

Editorial

Saliva is a Good Candidate to be the New Gold-Standard Sample for Neurodegenerative Diseases

Gorka Orive^{a,b,c,*}, Francisco Lopera^d and Eva Carro^{e,f,*}

^a*Laboratory of Pharmacy and Pharmaceutical Technology, Faculty of Pharmacy, University of the Basque Country/Euskal Herriko Unibertsitatea (UPV/EHU), Vitoria, Spain*

^b*Bioaraba, NanoBioCel Research Group, Vitoria-Gasteiz, Spain*

^c*Networking Center for Biomedical Research in Bioengineering Biomaterials and Nanomedicine (CIBER-BBN) Barcelona, Spain*

^d*Grupo de Neurociencias, Universidad de Antioquia. Medellín, Colombia*

^e*Neurobiology of Alzheimer's Disease Unit, Chronic Disease Programme, Instituto de Salud Carlos III, Madrid, Spain*

^f*Network Centre for Biomedical Research in Neurodegenerative Diseases (CIBERNED), Spain*

Accepted 31 March 2022

Pre-press 25 April 2022

Although still not considered a traditional diagnostic sample type, saliva testing is a simple, non-invasive, sustainable, and affordable approach for the diagnosis of infectious and non-infectious diseases. In the last two years, diagnostic tests have been developed to fight the coronavirus disease 2019 (COVID-19) pandemic, and saliva specimens represent a highly specific and sensitive alternative diagnostic sample to detect SARS-CoV-2, even at the level of the nasopharyngeal swab [1]. Indeed, some countries are adopting saliva testing for SARS-CoV-2 detection, including South Korea, Germany, and

Japan [2, 3]. This global and unusual situation support new point of view for the potential of saliva specimens in the diagnosis and management of diseases.

Saliva has emerged as a good source of samples for detection of disease biomarkers. The use of saliva as a diagnostic sample has several advantages. Saliva offers a new and easily accessible physiological fluid that can be collected in a non-invasive manner and assessed using different analytical assays. It is easy to collect and does not need specialized personnel. It may save time, is comfortable, and may be even more relevant as it is scalable and inexpensive, being able to extend its use to developing countries. In addition to its ease of collection, saliva is generally safer than blood and cerebrospinal fluid (CSF), and its collection does not expose the healthcare provider to needles, thus reducing the risk of pathogen transmission from patients suffering from chronic infection.

Several products based on saliva testing are already on the market. Commercially available kits can gauge the levels of a handful of hormones, including

* Corresponding authors. Dr. Gorka Orive, Laboratory of Pharmacy and Pharmaceutical Technology, Faculty of Pharmacy, University of the Basque Country/Euskal Herriko Unibertsitatea (UPV/EHU), Vitoria, Spain. Tel.: +34663027696; E-mail: gorka.orive@ehu.es.; ORCID: 0000-0002-0773-300X and Dr. Eva Carro, Neurobiology of Alzheimer's disease Unit, Chronic Disease Programme, Instituto de Salud Carlos III, 28222 Majadahonda, Madrid, Spain. Tel.: +34 918223995; E-mail: eva.carro@isciii.es.; ORCID: 0000-0002-6504-4579

estrogen, testosterone, and cortisol, from a sample of saliva. Saliva has the potential to diagnose diseases with more complex origins, including cancer [4]. Accumulating evidence has demonstrated the diagnostic and prognostic value of saliva as promising novel and revolutionary liquid biopsy in cancer [5].

Recent progress suggest saliva testing could be a useful approach for the diagnosis of neurodegenerative diseases. Early disease detection is critical in assigning proper treatment therapy to affected patients with these diseases. Easily accessible, cost-effective, and accurate diagnostic biomarkers are need to improve the early diagnosis of these neurodegenerative diseases, especially in primary care [6]. Usually the tests performed for the diagnosis of neurological conditions are lumbar puncture or more recently blood tests. Their invasive nature, especially for the lumbar puncture, usually results in discomfort, pain, and disagreeable side effects for patients, which necessitates the search for accurate, more advanced, and less invasive testing methods. Positron emission tomography (PET) imaging is also a very valuable diagnostic tool, but very costly which also limits its use in clinical practice. Therefore, it is essential to establish a substitute that is less invasive but remains representative of the brain's pathological changes.

