

# Interstitial lung opacities in patients with severe COVID-19 pneumonia by bedside high-resolution ultrasound in association to CO<sub>2</sub> retention

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## Abstract.

**BACKGROUND:** Coronavirus disease 2019 (COVID-19) can cause acute respiratory distress Syndrome (ARDS).

**OBJECTIVE:** This single centre cross-section study aimed to grade the severity of pneumonia by bed-side lung ultrasound (LUS).

**METHODS:** A scoring system discriminates 5 levels of lung opacities: A-lines (0 points),  $\geq 3$  B-Line (1 point), coalescent B-Lines (2 points), marked pleural disruptions (3 points), consolidations (4 points). LUS (convex 1–5 MHz probe) was performed at 6 defined regions for each hemithorax either in supine or prone position. A lung aeration score (LAS, maximum 4 points) was allocated for each patient by calculating the arithmetic mean of the examined lung areas. Score levels were correlated with ventilation parameters and laboratory markers.

**RESULTS:** LAS of 20 patients with ARDS reached from 2.58 to 3.83 and was highest in the lateral right lobe (Mean 3.67). Ferritin levels (Mean 1885  $\mu\text{g/l}$ ;  $r=0.467$ ;  $p=0.051$ ) showed moderate correlation in spearman roh calculation. PaCO<sub>2</sub> level (Mean 46.75 mmHg;  $r=0.632$ ;  $p=0.005$ ) correlated significantly with LAS, while duration of ventilation, Horovitz-Index, CRP, LDH and IL-6 did not.

**CONCLUSIONS:** The proposed LAS describes severity of lung opacities in COVID-19 patients and correlates with CO<sub>2</sub> retention in patients with ARDS.

Keywords: COVID-19, lung ultrasound, scoring system, PaCO<sub>2</sub>.

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## 31 1. Introduction

32 Coronavirus disease 2019 (COVID-19) was described first in China in December 2019 and led  
33 to a global pandemic now affecting most countries of the world. Mortality is driven by a severe  
34 pneumonia which occurs in about 25 % of patients infected with the corona virus [1–3] Mortality rate  
35 differs by countries and infection rate is still raising in some countries. COVID-19 pneumonia shows  
36 characteristic signs in computed tomography (CT) scan with pleura-near parenchymal infiltrations,  
37 described as ground-glass opacities and patchy shadowing, with most prominent findings in patients  
38 after one to three weeks after onset of disease [4]. CT scan turned out to be the gold standard for  
39 detection of COVID-19 pneumonia in the initial phase of the pandemic [5]. The peripheral localisation  
40 of the parenchymal involvement of COVID-19 pneumonia makes these abnormalities accessible to lung  
41 ultrasound (LUS). However, data on specificity of LUS in Covid-19 is lacking, since the differentiation  
42 of bacterial consolidations and COVID-19 inflammatory pulmonary changes may be difficult to detect  
43 for LUS and central pulmonary changes can only be detected by CT. In addition, for central pulmonary  
44 changes high resolution CT is superior, while for the detection of peripheral embolism CEUS could be  
45 helpful [4–7]. On the one hand, diagnosis by bed-side point-of-care LUS minimizes the need of patient  
46 transport and therefore might have less risk of contamination than CT scan. On the other hand, LUS  
47 needs time to perform and special protection and cleaning procedures are necessary for the ultrasound  
48 machines. Findings in LUS describe different grades, ranging from normal A-lines, to more than 3  
49 single B-lines, confluent B-lines, pleural disruptions and consolidations with air bronchograms [8].

50 Aim of this pilot study was to implement a scoring system that quantifies lung alterations in ventilated  
51 patients with ARDS caused by COVID-19 pneumonia and to correlate the grade of the score with  
52 ventilation parameters and laboratory variables.

## 53 2. Material and methods

54 LUS is a very competent diagnostic tool for the detection and characterization of pulmonal changes,  
55 especially in patients with acute respiratory distress syndrome (ARDS) [9].

