, an inflammatory
579 bowel disorder, is thought to be partly explained
580 by the contribution of variants in the *LRRK2* gene,
581 which is considered among the main genetic con-
582 tributors to PD risk [105]. Exome sequencing and
583 genotyping of CD cases and controls has recently
584 suggested that the N551K variant in *LRRK2* pro-
585 vides protection from CD, while the N2081D variant
586 confers risk, reinforcing the notion that both protec-
587 tive and harmful influences contribute to the complex
588 genetic architecture of immune contributors [20].
589 Additionally, one of the first identified shared poly-
590 morphisms between CD and PD was M2397T [106]
591 which presumably enhances interferon- γ responses
592 in monocytes thus modifying the immune response
593 [107]. Besides *LRRK2*, other genes have also been
594 studied in CD/PD. The Solute Carrier Family 39
595 Member 8 (*SLC39A8*) gene encodes for a membrane
596 manganese transporter protein, and the missense vari-
597 ant A391T has been previously linked to both CD
598 [108] and PD [109].

599 Additional candidate shared genes between PD and
600 CD have recently come to attention. There are hints
601 that both alpha-synuclein and tau proteins, encoded
602 by the *SNCA* and *MAPT* genes respectively, and con-
603 sidered pathological hallmarks in PD, may also be
604 involved in CD: increased expression levels of alpha-
605 synuclein protein were detected in inflamed tissues
606 of CD patients [110] and tau protein expression was
607 found upregulated in the enteric nervous system in
608 CD patients [111]. However, these studies were car-
609 ried out at a protein level and do not provide enough
610 evidence to assess genetic comorbidity.

611 Epigenetic changes have also been investigated
612 between PD and autoimmune diseases. Estimation
613 of Pearson's correlation coefficient between PD and
614 RA revealed that both traits share 337 gene pairs
615 with co-methylation changes [112]. Also, a signifi-
616 cant overlap in epigenetic patterns of risk genes for
617 PD, RA, and CD was found in butyrate-associated
618 methylation sites [113]. Investigating PD from an

619 epigenetic angle may clarify the role of immune
620 response and immune-mediated mechanisms.

621 FUTURE DIRECTIONS

622 Increasing evidence from recent literature sug-
623 gests that PD presents genetically as more of a
624 systemic disorder in which inflammation might play
625 a causative role rather than being a consequence
626 or an epiphenomenon of the neurodegenerative pro-
627 cess. As discussed, this notion is reinforced by the
628 overlapping genetic pleiotropy that seems to exist
629 between PD and several autoimmune diseases. Addi-
630 tionally, genes implicated in monogenic forms of PD
631 and genome-wide loci linked to idiopathic PD have
632 been seen to be widely implicated in immune sys-
633 tem related processes including antigen presentation,
634 inflammation regulation and the complement system.

635 Despite efforts at dissecting genetic drivers and
636 modifiers of PD etiology that modulate the immune
637 response, gene-environment interactions merit fur-
638 ther investigation. Neuroinflammation is triggered
639 by immunological insults likely at the convergence
640 of genetic and environmental factors for which the
641 exact mechanisms of such association remain to be
642 elucidated. The challenge ahead lies in our ability
643 to further dissect immune targets by defining the
644 complex genetic architecture of the immune system
645 contributors to disease risk, onset and progression, as
646 well as its functional implications, and the interplay
647 that exists with the environment.

648 The field of immunogenetics is rapidly expand-
649 ing but there are still caveats to be addressed. Most
650 of our current knowledge on the role of the immune
651 system in PD etiology comes from human studies
652 conducted in European populations. As we move
653 forward, shedding light on immunogenetic contribu-
654 tors in Non-European populations and its interaction
655 to specific environmental factors is essential to
656 provide generalised insights into disease pathophys-
657 iology. Looking to the future of immunogenetics in
658 PD, multimodal data integration with harmonised
659 large-scale, clinica-longitudinal data and collabora-
660 tive worldwide initiatives will facilitate translation of
661 immunogenetic factors to specific functional mecha-
662 nisms. Although much work needs to be done from
663 the genetics and genomics perspective to inform biol-
664 ogy, future immune-based therapeutic approaches
665 aimed at modifying the expression of PD risk loci in
the right patients and at the right time may succeed.

ACKNOWLEDGMENTS

666 This work was carried out with the support and
667 guidance of the 'GP2 Trainee Network' which is
668 part of the Global Parkinson's Genetics Program and
669 funded by the Aligning Science Across Parkinson's
670 (ASAP) initiative. 671

672 This research was supported, in part, by the Intra-
673 mural Research Program of the National Institutes of
674 Health (National Institute on Aging, National Insti-
675 tute of Neurological Disorders and Stroke; project
676 numbers 1ZIA-NS003154, Z01-AG000949-02 and
677 Z01-ES10198).

CONFLICT OF INTEREST

678 The authors have no conflict of interest to report. 679

REFERENCES

- 680
- 681 [1] Tansey MG, Goldberg MS (2010) Neuroinflammation in
682 Parkinson's disease: Its role in neuronal death and impli-
683 cations for therapeutic intervention. *Neurobiol Dis* **37**,
684 510-518.
 - 685 [2] Reed X, Bandrés-Ciga S, Blauwendraat C, Cookson MR
686 (2019) The role of monogenic genes in idiopathic Parkin-
687 son's disease. *Neurobiol Dis* **124**, 230-239.
 - 688 [3] Bandrés-Ciga S, Diez-Fairen M, Kim JJ, Singleton AB
689 (2020) Genetics of Parkinson's disease: An introspection
690 of its journey towards precision medicine. *Neurobiol Dis*
691 **137**, 104782.
 - 692 [4] Pierce S, Coetzee GA (2017) Parkinson's disease-
693 associated genetic variation is linked to quantitative
694 expression of inflammatory genes. *PLoS One* **12**,
695 e0175882.
 - 696 [5] Nalls MA, Blauwendraat C, Vallerga CL, Heilbron K,
697 Bandrés-Ciga S, Chang D, Tan M, Kia DA, Noyce AJ,
698 Xue A, Bras J, Young E, von Coelln R, Simón-Sánchez J,
699 Schulte C, Sharma M, Krohn L, Pihlström L, Siitonen A,
700 Iwaki H, Leonard H, Faghri F, Gibbs JR, Hernandez DG,
701 Scholz SW, Botia JA, Martinez M, Corvol J-C, Lesage S,
702 Jankovic J, Shulman LM, Sutherland M, Tienari P, Maja-
703 ma K, Toft M, Andreassen OA, Bangale T, Brice A, Yang
704 J, Gan-Or Z, Gasser T, Heutink P, Shulman JM, Wood
705 NW, Hinds DA, Hardy JA, Morris HR, Gratten J, Visscher
706 PM, Graham RR, Singleton AB, 23andMe Research Team,
707 System Genomics of Parkinson's Disease Consortium,
708 International Parkinson's Disease Genomics Consortium
709 (2019) Identification of novel risk loci, causal insights,
710 and heritable risk for Parkinson's disease: A meta-analysis
711 of genome-wide association studies. *Lancet Neurol* **18**,
712 1091-1102.
 - 713 [6] Tan E-K, Chao Y-X, West A, Chan L-L, Poewe W,
714 Jankovic J (2020) Parkinson disease and the immune sys-
715 tem - associations, mechanisms and therapeutics. *Nat Rev*
716 *Neurol* **16**, 303-318.
 - 717 [7] Dächsel JC, Farrer MJ (2010) LRRK2 and Parkinson dis-
718 ease. *Arch Neurol* **67**, 542-547.
 - 719 [8] Lesage S, Houot M, Mangone G, Tesson C, Bertrand H,
720 Forlani S, Anheim M, Brefel-Courbon C, Broussolle E,