Saliva is being explored as an alternative to CSF and imaging biomarkers in the accurate detection of Alzheimer disease (AD) [7]. A collection of studies have shown that salivary amyloid- β ($A\beta$)₄₂ is detectable and increased in AD [8–10], whereas presence of tau species has been also explored in saliva but with inconclusive results to support these molecules as salivary biomarkers for AD [11–13]. However, salivary lactoferrin has shown very high accuracy and specificity to differentiate prodromal and dementia stages of AD from healthy subjects [14]. Current research continues to supply evidence indicating that salivary lactoferrin concentration is significantly reduced in AD patients when compared to healthy controls, patients suffering from frontotemporal dementia and Parkinson's disease (PD) [15], and even in memory impaired older subjects associated with brain $A\beta$ burden [16]. Moreover, hypothalamic dysfunctions in AD may preclude immunity alterations, including reduction in salivary lactoferrin [17, 18], although other authors have not been able to replicate these findings [19]. Other recent and remarkable study showed that salivary GFAP levels decreased in mild cognitive impairment and AD patients and were proven a potential biomarker for AD [20].

Alternatively, to the use of saliva for the identification of circulating biological markers to help the diagnosis of early cognitive impairment associated with AD, saliva could also be useful to generate insights into the potential application of stem cells derived from salivary glands or saliva as therapeutics for the disease [21]. Previously, the presence of adult stem cells with mesenchymal characteristics in human parotid gland tissue was described [22]. New studies have reported the establishment of mesenchymal stem cell lines derived from mouse submandibular glands [23, 24]. As it is well known, mesenchymal stem cells have marked potential for use in cell therapy and regenerative medicine. In recent years, studies supporting the potential use of mesenchymal stem cell therapy for the treatment of autoimmune diseases have been published reporting their ability to modulate the immune response [25, 26]. Very recently, it has been observed that the treatment with salivary gland-derived mesenchymal stem cells via tail vein decreased the expression of interleukin 17 (IL-17), interferon gamma, and IL-6 levels and enhanced transforming growth factor beta and IL-10 secretion and restore salivary gland secretory function in the mouse models of Sjögren's syndrome [22].

PD is the second most common neurodegenerative disorder after AD, and it is also subject the difficulty in accurately diagnosing. Since α -synuclein (α -syn) is both genetically and pathologically linked to PD, it has been proposed as promising candidate biomarker [27]. More recently, it has reported α -syn as a potential biomarker of PD diagnosis in peripheral tissues [28]. Postmortem submandibular gland biopsies are positive for Lewy-type α -syn in patients with PD but not in healthy subjects [29, 30], and this finding has made salivary α -syn one of the most investigated salivary biomarkers in neurodegeneration. A decade ago, Devic and colleagues showed that α -syn concentrations significantly decrease in the saliva of PD patients as compared to healthy controls [31]. These results observed in saliva are mirrored in CSF [32]. These findings were later replicated by Al-Nimer [33] and Vivacqua [34] teams as they detected a significant decrease in total α -syn ($\text{syn}_{\text{total}}$) in saliva of PD patients when compared to healthy controls, while α -syn oligomers (α -syn_{olig}) and α -syn_{olig}/ α -syn_{total} ratio exhibited a significant increase in the saliva of PD patients as compared to healthy controls [34–36]. Moreover, salivary α -syn measurement revealed specific cut-off values able to differentiate PD patients from healthy subjects with high sensitivity and

specificity, being even higher for the α -syn olig/ α -syntotal ratio (69.77% and 95.16%, respectively) [35].

Interestingly, dysregulation of microRNAs (miRNAs) has been implicated in various neurodegenerative conditions, including PD [37, 38]. Salivary miR-153 and miR-223 levels may serve as useful, noninvasive, and relatively inexpensive diagnostic biomarkers of idiopathic PD based on their sensitivity (81% and 72%, respectively) and specificity (71%) [39]. These miRNAs were found to regulate α -syn expression in brain [40], and salivary miR-153 and miR-223 down-modulation in PD could reflect primary changes in the central nervous system.

In Huntington's disease (HD), huntingtin protein was successfully detected in saliva of HD patients and healthy controls. There was a significant increase in total huntingtin (Htt) protein concentration in saliva samples obtained from HD patients when compared to controls [41]. Given that currently a non-invasive measure of Htt CNS concentration does not exist, salivary Htt might be proposed as an early detection biomarker for HD, although new studies measuring sensitivity and specificity of salivary Htt will be needed.

