

Short Article

COVID-19 and the endothelium

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Abstract. There is growing evidence that COVID-19 not only affects the lungs but beyond that the endothelial system. Recent studies showed that this can lead to microcirculatory impairments and in consequence to functional disorders of all inner organs. The combination of endothelial dysfunction with a generalized inflammatory state and complement elements may together contribute to the overall pro-coagulative state described in COVID-19 patients leading to venular as well as to arteriolar occlusions.

In December 2019, an outbreak of pneumonia due to a novel corona virus – the severe acute respiratory syndrome corona virus 2 (SARS-CoV-2) – occurred in Wuhan, Hubei province, China. The virus causing the corona virus disease 2019 (COVID-19) has since spread to many countries resulting in a pandemic [1]. Symptoms most commonly reported include fever, cough, shortness of breath and loss of sense of taste and smell. Older patients with co-morbidities are more likely to develop respiratory failure due to severe alveolar damage [2]. In more severe cases, the disease can also show a rapid progression to organ failure, with complications, such as shock, acute respiratory distress syndrome (ARDS), acute cardiac injury, acute kidney injury, disseminated intravascular coagulopathy (DIC), which may ultimately prove fatal [3]. Recent observations suggest that respiratory failure in COVID-19 is not driven by the development of ARDS alone [4], but that macro-vascular as well as micro-vascular thrombotic processes may play a role [5, 6]. It is becoming apparent that severe cases of COVID-19 are characterized by hyper inflammation and a thrombotic phenomenon. Such major adverse clinical events seem to suggest that in advanced stages of this disease one target is the endothelium, one of the largest organs in the human body. Whether vascular derangements in COVID-19 are due to endothelial cell dysfunction is currently unknown.

The main transmission route is through virus containing droplet or aerosol but also smear transmission is possible. The virus replicates in the upper respiratory tract decent to lower respiratory tract. The spread of the virus in the host is thought to occur also via the vascular system and transgression

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34 into mucosal tissues of nose, throat and especially of the lungs, where the endothelium comes into
35 contact with SARS CoV-2 at an early stage. The virus enters endothelial cells by endocytosis via the
36 binding of its spike glycoprotein to a cellular receptor which facilitates viral attachment to the surface
37 of target cells [7–9]. Angiotensin-converting enzyme 2 (ACE2) was identified as the main receptor for
38 severe acute respiratory syndrome corona virus (SARS-CoV 2) [10], which is abundantly expressed
39 in the lungs, the respiratory epithelium and alveolar monocytes [11] and may explain the many cases
40 of rapidly occurring lung failure. However, ACE2 is also expressed by endothelial cells [12–14] in the
41 heart and the macro vascular system, gut, kidneys, liver, central nervous system, and adipose tissue
42 [15–17]. As the density of ACE2 differs in the various tissues – very high in the lungs – the receptor
43 density may correlate with the severity of the disease in those tissue [18–20]. In addition, there are
44 further receptors on the surface of human cells which can mediate the entry of SARS-CoV-2, includ-
45 ing transmembrane serine protease 2 (TMPRSS2 [19]), sialic acid receptors [21], and extracellular
46 matrix metalloproteinase inducer (CD147 [22]). These four receptors are known to be expressed by
47 endothelial cells [23–26].

48 Whereas unperturbed endothelial cells provide very potent anti-coagulant properties [27], exposure
49 to inflammatory stimuli can rapidly lead to a procoagulant behavior. Very recently, Varga et al. found
50 evidence of direct SARS-CoV-2 infection of endothelial cells in several organs and diffuse endothe-
51 lial inflammation associated with apoptosis [28]. They described this state as endotheliitis with viral
52 elements within endothelial cells and accumulation of inflammatory cells, with evidence of endothe-
53 lial and inflammatory cell death. Endothelial cell injury can strongly activate the coagulation system
54 via exposure of tissue factor and other pathways. Endothelial dysfunction refers to a systemic condi-
55 tion in which the endothelium loses its physiological properties, including the tendency to promote
56 vasodilation, fibrinolysis, and anti-aggregation [27]. Narrowing of organ supplying arteries as well
57 as microcirculatory disturbances in liver, spleen and kidneys in patients with severe COVID-19 were
58 already described recently [29].