- 721 Thobois S, Damier P, Durif F, Roze E, Tison F, Grabli D,
722 Ory-Magne F, Degos B, Viallet F, Cormier-Dequaire F,
723 Ouvrard-Hernandez A-M, Vidailhet M, Lohmann E, Sing-
724 leton A, Corvol J-C, Brice A, French Parkinson disease
725 Genetics Study Group (PDG) (2020) Genetic and pheno-
726 typic basis of autosomal dominant Parkinson's disease in
727 a large multi-center cohort. *Front Neurol* **11**, 682.
- [9] 728 Marder K, Wang Y, Alcalay RN, Mejia-Santana H, Tang
729 M-X, Lee A, Raymond D, Mirelman A, Saunders-Pullman
730 R, Clark L, Ozelius L, Orr-Urtreger A, Giladi N, Bressman
731 S, LRRK2 Ashkenazi Jewish Consortium (2015) Age-
732 specific penetrance of LRRK2 G2019S in the Michael J.
733 Fox Ashkenazi Jewish LRRK2 Consortium. *Neurology* **85**,
734 89-95.
- [10] 735 Lee AJ, Wang Y, Alcalay RN, Mejia-Santana H, Saunders-
736 Pullman R, Bressman S, Corvol J-C, Brice A, Lesage S,
737 Mangone G, Tolosa E, Pont-Sunyer C, Vilas D, Schüle B,
738 Kausar F, Foroud T, Berg D, Brockmann K, Goldwurm S,
739 Siri C, Asselta R, Ruiz-Martinez J, Mondragón E, Marras
740 C, Ghate T, Giladi N, Mirelman A, Marder K, Michael
741 J. Fox LRRK2 Cohort Consortium (2017) Penetrance
742 estimate of LRRK2 p.G2019S mutation in individu-
743 als of non-Ashkenazi Jewish ancestry. *Mov Disord* **32**,
744 1432-1438.
- [11] 745 Rideout HJ, Re DB (2017) LRRK2 and the "LRRK-
746 tosome" at the crossroads of programmed cell death:
747 Clues from RIP kinase relatives. *Adv Neurobiol* **14**,
748 193-208.
- [12] 749 Cook DA, Kannarkat GT, Cintron AF, Butkovich LM,
750 Fraser KB, Chang J, Grigoryan N, Factor SA, West AB,
751 Boss JM, Tansey MG (2017) LRRK2 levels in immune
752 cells are increased in Parkinson's disease. *NPJ Parkinsons*
753 *Dis* **3**, 11.
- [13] 754 Li T, Ning B, Kong L, Dai B, He X, Thomas JM, Sawa
755 A, Ross CA, Smith WW (2021) A LRRK2 GTP binding
756 inhibitor, 68, reduces LPS-induced signaling events and
757 TNF- α release in human lymphoblasts. *Cells* **10**, 480.
- [14] 758 Kim KS, Marcogliese PC, Yang J, Callaghan SM, Resende
759 V, Abdel-Messih E, Marras C, Visanji NP, Huang J,
760 Schlossmacher MG, Trinkle-Mulcahy L, Slack RS, Lang
761 AE, Canadian Lrrk2 in Inflammation Team (CLINT), Park
762 DS (2018) Regulation of myeloid cell phagocytosis by
763 LRRK2 via WAVE2 complex stabilization is altered in
764 Parkinson's disease. *Proc Natl Acad Sci U S A* **115**, E5164-
765 E5173.
- [15] 766 Levy DR, Udgate A, Tourlomousis P, Symmons MF,
767 Hopkins LJ, Bryant CE, Gay NJ (2020) The Parkin-
768 son's disease-associated kinase LRRK2 regulates genes
769 required for cell adhesion, polarization, and chemotaxis
770 in activated murine macrophages. *J Biol Chem* **295**, 10857-
771 10867.
- [16] 772 Nazish I, Arber C, Piers TM, Warner TT, Hardy JA, Lewis
773 PA, Pocock JM, Bandopadhyay R (2021) Abrogation of
774 LRRK2 dependent Rab10 phosphorylation with TLR4
775 activation and alterations in evoked cytokine release in
776 immune cells. *Neurochem Int* **147**, 105070.
- [17] 777 Atashrazm F, Hammond D, Perera G, Bolliger MF, Matar
778 E, Halliday GM, Schüle B, Lewis SJG, Nichols RJ,
779 Dzamko N (2019) LRRK2-mediated Rab10 phosphory-
780 lation in immune cells from Parkinson's disease patients.
781 *Mov Disord* **34**, 406-415.
- [18] 782 Kozina E, Sadasivan S, Jiao Y, Dou Y, Ma Z, Tan H, Kodali
783 K, Shaw T, Peng J, Smeyne RJ (2018) Mutant LRRK2
784 mediates peripheral and central immune responses leading
785 to neurodegeneration *in vivo*. *Brain* **141**, 1753-1769.
- [19] 786 Park J, Lee J-W, Cooper SC, Broxmeyer HE, Cannon
787 JR, Kim CH (2017) Parkinson disease-associated trans-
788 gene disrupts marrow myelopoiesis and peripheral Th17
789 response. *J Leukoc Biol* **102**, 1093-1102.
- [20] 790 Hui KY, Fernandez-Hernandez H, Hu J, Schaffner A,
791 Pankratz N, Hsu N-Y, Chuang L-S, Carmi S, Villaverde
792 N, Li X, Rivas M, Levine AP, Bao X, Labrias PR, Har-
793 itunians T, Ruane D, Gettler K, Chen E, Li D, Schiff ER,
794 Pontikos N, Barzilai N, Brant SR, Bressman S, Cheifetz
795 AS, Clark LN, Daly MJ, Desnick RJ, Duerr RH, Katz S,
796 Lencz T, Myers RH, Ostrer H, Ozelius L, Payami H, Peter
797 Y, Rioux JD, Segal AW, Scott WK, Silverberg MS, Vance
798 JM, Ubarretxena-Belandia I, Foroud T, Atzmon G, Pe'er
799 I, Ioannou Y, McGovern DPB, Yue Z, Schadt EE, Cho
800 JH, Peter I (2018) Functional variants in the gene confer
801 shared effects on risk for Crohn's disease and Parkinson's
802 disease. *Sci Transl Med* **10**, eaai7795.
- [21] 803 Witoelar A, Jansen IE, Wang Y, Desikan RS, Gibbs
804 JR, Blauwendraat C, Thompson WK, Hernandez DG,
805 Djurovic S, Schork AJ, Bettella F, Ellinghaus D, Franke
806 A, Lie BA, McEvoy LK, Karlsten TH, Lesage S, Morris
807 HR, Brice A, Wood NW, Heutink P, Hardy J, Singleton
808 AB, Dale AM, Gasser T, Andreassen OA, Sharma M,
809 International Parkinson's Disease Genomics Consortium
810 (IPDGC), North American Brain Expression Consor-
811 tium (NABEC), and United Kingdom Brain Expression
812 Consortium (UKBEC) Investigators (2017) Genome-wide
813 pleiotropy between Parkinson disease and autoimmune
814 diseases. *JAMA Neurol* **74**, 780-792.
- [22] 815 Polymeropoulos MH, Lavedan C, Leroy E, Ide SE, Dehe-
816 jia A, Dutra A, Pike B, Root H, Rubenstein J, Boyer
817 R, Stenroos ES, Chandrasekharappa S, Athanassiadou A,
818 Papapetropoulos T, Johnson WG, Lazzarini AM, Duvoisin
819 RC, Dj Iorio G, Golbe LI, Nussbaum RL (1997) Muta-
820 tion in the alpha-synuclein gene identified in families with
821 Parkinson's disease. *Science* **276**, 2045-2047.
- [23] 822 Proukakis C, Dudzik CG, Brier T, MacKay DS, Cooper
823 JM, Millhauser GL, Houlden H, Schapira AH (2013) A
824 novel α -synuclein missense mutation in Parkinson disease.
825 *Neurology* **80**, 1062-1064.
- [24] 826 Chartier-Harlin M-C, Kachergus J, Roumier C, Mouroux
827 V, Douay X, Lincoln S, Leveque C, Larvor L, Andrieux J,
828 Hulihan M, Waucquier N, Defebvre L, Amouyel P, Farrer
829 M, Destée A (2004) Alpha-synuclein locus duplication
830 as a cause of familial Parkinson's disease. *Lancet* **364**,
831 1167-1169.
- [25] 832 Braak H, Sastre M, Del Tredici K (2007) Develop-
833 ment of alpha-synuclein immunoreactive astrocytes in
834 the forebrain parallels stages of intraneuronal pathology
835 in sporadic Parkinson's disease. *Acta Neuropathol* **114**,
836 231-241.
- [26] 837 Kim C, Lee H-J, Masliah E, Lee S-J (2016) Non-
838 cell-autonomous Neurotoxicity of α -synuclein Through
839 Microglial Toll-like Receptor 2. *Exp Neurobiol* **25**,
840 113-119.
- [27] 841 Sarkar S, Dammer EB, Malovic E, Olsen AL, Raza SA,
842 Gao T, Xiao H, Oliver DL, Duong D, Joers V, Seyfried
843 N, Huang M, Kukar T, Tansey MG, Kanthasamy AG,
844 Rangaraju S (2020) Molecular signatures of neuroinflam-
845 mation induced by α Synuclein aggregates in microglial
846 cells. *Front Immunol* **11**, 33.
- [28] 847 Rostami J, Fotaki G, Sirois J, Mzezewa R, Bergström
848 J, Essand M, Healy L, Erlandsson A (2020) Astrocytes
849 have the capacity to act as antigen-presenting cells in the
850 Parkinson's disease brain. *J Neuroinflammation* **17**, 119.