However, investigation on salivary biomarkers for neurodegenerative diseases remains questionable. Although saliva is a fluid not in contact directly or indirectly with CNS, it is secreted by salivary glands directly regulated by cholinergic parasympathetic nerves connected with the hypothalamus, a brain area seriously affected in AD [42–45]. In addition to the structural abnormalities, including amyloid plaques and neurofibrillary tangles, functional studies suggest that hypothalamic dysfunction is a common AD manifestation, often an early event in the course of disease [43]. We propose that AD-related hypothalamic alterations could result in dysregulation of salivary gland function [17], as recent studies reported [18, 46]. Previous evidence suggests that A β can induce cholinergic hypofunction [47] whereas activation of muscarinic receptors can inhibit the generation of amyloidogenic A β [48, 49]. In our recent study, a reduction in M3 muscarinic receptor levels was observed in human submandibular glands from AD patients and APP/PS1 mice [18]. This last down-regulation of muscarinic receptors in AD salivary glands could be behind the increase in salivary A β levels. However, the mechanisms leading these salivary alterations in AD related proteins are not completely understood.

Although much evidence over the last decade supports the use of blood-based biomarkers for

investigating AD, as it has recently reviewed [50], blood drawing itself is also an invasive technique. Saliva collection is non-invasive, economical, safe, and simple and can be performed without the assistance of specialized health care personnel, even home collection, allowing for point-of-injury sampling.

While many diseases have confirmed salivary biomarkers [51], diseases affecting the nervous system have few confirmed markers available in saliva which are still being investigated. Important considerations must be taken in account when using saliva samples as diagnostic tools. Standardization and consensus in the saliva collection, processing and storage is needed in order to decrease biases and allow an accurate identification of salivary biomarkers, as we recently proposed [52]. Intrinsic and extrinsic factors affecting donors might have influenced the production of salivary proteins. Moreover, intra- or inter-laboratory variability of salivary analyses may influence the diagnostic classification. These variances may be attributable to differences between duplicate assays of the same sample within laboratories or associated with differences between laboratories. The need of reproducing research assay was also highlighted by Ashton and colleagues [53]. Thus, efforts on harmonization of procedures should be encouraged to optimize the accuracy of salivary biomarkers in AD. In any case, saliva-based methods may open new windows in disease diagnosis if more standardized, efficient, and broadly implementable kits are developed. Thought procedure, confounding variables, and protocols need to be optimized in the future, some of these testing products could become a glimmer of hope for many communities globally.

ACKNOWLEDGMENTS

Gorka Orive wishes to thank the Spanish Ministry of Economy, Industry, and Competitiveness (PID2019-106094RB-I00/AEI/10.13039/501100011033) and technical assistance from the ICTS NAN BIOSIS (Drug Formulation Unit, U10) at the University of the Basque Country. We also appreciate the support from the Basque Country Government (Grupos Consolidados, No ref: IT907-16). Eva Carro wishes to thank grants from Instituto de Salud Carlos III (FIS18/00118 to E.C.), FEDER, Comunidad de Madrid (S2017/BMD-3700; NEUROMETAB-CM to E.C), and CIBERNED (CB07/502 to E.C.). Francisco Lopera wishes to thank grants from Banner, NIH, Roche, Enroll-HD, Large-PD, and Open Philanthropy.

Dr. Eva Carro and Dr. Gorka Orive are co-founders of GEROA Diagnostics.

Authors' disclosures available online (<https://www.j-alz.com/manuscript-disclosures/22-0144r2>).