59 COVID-19-endotheliitis could explain the impaired microcirculation in different vascular beds [30]
60 and their clinical sequelae in patients with COVID-19. During inflammatory activation or apoptosis,
61 endothelial cells become pro-coagulant [31] and release microvesicles (MV) which can affect the
62 function of target cells through surface interaction and receptor activation, cellular fusion and the
63 delivery of intra-vesicular cargo [31]. Among others, endothelial MV have been described to affect
64 hemostasis [27]. Figure 1 shows a picture with apoptotic human umbilical venous endothelial cells
65 with massive signs of blebbing and detached microvesicles.

66 Pathological investigations and also imaging studies confirmed the COVID-19 disease as a thrombo-
67 inflammatory process that initially affects the lungs and in consequence the perfusion, which
68 consecutively can affect all organs of the body. It is well known that a variety of viruses can affect
69 the coagulation system including HIV, Dengue virus, and Ebola virus [32, 33]. Very recently, Spiezia
70 et al. reported that SARS-CoV-2 may predispose patients to thrombotic disease, both in the venous
71 and arterial circulations [34]. Excessive inflammation, platelet activation, endothelial dysfunction, and
72 stasis related to the infection were described [35], which can result in severe hypercoagulability and
73 predispose to thrombosis. That has been related to deaths in critically ill COVID-19 cases [36]. A
74 single-center retrospective cohort study of 183 patients with confirmed COVID-19 evaluated coag-
75 ulation abnormalities that mimic disseminated intravascular coagulation (DIC) [37]. According to
76 the International Society on Thrombosis and Hemostasis definition of DIC, 15 of 21 non-survivors
77 (71%) were classified as having overt-DIC (≥ 5 points) at any time during follow-up, whereas only 1
78 of 162 survivors (0.6%) met these criteria ($P < 0.001$). Likewise, Tang et al. reported that 71.4% of
79 non-survivors and 0.6% of survivors of COVID-19 showed evidence of overt DIC [38]. This is in line
80 with another study, in which a clear correlation between D-dimer levels, disease progression and chest
81 CT features suggesting venous thrombosis were reported [39]. Also, pulmonary embolism is with 30%



Fig. 1. Apoptotic endothelial cells with signs of blebbing and detached microvesicles (cLSM, objective $\times 40$).

82 significantly more frequent in COVID-19 patients [40] than usually occurring in critically ill patients
 83 without COVID-19 infection (1.3%, [41]) or in emergency department patients (3 to 10% [42]). Tee et
 84 al. were able show that in the lung a vascular areas occur which most likely represent 3–5 mm micro
 85 infarcts [43] which confirms the micro vascular involvement in the course of the disease.

86 In conclusion, it seems that COVID-19 is a disease affecting the lungs and, beyond that, the
 87 endothelial system. Recent studies show that this can lead to microcirculatory impairments, and in
 88 consequence to functional disorders of all inner organs. The combination of endothelial dysfunction
 89 with a generalized inflammatory state and complement elements may together contribute to the overall
 90 pro-coagulative state described in COVID-19 patients.

91 References

- 92 [1] World Health Organization. Coronavirus disease 2019 (COVID-19). Situation report – 70 [Internet]. Geneva:
 93 World Health Organization; 2020. Available at: [https://apps.who.int/iris/bitstream/handle/10665/331683/nCoVsitrep](https://apps.who.int/iris/bitstream/handle/10665/331683/nCoVsitrep30Mar2020-eng.pdf)
 94 [30Mar2020-eng.pdf](https://apps.who.int/iris/bitstream/handle/10665/331683/nCoVsitrep30Mar2020-eng.pdf) (Accessed: 26 April 2020).
- 95 [2] Yang X, Yu Y, Xu J, Shu H, Xia J, Liu H, Wu Y, Zhang L, Yu Z, Fang M, Yu T, Wang Y, Pan S, Zou X, Yuan S,
 96 Shang Y. Clinical Course and Outcomes of Critically Ill Patients With SARS-CoV-2 Pneumonia in Wuhan, China: A
 97 Single-Centered, Retrospective, Observational Study. *Lancet Respir Med.* 2020;8(5):475-81.
- 98 [3] Deng Y, Liu W, Liu K, et al. Clinical characteristics of fatal and recovered cases of coronavirus disease 2019 (COVID-19)
 99 in Wuhan, China: a retro-spective study. *Chin Med J (Engl).* 2020. <https://doi.org/10.1097/CM9.0000000000000824>
- 100 [4] Magro C, Mulvey JJ, Berlin D, Nuovo G, Salvatore S, Harp J, Baxter-Stoltzfus A, Laurence J. Complement associated
 101 microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: A report of five cases. *Transl*
 102 *Res.* 2020. doi: 10.1016/j.trsl.2020.04.007
- 103 [5] Jung F, Krieger V, Hufert FT, Küpper J-H. How we should respond to the Coronavirus SARS-CoV-2 outbreak: A German
 104 perspective. *Clin Hemorheol Microcirc.* 2020. DOI: 10.3233/CH-170277