- 851 [29] Ferreira SA, Romero-Ramos M (2018) Microglia response
852 during Parkinson's disease: Alpha-synuclein intervention.
853 *Front Cell Neurosci* **12**, 247.
- 854 [30] Stolzenberg E, Berry D, Yang D, Lee EY, Kroemer A,
855 Kaufman S, Wong GCL, Oppenheim JJ, Sen S, Fishbein
856 T, Bax A, Harris B, Barbut D, Zasloff MA (2017) A role
857 for neuronal alpha-synuclein in gastrointestinal immunity.
858 *J Innate Immun* **9**, 456-463.
- 859 [31] Olesen MN, Christiansen JR, Petersen SV, Jensen PH,
860 Paslawski W, Romero-Ramos M, Sanchez-Guajardo V
861 (2018) CD4 T cells react to local increase of α -synuclein
862 in a pathology-associated variant-dependent manner and
863 modify brain microglia in absence of brain pathology.
864 *Heliyon* **4**, e00513.
- 865 [32] Shameli A, Xiao W, Zheng Y, Shyu S, Sumodi J, Meyer-
866 son HJ, Harding CV, Maitta RW (2016) A critical role for
867 alpha-synuclein in development and function of T lym-
868 phocytes. *Immunobiology* **221**, 333-340.
- 869 [33] Xiao W, Shameli A, Harding CV, Meyerson HJ, Maitta
870 RW (2014) Late stages of hematopoiesis and B cell lym-
871 phopoiesis are regulated by α -synuclein, a key player in
872 Parkinson's disease. *Immunobiology* **219**, 836-844.
- 873 [34] Eittle B, Kuhbandner K, Jörg S, Hoffmann A, Winkler J,
874 Linker RA (2016) α -Synuclein deficiency promotes neu-
875 roinflammation by increasing Th1 cell-mediated immune
876 responses. *J Neuroinflammation* **13**, 201.
- 877 [35] Cebrián C, Zucca FA, Mauri P, Steinbeck JA, Studer L,
878 Scherzer CR, Kanter E, Budhu S, Mandelbaum J, Vonsattel
879 JP, Zecca L, Loike JD, Sulzer D (2014) MHC-I expression
880 renders catecholaminergic neurons susceptible to T-cell-
881 mediated degeneration. *Nat Commun* **5**, 3633.
- 882 [36] Sulzer D, Alcalay RN, Garretti F, Cote L, Kanter E, Agin-
883 Liebes J, Liang C, McMurtrey C, Hildebrand WH, Mao
884 X, Dawson VL, Dawson TM, Oseroff C, Pham J, Sidney J,
885 Dillon MB, Carpenter C, Weiskopf D, Phillips E, Mallal
886 S, Peters B, Frazier A, Lindestam Arlehamn CS, Sette
887 A (2017) T cells from patients with Parkinson's disease
888 recognize α -synuclein peptides. *Nature* **546**, 656-661.
- 889 [37] Lindestam Arlehamn CS, Dhanwani R, Pham J, Kuan R,
890 Frazier A, Rezende Dutra J, Phillips E, Mallal S, Roederer
891 M, Marder KS, Amara AW, Standaert DG, Goldman JG,
892 Litvan I, Peters B, Sulzer D, Sette A (2020) α -Synuclein-
893 specific T cell reactivity is associated with preclinical and
894 early Parkinson's disease. *Nat Commun* **11**, 1875.
- 895 [38] Hernandez DG, Reed X, Singleton AB (2016) Genetics
896 in Parkinson disease: Mendelian versus non-Mendelian
897 inheritance. *J Neurochem* **139**(Suppl 1), 59-74.
- 898 [39] Williams ET, Chen X, Moore DJ (2017) VPS35, the
899 retromer complex and Parkinson's disease. *J Parkinsons*
900 *Dis* **7**, 219-233.
- 901 [40] Li J-G, Chiu J, Praticò D (2020) Full recovery of
902 the Alzheimer's disease phenotype by gain of function
903 of vacuolar protein sorting 35. *Mol Psychiatry* **25**,
904 2630-2640.
- 905 [41] Ye S-Y, Apple JE, Ren X, Tang F-L, Yao L-L, Wang Y-G,
906 Mei L, Zhou Y-G, Xiong W-C (2019) Microglial VPS35
907 deficiency regulates microglial polarization and decreases
908 ischemic stroke-induced damage in the cortex. *J Neuroin-*
909 *flammation* **16**, 235.
- 910 [42] Yin J, Liu X, He Q, Zhou L, Yuan Z, Zhao S (2016)
911 Vps35-dependent recycling of Trem2 regulates microglial
912 function. *Traffic* **17**, 1286-1296.
- 913 [43] Appel JR, Ye S, Tang F, Sun D, Zhang H, Mei L, Xiong
914 W-C (2018) Increased microglial activity, impaired adult
915 hippocampal neurogenesis, and depressive-like behavior
916 in microglial VPS35-depleted mice. *J Neurosci* **38**, 5949-
917 5968.
- 918 [44] Steger M, Diez F, Dhekne HS, Lis P, Nirujogi RS, Karayel
919 O, Tonelli F, Martinez TN, Lorentzen E, Pfeffer SR,
920 Alessi DR, Mann M (2017) Systematic proteomic anal-
921 ysis of LRRK2-mediated Rab GTPase phosphorylation
922 establishes a connection to ciliogenesis. *Elife* **6**, e31012.
- 923 [45] Mir R, Tonelli F, Lis P, Macartney T, Polinski NK, Mar-
924 tinez TN, Chou M-Y, Howden AJM, König T, Hotzy C,
925 Milenkovic I, Brücke T, Zimprich A, Sammler E, Alessi
926 DR (2018) The Parkinson's disease VPS35[D620N]
927 mutation enhances LRRK2-mediated Rab protein phos-
928 phorylation in mouse and human. *Biochem J* **475**,
929 1861-1883.
- 930 [46] Day JO, Mullin S (2021) The genetics of Parkinson's
931 disease and implications for clinical practice. *Genes* **12**,
932 1006.
- 933 [47] Quinn PMJ, Moreira PI, Ambrósio AF, Alves CH (2020)
934 PINK1/PARKIN signalling in neurodegeneration and neu-
935 roinflammation. *Acta Neuropathol Commun* **8**, 189.
- 936 [48] Ge P, Dawson VL, Dawson TM (2020) PINK1 and
937 Parkin mitochondrial quality control: A source of regional
938 vulnerability in Parkinson's disease. *Mol Neurodegener*
939 **15**, 20.
- 940 [49] Xu S, Yang X, Qian Y, Xiao Q (2018) Parkinson's
941 disease-related DJ-1 modulates the expression of uncou-
942 pling protein 4 against oxidative stress. *J Neurochem* **145**,
943 312-322.
- 944 [50] Borsche M, König IR, Delcambre S, Petrucci S, Balck A,
945 Brüggemann N, Zimprich A, Wasner K, Pereira SL, Aven-
946 ali M, Deuschle C, Badanjak K, Ghelfi J, Gasser T, Kasten
947 M, Rosenstiel P, Lohmann K, Brockmann K, Valente EM,
948 Youle RJ, Grünewald A, Klein C (2020) Mitochondrial
949 damage-associated inflammation highlights biomarkers in
950 PRKN/PINK1 parkinsonism. *Brain* **143**, 3041-3051.
- 951 [51] Sliter DA, Martinez J, Hao L, Chen X, Sun N, Fischer
952 TD, Burman JL, Li Y, Zhang Z, Narendra DP, Cai H,
953 Borsche M, Klein C, Youle RJ (2018) Parkin and PINK1
954 mitigate STING-induced inflammation. *Nature* **561**,
955 258-262.
- 956 [52] Matheoud D, Sugiura A, Bellemare-Pelletier A, Laplante
957 A, Rondeau C, Chemali M, Fazel A, Bergeron JJ, Trudeau
958 L-E, Burelle Y, Gagnon E, McBride HM, Desjardins M
959 (2016) Parkinson's disease-related proteins PINK1 and
960 parkin repress mitochondrial antigen presentation. *Cell*
961 **166**, 314-327.
- 962 [53] Mouton-Liger F, Rosazza T, Sepulveda-Diaz J, Jeang A,
963 Hassoun S-M, Claire E, Mangone G, Brice A, Michel PP,
964 Corvol J-C, Corti O (2018) Parkin deficiency modulates
965 NLRP3 inflammasome activation by attenuating an A20-
966 dependent negative feedback loop. *Glia* **66**, 1736-1751.
- 967 [54] Lin Q, Li S, Jiang N, Shao X, Zhang M, Jin H, Zhang Z,
968 Shen J, Zhou Y, Zhou W, Gu L, Lu R, Ni Z (2019) PINK1-
969 parkin pathway of mitophagy protects against contrast-
970 induced acute kidney injury via decreasing mitochondrial
971 ROS and NLRP3 inflammasome activation. *Redox Biol*
972 **26**, 101254.
- 973 [55] Zhou Y, Shi X, Chen H, Zhang S, Salker MS, Mack AF,
974 Föllner M, Mak TW, Singh Y, Lang F (2017) DJ-1/Park7
975 sensitive Na/H exchanger 1 (NHE1) in CD4 T cells. *J Cell*
976 *Physiol* **232**, 3050-3059.
- 977 [56] Sun L, Shen R, Agnihotri SK, Chen Y, Huang Z,
978 Büeler H (2018) Lack of PINK1 alters glia innate
979 immune responses and enhances inflammation-induced,
980 nitric oxide-mediated neuron death. *Sci Rep* **8**, 383.