REFERENCES

- [1] Tan SH, Allicock O, Armstrong-Hough M, Wyllie AL (2021) Saliva as a gold-standard sample for SARS-CoV-2 detection. *Lancet Respir Med* **9**, 562-564.
- [2] Oba J, Taniguchi H, Sato M, Takamatsu R, Morikawa S, Nakagawa T, Takaishi H, Saya H, Matsuo K, Nishihara H (2021) RT-PCR screening tests for SARS-CoV-2 with saliva samples in asymptomatic people: Strategy to maintain social and economic activities while reducing the risk of spreading the virus. *Keio J Med* **70**, 35-43.
- [3] Yokota I, Shane PY, Okada K, Unoki Y, Yang Y, Iwasaki S, Fujisawa S, Nishida M, Teshima T (2021) A novel strategy for SARS-CoV-2 mass screening with quantitative antigen testing of saliva: A diagnostic accuracy study. *Lancet Microbe* **2**, e397-e404.
- [4] Kaur J, Jacobs R, Huang Y, Salvo N, Politis C (2018) Salivary biomarkers for oral cancer and pre-cancer screening: A review. *Clin Oral Investig* **22**, 633-640.
- [5] Rapado-González Ó, Majem B, Álvarez-Castro A, Díaz-Peña R, Abalo A, Suárez-Cabrera L, Gil-Moreno A, Santamaría A, López-López R, Muínelo-Romay L, Suarez-Cunqueiro MM (2019) A novel saliva-based miRNA signature for colorectal cancer diagnosis. *J Clin Med* **8**, 2029.
- [6] Hansson O (2021) Biomarkers for neurodegenerative diseases. *Nat Med* **27**, 954-963.
- [7] Ashton NJ, Ide M, Zetterberg H, Blennow K (2019) Salivary biomarkers for Alzheimer's disease and related disorders. *Neurol Ther* **8**, 83-94.
- [8] Bermejo-Pareja F, Antequera D, Vargas T, Molina JA, Carro E (2010) Saliva levels of Abeta1-42 as potential biomarker of Alzheimer's disease: A pilot study. *BMC Neurol* **10**, 108.
- [9] Lee M, Guo JP, Kennedy K, McGeer EG, McGeer PL (2017) A method for diagnosing Alzheimer's disease based on salivary amyloid-beta protein 42 levels. *J Alzheimers Dis* **55**, 1175-1182.
- [10] Sabbagh MN, Shi J, Lee M, Arnold L, Al-Hasan Y, Heim J, McGeer P (2018) Salivary beta amyloid protein levels are detectable and differentiate patients with Alzheimer's disease dementia from normal controls: Preliminary findings. *BMC Neurol* **18**, 155.
- [11] Ashton NJ, Ide M, Scholl M, Blennow K, Lovestone S, Hye A, Zetterberg H (2018) No association of salivary total tau concentration with Alzheimer's disease. *Neurobiol Aging* **70**, 125-127.
- [12] Pেকেles H, Qureshi HY, Paudel HK, Schipper HM, Gornistky M, Chertkow H (2019) Development and validation of a salivary tau biomarker in Alzheimer's disease. *Alzheimers Dement (Amst)* **11**, 53-60.
- [13] Gleerup HS, Hasselbalch SG, Simonsen AH (2019) Biomarkers for Alzheimer's disease in saliva: A systematic review. *Dis Markers* **2019**, 4761054.
- [14] Carro E, Bartolomé F, Bermejo-Pareja F, Villarejo-Galende A, Molina JA, Ortiz P, Calero M, Rabano A, Cantero JL, Orive G (2017) Early diagnosis of mild cognitive impairment and Alzheimer's disease based on salivary lactoferrin. *Alzheimers Dement (Amst)* **8**, 131-138.
- [15] González-Sánchez M, Bartolome F, Antequera D, Puertas-Martín V, González P, Gómez-Grande A, Llamas-Velasco S, Herrero-San Martín A, Pérez-Martínez D, Villarejo-Galende A, Atienza M, Palomar-Bonet M, Cantero JL, Perry G, Orive G, Ibañez B, Bueno H, Fuster V, Carro E (2020) Decreased salivary lactoferrin levels are specific to Alzheimer's disease. *EBioMedicine* **57**, 102834.