- 105 [6] Oudkerk M, Büller HR, Kuijpers D, van Es N, Oudkerk SF, McCloud TC, Gommers, van Dissel J, Ten Cate H, van Beek
106 EJ. Diagnosis, Prevention, and Treatment of thromboembolic Complications in COVID-19: Report of the National
107 Institute for Public Health of the Netherlands. *Radiology*. 2020;201629. doi: 10.1148/radiol.2020201629
- 108 [7] Letko M, Marzi A and Munster V. Functional assessment of cell entry and receptor usage for SARS-CoV-2 and other
109 lineage B betacoronaviruses. *Nat Microbiol*. 2020;5:562-9.
- 110 [8] Wang Q, Zhang Y, Wu L, Niu S, Song C, Zhang Z, Lu G, Qiao C, Hu Y, Yuen KY, Wang Q, Zhou H, Yan J, Qi J. Structural
111 and functional basis of SARS-CoV-2 entry by using human ACE2. *Cell*. 2020. <https://doi.org/10.1016/j.cell.2020.03.045>
- 112 [9] Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, Schiergens TS, Herrler G, Wu NH,
113 Nitsche A, Müller MA, Drosten C, Pöhlmann S. SARS-CoV-2. Cell Entry Depends on ACE2 and TMPRSS2 and Is
114 Blocked by a Clinically Proven Protease Inhibitor. *Cell*. 2020;181(2):271-280.e8.
- 115 [10] Monteil V KH, Prado P, Hagelkriüys A, et al. Inhibition of SARS-CoV-2 infections in engineered human tissues using
116 clinical-grade soluble human ACE2. *Cell*. 2020. https://www.cell.com/pbassets/products/coronavirus/CELL_CELL-D-20-00739.pdf
- 117 [11] Hamming I, Timens W, Bulthuis ML, Lely AT, Navis G, van Goor H. Tissue distribution of ACE2 protein, the functional
118 receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. *J Pathol*. 2004;203:631-7.
- 119 [12] Nicin L, Abplanalp WT, Mellentin H, Kattih B, Tombor L, John D, Schmitto JD, Heineke J, Emrich F, Arsalan M,
120 Holubec T, Walther T, Zeiher AM, Dimmeler S. Cell type-specific expression of the putative SARS-CoV-2 receptor
121 ACE2 in human hearts. *European Heart Journal*. 2020. doi:10.1093/eurheartj/ehaa311
- 122 [13] Sluimer JC, Gasc JM, Hamming I, van Goor H, Michaud A, van den Akker LH, Jutten B, Cleutjens J, Bijmens AP,
123 Corvol P, Daemen MJ and Heeneman S. Angiotensin-converting enzyme 2 (ACE2) expression and activity in human
124 carotid atherosclerotic lesions. *J Pathol*. 2008;215:273-9.
- 125 [14] Lu R, Zhao X, Li J, Ni P, Yang B, Wu H, Wang W, Song H, Huang B, Zhu N, Bi Y, Ma X, Zhan F, Wang L, Hu T, Zhou
126 H, Hu Z, Zhou W, Zhao L, Chen J, Meng Y, Wang J, Lin Y, Yuan J, Xie Z, Ma J, Liu WJ, Wang D, Xu W, Holmes
127 EC, Gao GF, Wu G, Chen W, Shi W, Tan W. Genomic Characterisation and Epidemiology of 2019 Novel Coronavirus:
128 Implications for Virus Origins and Receptor Binding. *Lancet*. 2020;395(10224):565-74.
- 129 [15] Ferrario CM, Jessup J, Chappell MC, et al. Effect of angiotensin-converting enzyme inhibition and angiotensin II
130 receptor blockers on cardiac angiotensin-converting enzyme 2. *Circulation*. 2005;111:2605-10.