56 Bed-side high resolution LUS was performed in 20 patients with COVID-19 pneumonia, who were  
57 ventilated at intensive care units of the University hospital of Regensburg from 03/2020 to 06/2020.  
58 Diagnosis of COVID-19 was verified by positive polymerase chain reaction of SARS-CoV-19 (EZ-1  
59 Advanced XL extraction (Qiagen)) and CT scan with COVID-19-typical findings. All patients included  
60 in this study were older than 18 years and required ventilation due to severe respiratory failure caused  
61 by SARS-CoV-19.

### 62 2.1. Technical procedure

63 Bed-side LUS at the intensive care unit was performed by an experienced physician of the interdis-  
64 ciplinary ultrasound centre of the university hospital of Regensburg. A hygienic concept, describing how  
65 to handle the mobile ultrasound device was defined and approved by the hospitals hygienic staff. Fun-  
66 damental B-mode LUS was performed using a LOGIQ S8 device from General Electrics® (convex 1–5  
67 MHz array probe) at a bed side setting on intensive care units. The images were saved as DICOM (digi-  
68 tal imaging and communications in the medicine) files and were imported to the local picture archiving  
69 and communication system (PACS, Siemens) for further evaluation. Pictograms were retrospectively  
70 analysed on a workstation by one radiologist and one gastroenterologist and grade of alteration was  
71 defined in consensus. Twelve-area-examination of the thorax was done, as fare as accessible, by divid-  
72 ing the two lobes of the lung in 6 areas each, including the anterior parasternal upper area (A up),  
73 the anterior parasternal lower area (A low), the para-axillary upper area (anterior lateral / AL up), the