- 981 [57] Choi D-J, An J, Jou I, Park SM, Joe E-H (2019) A Parkinson's disease gene, DJ-1, regulates anti-inflammatory roles of astrocytes through prostaglandin D synthase expression. *Neurobiol Dis* **127**, 482-491. 1046
- 982 1047
- 983 1048
- 984 1049
- 985 [58] Singh Y, Chen H, Zhou Y, Föllmer M, Mak TW, Salker MS, Lang F (2015) Differential effect of DJ-1/PARK7 on development of natural and induced regulatory T cells. *Sci Rep* **5**, 17723. 1050
- 986 1051
- 987 1052
- 988 1053
- 989 [59] Avenali M, Blandini F, Cerri S (2020) Glucocerebrosidase defects as a major risk factor for Parkinson's disease. *Front Aging Neurosci* **12**, 97. 1054
- 990 1055
- 991 1056
- 992 [60] Panicker LM, Miller D, Park TS, Patel B, Azevedo JL, Awad O, Masood MA, Veenstra TD, Goldin E, Stubblefield BK, Tayebi N, Polumuri SK, Vogel SN, Sidransky E, Zambidis ET, Feldman RA (2012) Induced pluripotent stem cell model recapitulates pathologic hallmarks of Gaucher disease. *Proc Natl Acad Sci U S A* **109**, 18054-18059. 1057
- 993 1058
- 994 1059
- 995 1060
- 996 1061
- 997 1062
- 998 1063
- 999 1064
- 1000 [61] Mizukami H, Mi Y, Wada R, Kono M, Yamashita T, Liu Y, Werth N, Sandhoff R, Sandhoff K, Proia RL (2002) Systemic inflammation in glucocerebrosidase-deficient mice with minimal glucosylceramide storage. *J Clin Invest* **109**, 1215-1221. 1065
- 1001 1066
- 1002 1067
- 1003 1068
- 1004 [62] Hughes LP, Pereira MMM, Hammond DA, Kwok JB, Halliday GM, Lewis SJG, Dzamko N (2021) Glucocerebrosidase activity is reduced in cryopreserved Parkinson's disease patient monocytes and inversely correlates with motor severity. *J Parkinsons Dis* **11**, 1157-1165. 1069
- 1005 1070
- 1006 1071
- 1007 1072
- 1008 1073
- 1009 1074
- 1010 [63] Atashrazm F, Hammond D, Perera G, Dobson-Stone C, Mueller N, Pickford R, Kim WS, Kwok JB, Lewis SJG, Halliday GM, Dzamko N (2018) Reduced glucocerebrosidase activity in monocytes from patients with Parkinson's disease. *Sci Rep* **8**, 15446. 1075
- 1011 1076
- 1012 1077
- 1013 1078
- 1014 1079
- 1015 [64] Mullin S, Stokholm MG, Hughes D, Mehta A, Parbo P, Hinz R, Pavese N, Brooks DJ, Schapira AHV (2021) Brain microglial activation increased in glucocerebrosidase (GBA) mutation carriers without Parkinson's disease. *Mov Disord* **36**, 774-779. 1080
- 1016 1081
- 1017 1082
- 1018 1083
- 1019 1084
- 1020 [65] Sanyal A, DeAndrade MP, Novis HS, Lin S, Chang J, Lengacher N, Tomlinson JJ, Tansey MG, LaVoie MJ (2020) Lysosome and inflammatory defects in GBA1-mutant astrocytes are normalized by LRRK2 inhibition. *Mov Disord* **35**, 760-773. 1085
- 1021 1086
- 1022 1087
- 1023 1088
- 1024 1089
- 1025 [66] Keatinge M, Bui H, Menke A, Chen Y-C, Sokol AM, Bai Q, Ellett F, Da Costa M, Burke D, Gegg M, Trollope L, Payne T, McTighe A, Mortiboys H, de Jager S, Nuthall H, Kuo M-S, Fleming A, Schapira AHV, Renshaw SA, Highley JR, Chacinska A, Panula P, Burton EA, O'Neill MJ, Bandmann O (2015) Glucocerebrosidase 1 deficient Danio rerio mirror key pathological aspects of human Gaucher disease and provide evidence of early microglial activation preceding alpha-synuclein-independent neuronal cell death. *Hum Mol Genet* **24**, 6640-6652. 1090
- 1026 1091
- 1027 1092
- 1028 1093
- 1029 1094
- 1030 1095
- 1031 [67] Thaler A, Omer N, Giladi N, Gurevich T, Bar-Shira A, Gana-Weisz M, Goldstein O, Kestenbaum M, Shirvan JC, Cedarbaum JM, Orr-Urtreger A, Regev K, Shenhar-Tsarfaty S, Mirelman A (2021) Mutations in GBA and LRRK2 are not associated with increased inflammatory markers. *J Parkinsons Dis* **11**, 1285-1296. 1096
- 1032 1097
- 1033 1098
- 1034 1099
- 1035 1100
- 1036 1101
- 1037 1102