- 131 [16] Gheblawi M, Wang K, Viveiros A, Q Nguyen, Zhong J-C, Turner AJ, Raizada MK, Grant MB, Oudit GY. Angiotensin-
132 Converting Enzyme 2: SARS-CoV-2 Receptor and Regulator of the Renin-Angiotensin System. *Circ Res*. 2020;126(10).
133 doi: 10.1161/CIRCRESAHA.120.317015
- 134 [17] Chai X, Hu L, Zhang Y, Han W, Lu Z, Ke A, Zhou J, Shi G, Fang N, Fan J, Cai J, Fan J, Lan F. Specific ACE2
135 Expression in Cholangiocytes May Cause Liver Damage After 2019-nCoV Infection. *BioRxiv preprint Server*. doi:
136 <https://doi.org/10.1101/2020.02.03.931766>
- 137 [18] Xu H, Zhong L, Deng J, Peng J, Dan H, Zeng X, Li T and Chen Q. High expression of ACE2 receptor of 2019-nCoV
138 on the epithelial cells of oral mucosa. *Int J Oral Sci*. 2020;12:8.
- 139 [19] Perico L, Benigni A, Remuzzi G. Should COVID-19 Concern Nephrologists? Why and to What Extent? The Emerging
140 Impasse of Angiotensin Blockade. *Nephron*. 2020:1-9.
- 141 [20] Matsuyama S, Nao N, Shirato K, Kawase M, Saito S, Takayama I, Nagata N, Sekizuka T, Katoh H, Kato F, Sakata M,
142 Tahara M, Kutsuna S, Ohmagari N, Kuroda M, Suzuki T, Kageyama T, Takeda M. Enhanced isolation of SARS-CoV-2
143 by TMPRSS2-expressing cells. *Proc Natl Acad Sci U S A*. 2020;117:7001-3.
- 144 [21] Tortorici MA, Walls AC, Lang Y, Wang C, Li Z, Koerhuis D, Boons GJ, Bosch BJ, Rey FA, de Groot RJ and Veisler
145 D. Structural basis for human coronavirus attachment to sialic acid receptors. *Nat Struct Mol Biol*. 2019;26:481-9.
- 146 [22] Chen Z, Mi L, Xu J, Yu J, Wang X, Jiang J, Xing J, Shang P, Qian A, Li Y, Shaw PX, Wang J, Duan S, Ding J, Fan C,
147 Zhang Y, Yang Y, Yu X, Feng Q, Li B, Yao X, Zhang Z, Li L, Xue X, Zhu P. Function of HAb18G/CD147 in invasion
148 of host cells by severe acute respiratory syndrome coronavirus. *J Infect Dis*. 2005;191:755-60.
- 149 [23] Yang J, Feng X, Zhou Q, Cheng W, Shang C, Han P, Lin CH, Chen HS, Quertermous T, Chang CP. Pathological Ace2-
150 to-Ace enzyme switch in the stressed heart is transcriptionally controlled by the endothelial Brg1-FoxM1 complex.
151 *Proc Natl Acad Sci U S A*. 2016;113:E5628-35.
- 152 [24] Aimes RT, Zijlstra A, Hooper JD, Ogbourne SM, Sit ML, Fuchs S, Gotley DC, Quigley JP, Antalis TM. Endothelial
153 cell serine proteases expressed during vascular morphogenesis and angiogenesis. *Thromb Haemost*. 2003;89:561-72.
- 154 [25] Huang DT, Lu CY, Chi YH, Li WL, Chang LY, Lai MJ, Chen JS, Hsu WM, Huang LM. Adaptation of influenza A
155 (H7N9) virus in primary human airway epithelial cells. *Sci Rep*. 2017;7:11300.
- 156 [26] Vanarsdall AL, Pritchard SR, Wisner TW, Liu J, Jardetzky TS and Johnson DC. CD147 Promotes Entry of
157 Pentamer-Expressing Human Cytomegalovirus into Epithelial and Endothelial Cells. *mbio*. 2018;9(3):e00781-18. doi:
158 10.1128/mBio.00781-18
- 159