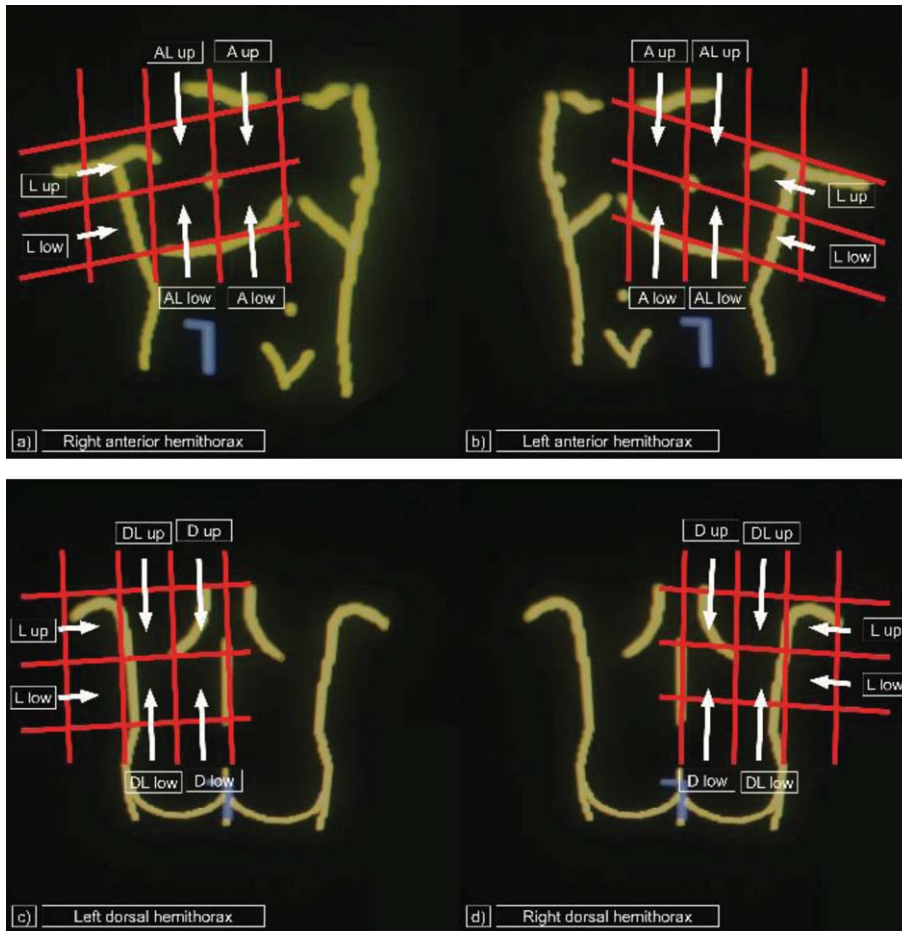


Fig. 1. Areas examined by lung ultrasound. Twelve-area-examination with a convex 1-5 MHz array probe by bed side point-of-care lung ultrasound. a) Right anterior hemithorax: A up = anterior parasternal upper area, A low = anterior parasternal lower area, AL up = anterior lateral / para-axillary upper area, AL low = AL up = anterior lateral / para-axillary upper area, L up = lateral upper area, L low = lateral lower area. b) Left anterior hemithorax: A up = anterior parasternal upper area, A low = anterior parasternal lower area, AL up = anterior lateral / para-axillary upper area, AL up = anterior lateral / para-axillary upper area, L up = lateral upper area, L low = lateral lower area. c) Left dorsal hemithorax: D up = dorsal posterior paravertebral upper area, D low = dorsal posterior paravertebral lower area, DL up = dorsal lateral / dorsal para-axillary upper area, DL low = dorsal lateral / dorsal para-axillary lower area, L up = lateral upper area, L low = lateral lower area. d) Right dorsal hemithorax: D up = dorsal posterior paravertebral upper area, D low = dorsal posterior paravertebral lower area, DL up = dorsal lateral / dorsal para-axillary upper area, DL low = dorsal lateral / dorsal para-axillary lower area, L up = lateral upper area, L low = lateral lower area.

74 para-axillary lower area (AL low) as well as the lateral upper area (L up) and the lateral lower area (L  
 75 low) in patients at supine position. In patients at prone position, lateral areas were defined as above. The  
 76 following dorsal areas were examined for the right and left lobe each: the dorsal posterior paravertebral  
 77 upper area (D up), the dorsal posterior paravertebral lower area (D low), the dorsal para-axillary upper  
 78 area (dorsal lateral / DL up) and the dorsal para-axillary lower area (DL low) (Fig. 1).

## 79 2.2. Lung aeration score

80 LUS findings described by Huang et al. [10] and Peng et al. [11] discriminated different findings,  
 81 describing focal, multifocal and confluent B-lines, pleural line irregularity, and consolidations. We