- 1038 [68] Foo JN, Chew EGY, Chung SJ, Peng R, Blauwendraat C, Nalls MA, Mok KY, Satake W, Toda T, Chao Y, Tan LCS, Tandiono M, Lian MM, Ng EY, Prakash K-M, Au W-L, Meah W-Y, Mok SQ, Annuar AA, Chan AYY, Chen L, Chen Y, Jeon BS, Jiang L, Lim JL, Lin J-J, Liu C, Mao C, Mok V, Pei Z, Shang H-F, Shi C-H, Song K, Tan AH, Wu Y-R, Xu Y-M, Xu R, Yan Y, Yang J, Zhang B, Koh W-P, Lim S-Y, Khor CC, Liu J, Tan E-K (2020) Identification of risk loci for Parkinson disease in Asians and comparison of risk between Asians and Europeans: A genome-wide association study. *JAMA Neurol* **77**, 746-754. 1103
- 1039 1104
- 1040 1105
- 1041 1106
- 1042 1107
- 1043 1108
- 1044 1109
- 1045 1110
- 1110 [69] Loesch DP, Horimoto ARVR, Heilbron K, Sarihan EI, Inca-Martinez M, Mason E, Cornejo-Olivas M, Torres L, Mazzetti P, Cosentino C, Sarapura-Castro E, Rivera-Valdivia A, Medina AC, Dieguez E, Raggio V, Lescano A, Tumas V, Borges V, Ferraz HB, Rieder CR, Schumacher-Schuh A, Santos-Lobato BL, Velez-Pardo C, Jimenez-Del-Rio M, Lopera F, Moreno S, Chana-Cuevas P, Fernandez W, Arboleda G, Arboleda H, Arboleda-Bustos CE, Yearout D, Zabetian CP, 23andMe Research Team, Cannon P, Thornton TA, O'Connor TD, Mata IF, Latin American Research Consortium on the Genetics of Parkinson's Disease (LARGE-PD) (2021) Characterizing the genetic architecture of Parkinson's disease in Latinos. *Ann Neurol* **90**, 353-365. 1111
- [70] Bandres-Ciga S, Saez-Atienzar S, Kim JJ, Makarios MB, Faghri F, Diez-Fairen M, Iwaki H, Leonard H, Botia J, Ryten M, Hernandez D, Gibbs JR, Ding J, Gan-Or Z, Noyce A, Pihlstrom L, Torkamani A, Soltis AR, Dalgard CL, American Genome Center, Scholz SW, Traynor BJ, Ehrlich D, Scherzer CR, Bookman M, Cookson M, Blauwendraat C, Nalls MA, Singleton AB, International Parkinson Disease Genomics Consortium (2020) Large-scale pathway specific polygenic risk and transcriptomic community network analysis identifies novel functional pathways in Parkinson disease. *Acta Neuropathol* **140**, 341-358. 1112
- [71] Li YI, Wong G, Humphrey J, Raj T (2019) Prioritizing Parkinson's disease genes using population-scale transcriptomic data. *Nat Commun* **10**, 994. 1113
- [72] Gagliano SA, Pouget JG, Hardy J, Knight J, Barnes MR, Ryten M, Weale ME (2016) Genomics implicates adaptive and innate immunity in Alzheimer's and Parkinson's diseases. *Ann Clin Transl Neurol* **3**, 924-933. 1114
- [73] Andersen MS, Bandres-Ciga S, Reynolds RH, Hardy J, Ryten M, Krohn L, Gan-Or Z, Holtman IR, Pihlstrom L, International Parkinson's Disease Genomics Consortium (2021) Heritability enrichment implicates microglia in Parkinson's disease pathogenesis. *Ann Neurol* **89**, 942-951. 1115
- [74] Walker DG, Tang TM, Mendsaikhan A, Tooyama I, Serano GE, Sue LI, Beach TG, Lue L-F (2020) Patterns of expression of purinergic receptor P2RY12, a putative marker for non-activated microglia, in aged and Alzheimer's disease brains. *Int J Mol Sci* **21**, 678. 1116
- [75] Lopes K de P, Snijders GJL, Humphrey J, Allan A, Sneebouer MAM, Navarro E, Schilder BM, Vialle RA, Parks M, Missall R, van Zuiden W, Gigase FAJ, Kübler R, van Berlekom AB, Hicks EM, Böttcher C, Priller J, Kahn RS, de Witte LD, Raj T (2022) Genetic analysis of the human microglial transcriptome across brain regions, aging and disease pathologies. *Nat Genet* **54**, 4-17. 1117
- [76] Chang D, Nalls MA, Hallgrímsson IB, Hunkapiller J, van der Brug M, Cai F, International Parkinson's Disease Genomics Consortium, 23andMe Research Team, Kerchner GA, Ayalon G, Bingol B, Sheng M, Hinds D, Behrens TW, Singleton AB, Bhargava TR, Graham RR (2017) A meta-analysis of genome-wide association studies identifies 17 new Parkinson's disease risk loci. *Nat Genet* **49**, 1511-1516. 1118