- [16] Reseco L, Atienza M, Fernandez-Alvarez M, Carro E, Cantero JL (2021) Salivary lactoferrin is associated with cortical amyloid-beta load, cortical integrity, and memory in aging. *Alzheimers Res Ther* **13**, 150.
- [17] Bermejo-Pareja F, Del Ser T, Valentí M, de la Fuente M, Bartolome F, Carro E (2020) Salivary lactoferrin as biomarker for Alzheimer's disease: Brain-immunity interactions. *Alzheimers Dement* **16**, 1196-1204.
- [18] Antequera D, Moneo D, Carrero L, Bartolome F, Ferrer I, Proctor G, Carro E (2021) Salivary lactoferrin expression in a mouse model of Alzheimer's disease. *Front Immunol* **12**, 749468.
- [19] Gleerup HS, Jensen CS, Høgh P, Hasselbalch SG, Simonsen AH (2021) Lactoferrin in cerebrospinal fluid and saliva is not a diagnostic biomarker for Alzheimer's disease in a mixed memory clinic population. *EBioMedicine* **67**, 103361.
- [20] Katsipis G, Tzekaki EE, Tsolaki M, Pantazaki AA (2021) Salivary GFAP as a potential biomarker for diagnosis of mild cognitive impairment and Alzheimer's disease and its correlation with neuroinflammation and apoptosis. *J Neuroimmunol* **361**, 577744.
- [21] Reale M, Gonzales-Portillo I, Borlongan CV (2020) Saliva, an easily accessible fluid as diagnostic tool and potent stem cell source for Alzheimer's disease: Present and future applications. *Brain Res* **1727**, 146535.
- [22] Rotter N, Oder J, Schlenke P, Lindner U, Böhrnsen F, Kramer J, Rohwedel J, Huss R, Brandau S, Wollenberg B, Lang S (2008) Isolation and characterization of adult stem cells from human salivary glands. *Stem Cells Dev* **17**, 509-518.
- [23] Furukawa S, Kuwajima Y, Chosa N, Satoh K, Ohtsuka M, Miura H, Kimura M, Inoko H, Ishisaki A, Fujimura A, Miura H (2015) Establishment of immortalized mesenchymal stem cells derived from the submandibular glands of tdTomato transgenic mice. *Exp Ther Med* **10**, 1380-1386.
- [24] Lim JY, Yi T, Lee S, Kim J, Kim SN, Song SU, Kim YM (2015) Establishment and characterization of mesenchymal stem cell-like clonal stem cells from mouse salivary glands. *Tissue Eng Part C Methods* **21**, 447-457.
- [25] Tyndall A, van Laar JM (2016) Stem cell transplantation and mesenchymal cells to treat autoimmune diseases. *Presse Med* **45**, e159-169.
- [26] Li B, Xing Y, Gan Y, He J, Hua H (2021) Labial gland-derived mesenchymal stem cells and their exosomes ameliorate murine Sjögren's syndrome by modulating the balance of Treg and Th17 cells. *Stem Cell Res Ther* **12**, 478.
- [27] El-Agnaf OM, Salem SA, Paleologou KE, Curran MD, Gibson MJ, Court JA, Schlossmacher MG, Allsop D (2006) Detection of oligomeric forms of alpha-synuclein protein in human plasma as a potential biomarker for Parkinson's disease. *FASEB J* **20**, 419-425.
- [28] Fayyad M, Salim S, Majbour N, Erskine D, Stoops E, Mollenhauer B, El-Agnaf OMA (2019) Parkinson's disease biomarkers based on alpha-synuclein. *J Neurochem* **150**, 626-636.
- [29] Del Tredici K, Hawkes CH, Ghebremedhin E, Braak H (2010) Lewy pathology in the submandibular gland of