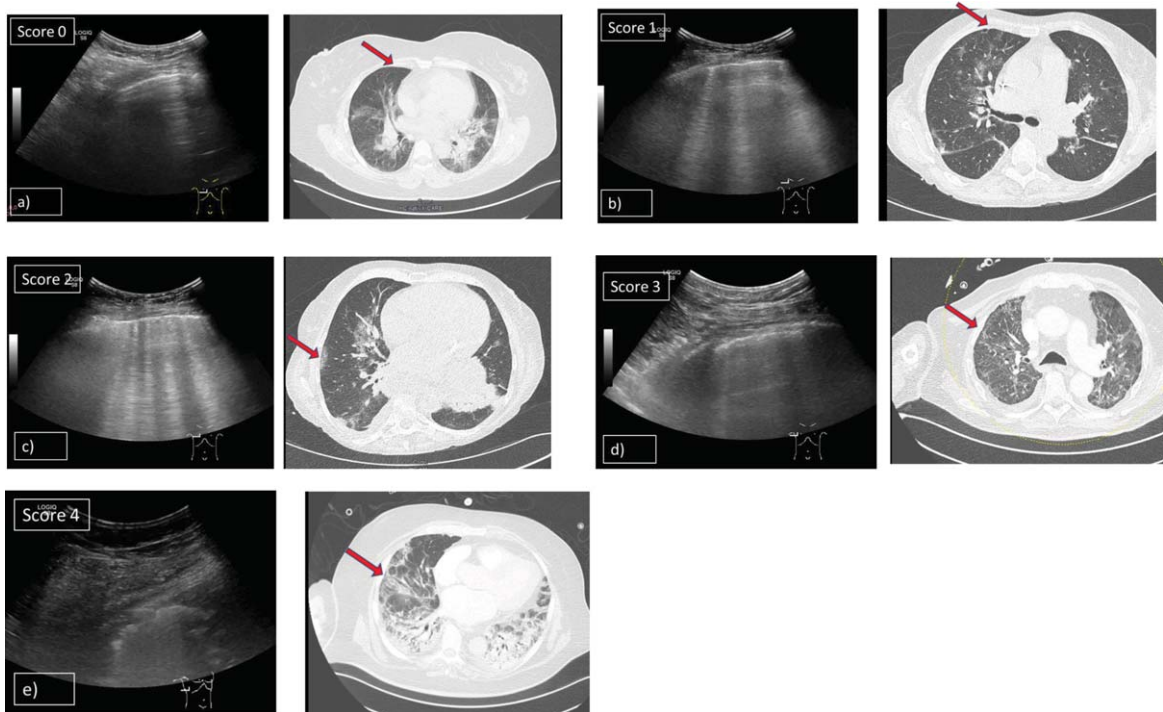


Fig. 2. Grading system of lung opacities used for the lung aeration score in patients with ARDS and COVID-19 pneumonia. Shows findings of LUS in patients with COVID-19 pneumonia and corresponding HRCT scans. 5 levels of typical opacities of the lung could be observed and were graded in a scoring system from minimum 0 points to maximum 4 points. a) Score 0: A-lines detected by ultrasound B-mode using a 1–5 MHz convex probe and corresponding HRCT scan (CT was done 3 days before LUS). b) Score 1: More than 2 ( $\geq 3$ ) single B-lines detected by ultrasound B-mode using a 1–5 MHz convex probe and corresponding HRCT scan (CT was done 3 days before LUS). c) Score 2: Multiple, coalescent B-lines detected by ultrasound B-mode using a 1–5 MHz convex probe and corresponding HRCT scan (CT was done 4 days before LUS). d) Score 3: Marked pleural disruptions detected by ultrasound B-mode using a 1–5 MHz convex probe and corresponding HRCT scan (CT was done at the same day as LUS). e) Score 4: Severe consolidations with air bronchogram sign detected by ultrasound B-mode using a 1–5 MHz convex probe and corresponding HRCT scan (CT was done at the same day as LUS).

82 graded typical findings of COVID-19 in five levels: 0 points were given if A-lines were present,  
 83 1 point if there were more than 2 ( $\geq 3$ ) single B-lines, 2 points if there were waterfall B-lines or  
 84 coalescent B-lines, 3 points if there were marked pleural disruptions and 4 points if consolidations  
 85 with air bronchogram sign were apparent (Fig. 2). Assessment of a lung aeration score (LAS) was done  
 86 by calculating the arithmetic mean of the grades defined of all examined areas of the lung (maximum  
 87 12 areas). When LUS was performed we retrospectively graded each examined area of a patient's lung  
 88 and gave 0 to 4 points for each area as shown in Fig. 2. The sum of all points was divided through the  
 89 number of examined areas. This calculation resulted in an arithmetic mean which was defined as the  
 90 patient's LAS.

### 91 2.3. Clinical parameters

92 Demographics, ventilatory parameters, parameters of blood gas analysis and laboratory markers  
 93 were obtained from the local electronic database (SAP® and MetaVision®). Time interval between  
 94 LUS and ventilatory parameters and chemistries were at maximum 6 hours, 12 hours respectively.  
 95 The following parameters defining ventilation were obtained by results of blood gas analysis: PaCO<sub>2</sub>

Table 1  
 Characteristics of patients with COVID-19 pneumonia who had lung ultrasound

Factor	N = 20	%
male	15	83.3
female	5	16.7
mean age (years)*	59,5 (36–68)*	
BMI	28.7 kg/m <sup>2</sup> (19.2–41.5)*	
Time from first diagnosis of COVID-19 to examination (days)	15.5 (1–52)*	
Supine position	13	72.2
Prone position	7	38.8
Parameters of Intensive care		
Pressure control ventilation	20	100
Diagnosis of ARDS	20	100
ECMO at timepoint of examination	5	25
Dialysis at timepoint of examination	7	35

\*median (range).