- 1111 [77] Kondo M, Tanaka Y, Kuwabara T, Naito T, Kohwi-
1112 Shigematsu T, Watanabe A (2016) SATB1 plays a critical
1113 role in establishment of immune tolerance. *J Immunol* **196**,
1114 563-572.
- 1115 [78] Galiano-Landeira J, Torra A, Vila M, Bové J (2020) CD8
1116 T cell nigral infiltration precedes synucleinopathy in early
1117 stages of Parkinson's disease. *Brain* **143**, 3717-3733.
- 1118 [79] Zhang W, Wang T, Pei Z, Miller DS, Wu X, Block ML,
1119 Wilson B, Zhang W, Zhou Y, Hong J-S, Zhang J (2005)
1120 Aggregated alpha-synuclein activates microglia: A process
1121 leading to disease progression in Parkinson's disease.
1122 *FASEB J* **19**, 533-542.
- 1123 [80] Klegeris A, Giasson BI, Zhang H, Maguire J, Pelech
1124 S, McGeer PL (2006) Alpha-synuclein and its disease-
1125 causing mutants induce ICAM-1 and IL-6 in human
1126 astrocytes and astrocytoma cells. *FASEB J* **20**, 2000-2008.
- 1127 [81] Allen Reish HE, Standaert DG (2015) Role of α -synuclein
1128 in inducing innate and adaptive immunity in Parkinson
1129 disease. *J Parkinsons Dis* **5**, 1-19.
- 1130 [82] Grozdanov V, Bousset L, Hoffmeister M, Bliederhaeuser
1131 C, Meier C, Madiona K, Pieri L, Kiechle M, McLean
1132 PJ, Kassubek J, Behrends C, Ludolph AC, Weishaupt JH,
1133 Melki R, Danzer KM (2019) Increased immune activa-
1134 tion by pathologic α -synuclein in Parkinson's disease. *Ann*
1135 *Neurol* **86**, 593-606.
- 1136 [83] Saporito MS, Thomas BA, Scott RW (2000) MPTP acti-
1137 vates c-Jun NH(2)-terminal kinase (JNK) and its upstream
1138 regulatory kinase MKK4 in nigrostriatal neurons *in vivo*.
1139 *J Neurochem* **75**, 1200-1208.
- 1140 [84] Huang G, Shi LZ, Chi H (2009) Regulation of JNK and
1141 p38 MAPK in the immune system: Signal integration,
1142 propagation and termination. *Cytokine* **48**, 161-169.
- 1143 [85] Zhang Y, Nallaparaju KC, Liu X, Jiao H, Reynolds JM,
1144 Wang Z-X, Dong C (2015) MAPK phosphatase 7 regulates
1145 T cell differentiation via inhibiting ERK-mediated IL-2
1146 expression. *J Immunol* **194**, 3088-3095.
- 1147 [86] Bachstetter AD, Xing B, de Almeida L, Dimayuga ER,
1148 Watterson DM, Van Eldik LJ (2011) Microglial p38 α
1149 MAPK is a key regulator of proinflammatory cytokine up-
1150 regulation induced by toll-like receptor (TLR) ligands or
1151 beta-amyloid (A β). *J Neuroinflammation* **8**, 79.
- 1152 [87] Gallagher MD, Posavi M, Huang P, Unger TL, Berlyand Y,
1153 Gruenewald AL, Chesi A, Manduchi E, Wells AD, Grant
1154 SFA, Blobel GA, Brown CD, Chen-Plotkin AS (2017) A
1155 dementia-associated risk variant near TMEM106B alters
1156 chromatin architecture and gene expression. *Am J Hum*
1157 *Genet* **101**, 643-663.
- 1158 [88] Tropea TF, Mak J, Guo MH, Xie SX, Suh E, Rick J,
1159 Siderowf A, Weintraub D, Grossman M, Irwin D, Wolk
1160 DA, Trojanowski JQ, Van Deerlin V, Chen-Plotkin AS
1161 (2019) TMEM106B Effect on cognition in Parkinson
1162 disease and frontotemporal dementia. *Ann Neurol* **85**, 801-
1163 811.
- 1164 [89] Rhinn H, Abeliovich A (2017) Differential aging anal-
1165 ysis in human cerebral cortex identifies variants in
1166 TMEM106B and GRN that regulate aging phenotypes.
1167 *Cell Syst* **4**, 404-415.e5.
- 1168 [90] Ren Y, van Blitterswijk M, Allen M, Carrasquillo MM,
1169 Reddy JS, Wang X, Beach TG, Dickson DW, Ertekin-
1170 Taner N, Asmann YW, Rademakers R (2018) TMEM106B
1171 haplotypes have distinct gene expression patterns in aged
1172 brain. *Mol Neurodegener* **13**, 35.
- 1173 [91] Li P, Marshall L, Oh G, Jakubowski JL, Groot D, He Y,
1174 Wang T, Petronis A, Labrie V (2019) Epigenetic dys-
1175 regulation of enhancers in neurons is associated with