- 160 [27] Krüger-Genge A, Blocki A, Franke RP, Jung F. Vascular Endothelial Cell Biology: An Update. *Int J Mol Sci.* 2019;20.pii:
161 E4411. doi: 10.3390/ijms20184411
- 162 [28] Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, Mehra MR, Schuepbach RA, Ruschitzka
163 F, Holger Moch H. Endothelial cell infection and endotheliitis in COVID-19. *The Lancet.* 2020. doi.org/10.1016/S0140-
164 6736(20)30937-5
- 165 [29] Jung EM, Stroszczinski C, Jung F. Contrast enhanced ultrasonography (CEUS) to detect abdominal microcirculatory
166 disorders in severe cases of COVID-19 infection: First experience. *Clin Hemorheol Microcirc.* 2020. DOI:10.3233/CH-
167 209003
- 168 [30] Bombeli T, Karsan A, Tait JF, Harlan JM. Apoptotic Vascular Endothelial Cells Become Procoagulant. *Blood.*
169 1997;89:2429-42.
- 170 [31] Vítková V, Živný J, Janota J. Endothelial Cell-Derived Microvesicles: Potential Mediators and Biomarkers of Pathologic
171 Processes. *Biomark Med.* 2018;12(2):161-75.
- 172 [32] Antoniak S, Mackman N. Multiple roles of the coagulation protease cascade during virus infection. *Blood.*
173 2014;123:2605-13.
- 174 [33] Antoniak S. The coagulation system in host defense. *Res Pract Thromb Haemost.* 2018;2:549-57.
- 175 [34] Spiezia L, Boscolo A, Poletto F, et al. COVID-19-related severe hypercoagulability in patients admitted to intensive
176 care unit for acute respiratory failure. *Thromb Haemost.* 2020.
- 177 [35] Bikdeli B, Madhavan MV, Jimenez D, et al. COVID-19 and thrombotic or thromboembolic disease: implications for
178 prevention, antithrombotic therapy, and follow-up. *J Am Coll Cardiol.* 2020. pii: S0735-1097(20)35008-7.
- 179 [36] Zhai Z, Li C, Chen Y, et al. Prevention and treatment of venous thromboembolism associated with coronavirus disease
180 2019 infection: a consensus statement before guidelines. *Thromb Haemost.* 2020.
- 181 [37] Ai T, Yang Z, Hou H, Zhan C, Chen C, Lv W, Tao Q, Sun Z, Xia L. Correlation of chest CT and RT-PCR testing in coro-
182 navirus disease 2019 (COVID-19) in China: a report of 1014 cases. *Radiology.* 2020. doi: 10.1148/radiol.2020200642
- 183 [38] Tang N, Li D, Wang X, Sun Z. Abnormal Coagulation Parameters Are Associated With Poor Prognosis in Patients With
184 Novel Coronavirus Pneumonia. *J Thromb Haemost.* 2020;18(4):844-7.
- 185 [39] Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, Xiang J, Wang Y, Song B, Gu X, Guan L, Wei Y, Li H, Wu X, Xu J, Tu S,
186 Zhang Y, Chen H, Cao B. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan,
187 China: a retrospective cohort study. *Lancet.* 2020;395:1054-62.
- 188 [40] Leonard-Lorant I, Delabranche X, Severac F, Helms J, Pautet C, Collange O, Schneider F, Labani A, Bilbault P,
189 Moliere S, Leyendecker P, Roy C, Ohana M. Acute Pulmonary Embolism in COVID-19 Patients on CT Angiography
190 and Relationship to D-Dimer Levels. *Radiology.* 2020:201561. doi: 10.1148/radiol.2020201561
- 191 [41] Lim W, Meade M, Lauzier F, et al. Failure of anticoagulant thrombo-prophylaxis: risk factors in medical-surgical
192 critically ill patients. *Crit Care Med.* 2015;43:401-10.
- 193 [42] Corrigan D, Prucnal C, Kabrhel C. Pulmonary embolism: the diagnosis, risk-stratification, treatment and disposition of
194 emergency department patients. *Clin Exp Emerg Med.* 2016;3:117-25.
- 195 [43] Tee A, Wong A, Yusuff T, Rao D, Sidhu P. Contrast-enhanced ultrasound (CEUS) of the lung reveals multiple areas of
196 microthrombi in a COVID-19 patient. *Intensive Care Med.* <https://doi.org/10.1007/s00134-020-06085-4>