- individuals with incidental Lewy body disease and sporadic Parkinson's disease. *Acta Neuropathol* **119**, 703-713.
- [30] Vilas D, Iranzo A, Tolosa E, Aldecoa I, Berenguer J, Vilaseca I, Martí C, Serradell M, Lomena F, Alos L, Gaig C, Santamaria J, Gelpi E (2016) Assessment of alpha-synuclein in submandibular glands of patients with idiopathic rapid-eye-movement sleep behaviour disorder: A case-control study. *Lancet Neurol* **15**, 708-718.
- [31] Devic I, Hwang H, Edgar JS, Izutsu K, Presland R, Pan C, Goodlett DR, Wang Y, Armaly J, Tumas V, Zabetian CP, Leverenz JB, Shi M, Zhang J (2011) Salivary alpha-synuclein and DJ-1: Potential biomarkers for Parkinson's disease. *Brain* **134**, e178.
- [32] Parnetti L, Castrioto A, Chiasserini D, Persichetti E, Tambasco N, El-Agnaf O, Calabresi P (2013) Cerebrospinal fluid biomarkers in Parkinson disease. *Nat Rev Neurol* **9**, 131-140.
- [33] Al-Nimer MS, Mshat SF, Abdulla HI (2014) Saliva alpha-synuclein and a high extinction coefficient protein: A novel approach in assessment biomarkers of Parkinson's disease. *N Am J Med Sci* **6**, 633-637.
- [34] Vivacqua G, Latorre A, Suppa A, Nardi M, Pietracupa S, Mancinelli R, Fabbrini G, Colosimo C, Gaudio E, Berardelli A (2016) Abnormal salivary total and oligomeric alpha-synuclein in Parkinson's disease. *PLoS One* **11**, e0151156.
- [35] Vivacqua G, Suppa A, Mancinelli R, Belvisi D, Fabbrini A, Costanzo M, Formica A, Onori P, Fabbrini G, Berardelli A (2019) Salivary alpha-synuclein in the diagnosis of Parkinson's disease and progressive supranuclear palsy. *Parkinsonism Relat Disord* **63**, 143-148.
- [36] Kang W, Chen W, Yang Q, Zhang L, Zhang L, Wang X, Dong F, Zhao Y, Chen S, Quinn TJ, Zhang J, Chen S, Liu J (2016) Salivary total alpha-synuclein, oligomeric alpha-synuclein and SNCA variants in Parkinson's disease patients. *Sci Rep* **6**, 28143.
- [37] Basak I, Patil KS, Alves G, Larsen JP, Møller SG (2016) microRNAs as neuroregulators, biomarkers and therapeutic agents in neurodegenerative diseases. *Cell Mol Life Sci* **73**, 811-827.
- [38] Martinez B, Peplow PV (2017) MicroRNAs in Parkinson's disease and emerging therapeutic targets. *Neural Regen Res* **12**, 1945-1959.
- [39] Cressatti M, Juwara L, Galindez JM, Velly AM, Nkurunziza ES, Marier S, Canie O, Gornistky M, Schipper HM (2020) Salivary microR-153 and microR-223 levels as potential diagnostic biomarkers of idiopathic Parkinson's disease. *Mov Disord* **35**, 468-477.
- [40] Cressatti M, Song W, Turk AZ, Garabed LR, Benchaya JA, Galindez C, Liberman A, Schipper HM (2019) Glial HMOX1 expression promotes central and peripheral α -synuclein dysregulation and pathogenicity in parkinsonian mice. *Glia* **67**, 1730-1744.
- [41] Corey-Bloom J, Haque AS, Park S, Nathan AS, Baker RW, Thomas EA (2018) Salivary levels of total huntingtin are elevated in Huntington's disease patients. *Sci Rep* **8**, 7371.
- [42] Baloyannis SJ, Mavroudis I, Mitiileos D, Baloyannis IS, Costa VG (2015) The hypothalamus in Alzheimer's disease: A Golgi and electron microscope study. *Am J Alzheimers Dis Other Demen* **30**, 478-487.
- [43] Ishii M, Iadecola C (2015) Metabolic and non-cognitive manifestations of Alzheimer's disease: The hypothalamus as both culprit and target of pathology. *Cell Metab* **22**, 761-776.
- [44] Goldstein DS, Kopin IJ (2017) Homeostatic systems, biocybernetics, and autonomic neuroscience. *Auton Neurosci* **208**, 15-28.
- [45] Zheng H, Zhou Q, Du Y, Li C, Xu P, Lin L, Xiao J, Gao H (2018) The hypothalamus as the primary brain region of metabolic abnormalities in APP/PS1 transgenic mouse model of Alzheimer's disease. *Biochim Biophys Acta Mol Basis Dis* **1864**, 263-273.
- [46] Zalewska A, Klimiuk A, Zięba S, Wnorowska O, Rusak M, Waszkiewicz N, Szarmach I, Dzierżanowski K, Maciejczyk M (2021) Salivary gland dysfunction and salivary redox imbalance in patients with Alzheimer's disease. *Sci Rep* **11**, 23904.
- [47] Auld DS, Kar S, Quirion R (1998) Beta-amyloid peptides as direct cholinergic neuromodulators: A missing link? *Trends Neurosci* **21**, 43-49.
- [48] Lin L, Georgievska B, Mattsson A, Isacson O (1999) Cognitive changes and modified processing of amyloid precursor protein in the cortical and hippocampal system after cholinergic synapse loss and muscarinic receptor activation. *Proc Natl Acad Sci U S A* **96**, 12108-12113.
- [49] Gu Z, Zhong P, Yan Z (2003) Activation of muscarinic receptors inhibits beta-amyloid peptide-induced signaling in cortical slices. *J Biol Chem* **278**, 17546-17556.
- [50] Leuzy A, Mattsson-Carlgrén N, Palmqvist S, Janelidze S, Dage JL, Hansson O (2022) Blood-based biomarkers for Alzheimer's disease. *EMBO Mol Med* **14**, e14408.
- [51] Streckfus CF, Bigler LR (2002) Saliva as a diagnostic fluid. *Oral Dis* **8**, 69-76.
- [52] Bartolome F, Orive G, Carro E (2021) Standardizing salivary lactoferrin measurements to obtain a robust diagnostic biomarker for Alzheimer's disease. *Alzheimers Dement (Amst)* **13**, e12173.
- [53] Ashton NJ, Blennow K, Zetterberg H (2021) Spitting image: Can saliva biomarkers reflect Alzheimer's disease? *EBioMedicine* **68**, 103437.