1176 Alzheimer's disease pathology and cognitive symptoms. *Nat Commun* **10**, 2246.
- 1177 [92] Sharma A, Osato N, Liu H, Asthana S, Dakal TC,
1178 Ambrosini G, Bucher P, Schmitt I, Wüllner U (2019) Com-
1179 mon genetic variants associated with Parkinson's disease
1180 display widespread signature of epigenetic plasticity. *Sci*
1181 *Rep* **9**, 18464.
- 1182 [93] Zakharova MY, Belyanina TA, Sokolov AV, Kiselev IS,
1183 Mamedov AE (2019) The contribution of major histocom-
1184 patibility complex class II genes to an association with
1185 autoimmune diseases. *Acta Naturae* **11**, 4-12.
- 1186 [94] Muñoz-Castrillo S, Vogrig A, Honnorat J (2020) Asso-
1187 ciations between HLA and autoimmune neurological
1188 diseases with autoantibodies. *Auto Immun Highlights*
1189 **11**, 2.
- 1190 [95] Hamza TH, Zabetian CP, Tenesa A, Laederach A, Mon-
1191 timurro J, Yearout D, Kay DM, Doheny KF, Paschall J,
1192 Pugh E, Kusel VI, Collura R, Roberts J, Griffith A, Samii
1193 A, Scott WK, Nutt J, Factor SA, Payami H (2010) Com-
1194 mon genetic variation in the HLA region is associated
1195 with late-onset sporadic Parkinson's disease. *Nat Genet*
1196 **42**, 781-785.
- 1197 [96] Yu E, Ambati A, Andersen MS, Krohn L, Estiar MA, Saini
1198 P, Senkevich K, Sosero YL, Sreelatha AAK, Ruskey JA,
1199 Asayesh F, Spiegelman D, Toft M, Viken MK, Sharma M,
1200 Blauwendraat C, Pihlström L, Mignot E, Gan-Or Z (2021)
1201 Fine mapping of the HLA locus in Parkinson's disease in
1202 Europeans. *NPJ Parkinsons Dis* **7**, 84.
- 1203 [97] Hill-Burns EM, Factor SA, Zabetian CP, Thomson G,
1204 Payami H (2011) Evidence for more than one Parkinson's
1205 disease-associated variant within the HLA region. *PLoS*
1206 *One* **6**, e27109.
- 1207 [98] Hollenbach JA, Norman PJ, Creary LE, Damotte V,
1208 Montero-Martin G, Caillier S, Anderson KM, Misra MK,
1209 Nemat-Gorgani N, Osoegawa K, Santaniello A, Renschen
1210 A, Marin WM, Dandekar R, Parham P, Tanner CM, Hauser
1211 SL, Fernandez-Viña M, Oksenberg JR (2019) A specific
1212 amino acid motif of mediates risk and interacts with smok-
1213 ing history in Parkinson's disease. *Proc Natl Acad Sci U*
1214 *S A* **116**, 7419-7424.
- 1215 [99] Chuang Y-H, Lee P-C, Vlaar T, Mulot C, Loriot M-A,
1216 Hansen J, Lill CM, Ritz B, Elbaz A (2017) Pooled analysis
1217 of the HLA-DRB1 by smoking interaction in Parkinson
1218 disease. *Ann Neurol* **82**, 655-664.
- 1219 [100] Noyce AJ, Bandres-Ciga S, Kim J, Heilbron K, Kia D,
1220 Hemani G, Xue A, Lawlor DA, Smith GD, Duran R,
1221 Gan-Or Z, Blauwendraat C, Gibbs JR, 23andMe Research
1222 Team5, International Parkinson's Disease Genomics Con-
1223 sortium (IPDGC), Hinds DA, Yang J, Visscher P, Cuzick
1224 J, Morris H, Hardy J, Wood NW, Nalls MA, Singleton AB
1225 (2019) The Parkinson's Disease Mendelian Randomiza-
1226 tion Research Portal. *Mov Disord* **34**, 1864-1872.
- 1227 [101] Chang C-C, Lin T-M, Chang Y-S, Chen W-S, Sheu J-
1228 J, Chen Y-H, Chen J-H (2018) Autoimmune rheumatic
1229 diseases and the risk of Parkinson disease: A nationwide
1230 population-based cohort study in Taiwan. *Ann Med* **50**,
1231 83-90.
- 1232 [102] Li CY, Yang TM, Ou RW, Wei QQ, Shang HF (2021)
1233 Genome-wide genetic links between amyotrophic lateral
1234 sclerosis and autoimmune diseases. *BMC Med* **19**, 27.
- 1235 [103] Li C, Ou R, Shang H (2021) Rheumatoid arthritis
1236 decreases risk for Parkinson's disease: A Mendelian ran-
1237 domization study. *NPJ Parkinsons Dis* **7**, 17.
- 1238 [104] Bacelis J, Compagno M, George S, Pospisilik JA, Brundin
1239 P, Nalwai ÁT, Brundin L (2021) Decreased risk of
1240

- 1241 Parkinson's disease after rheumatoid arthritis diagnosis: A
1242 nested case-control study with matched cases and controls.
1243 *J Parkinsons Dis* **11**, 821-832.
- 1244 [105] Derkinderen P, Neunlist M (2018) Crohn's and Parkinson
1245 disease: Is LRRK2 lurking around the corner? *Nat Rev*
1246 *Gastroenterol Hepatol* **15**, 330-331.
- 1247 [106] Liu Z, Lee J, Krummey S, Lu W, Cai H, Lenardo MJ (2011)
1248 The kinase LRRK2 is a regulator of the transcription factor
1249 NFAT that modulates the severity of inflammatory bowel
1250 disease. *Nat Immunol* **12**, 1063-1070.
- 1251 [107] Ikezu T, Koro L, Wolozin B, Farraye FA, Strongosky
1252 AJ, Wszolek ZK (2020) Crohn's and Parkinson's disease-
1253 associated LRRK2 mutations alter type II interferon
1254 responses in human CD14+ blood monocytes *ex vivo*. *J*
1255 *Neuroimmune Pharmacol* **15**, 794-800.
- 1256 [108] Li D, Achkar J-P, Haritunians T, Jacobs JP, Hui KY,
1257 D'Amato M, Brand S, Radford-Smith G, Halfvarson J,
1258 Niess J-H, Kugathasan S, Büning C, Schumm LP, Klei L,
1259 Ananthakrishnan A, Aumais G, Baidoo L, Dubinsky M,
1260 Fiocchi C, Glas J, Milgrom R, Proctor DD, Regueiro M,
1261 Simms LA, Stempak JM, Targan SR, Törkvist L, Sharma
1262 Y, Devlin B, Borneman J, Hakonarson H, Xavier RJ, Daly
1263 M, Brant SR, Rioux JD, Silverberg MS, Cho JH, Braun J,
1264 McGovern DPB, Duerr RH (2016) A pleiotropic missense
1265 variant in SLC39A8 is associated with Crohn's disease
1266 and human gut microbiome composition. *Gastroenterol-*
1267 *ogy* **151**, 724-732.
- [109] Pickrell JK, Berisa T, Liu JZ, Ségurel L, Tung JY, Hinds
1268 DA (2016) Detection and interpretation of shared genetic
1269 influences on 42 human traits. *Nat Genet* **48**, 709-717.
1270
- [110] Prigent A, Lionnet A, Durieu E, Chapelet G, Bourreille A,
1271 Neunlist M, Rolli-Derkinderen M, Derkinderen P (2019)
1272 Enteric alpha-synuclein expression is increased in Crohn's
1273 disease. *Acta Neuropathol* **137**, 359-361.
1274
- [111] Prigent A, Chapelet G, De Guilhem de Lataillade A, Oul-
1275 lier T, Durieu E, Bourreille A, Duchalais E, Hardonnière K,
1276 Neunlist M, Noble W, Kerdine-Römer S, Derkinderen P,
1277 Rolli-Derkinderen M (2020) Tau accumulates in Crohn's
1278 disease gut. *FASEB J* **34**, 9285-9296.
1279
- [112] Tang G, Pan H, Xu L, Feng R, Jiang Y, Kong F, Hu S (2018)
1280 A comparison of co-methylation relationships between
1281 rheumatoid arthritis and Parkinson's disease. *Front Neu-*
1282 *rosci* **12**, 1001.
1283
- [113] Xie A, Ensink E, Li P, Gordevičius J, Marshall LL, George
1284 S, Andrew Pospisilik J, Aho VTE, Houser MC, Pereira
1285 PAB, Rudi K, Paulin L, Tansey MG, Auvinen P, Brundin
1286 P, Brundin L, Labrie V, Scheperjans F (2021) Butyrate
1287 and related epigenetic changes link Parkinson's disease
1288 to inflammatory bowel disease and depressive symptoms.
1289 *medRxiv* 2021.09.17.21263343.
1290