(mmHg), PaO<sub>2</sub> (mmHg), FiO<sub>2</sub> (%), Horovitz-Index (PaO<sub>2</sub>/FiO<sub>2</sub>) and oxygen saturation SO<sub>2</sub> (%). Positive end-expiratory pressure (PEEP) was given by automatic transfer by the ventilation device to the digital patient file (MetaVision®). C-reactive protein (CRP, mg/l) Lactate dehydrogenase (LDH, U/l), interleukin 6 (IL-6, pg/ml) and ferritin (µg/l) levels were obtained by the digital documentation system SAP®).

#### 2.4. Statistics

Statistical analyses were performed using Excel (Microsoft®, US) and SPSS (SPSS Statistics 25, IBM®, USA). Spearman rho test was calculated, comparing the value of the assessed lung aeration score with ventilation parameters and laboratory findings. A correlation coefficient  $r \geq 0,3$  was considered as a moderate correlation and  $r \geq 0,5$  as a strong correlation. *T*-Test was done to compare different groups. A *p* value < 0,05 was considered as significant. The study was conducted according to the declaration of Helsinki and was approved by the local ethical commission board in cooperation with the ethical board of the University of Tübingen (Number: 432/2020BO). The manuscript was written in accordance with the guidelines of Clinical Hemorheology and Microcirculation (Ethical guidelines for publication in Clinical Hemorheology and Microcirculation: Update 2016, Clin Hemorheol Microcirc. 2016;63(1):1-2.).

### 3. Results

Median body mass index (BMI) was  $28.6 \pm 5.58$  kg/m<sup>2</sup>, median time from first diagnosis of COVID-19 to LUS assessment was  $15 \pm 14.9$  days. In seven patients, LUS was performed in prone position, in 13 LUS was done in supine position. Patients were ventilated by pressure control. Five patients were treated with veno-venous extracorporeal membrane oxygenation (vvECMO) and seven patients required dialysis when LUS was performed. Patients characteristics are summarized in Table 1.

Results of the LAS are shown in Table 2. A maximum of 12 defined areas were examined (Fig. 1) and a grade ranging from 0 to 4 was classified as described at the section patients and methods (Fig. 2).

Table 2  
Lung aeration score in patients with COVID-19 pneumonia

a)	Supine position / segment												Lung aeration score Mean
	Anterior left lobe				Anterior right lobe				Lateral left lobe		Lateral right lobe		
	A <sub>1</sub> up	A low	AL <sub>2</sub> up	AL low	A up	A low	AL up	AL low	L <sub>3</sub> up	L low	L up	L low	
P001	1	3	2	2	1	2	4	4	4	2	4	4	2.75
P004	3	3	2	3	3	0	3	2	2	3	4	3	2.58
P005	2	3	4	3	3	4	4	2	4	4	4	4	3.42
P006	4	4	4	2	4	4	3	4	4	4	4	2	3.58
P007	3				3	4	2	3			1	3	2.38*
P008	2	3	3		3	2	4	4			4	4	3.22
P011	3	3	2		3	2	3	4	2	3	4	4	3.00
P013	4	3			4	3		4	3			4	3.57
P014	4	4		4	2	3	1	3	4		4		2.90
P015	4	4			3	4	4	4		4	3		3.75
P016			4		3	4					4		3.75*
P017	3	4		4	4	4	3	2			4		3.5
P018	4	4	4	4	4	4	3	2			4	4	3.82
Mean per area	3.08	3.45	3.13	3.14	3.08	3.08	3.09	3.17	3.29	3.33	3.67	3.56	3.42
b)	Prone position / segment												Mean
Patient	Dorsal left lobe				Dorsal right lobe				Lateral left lobe		Lateral right lobe		
	D <sub>4</sub> up	D low	DL <sub>5</sub> up	DL low	D up	D low	DL up	DL low	L up	L low	L up	L low	
P002	4	2	2	3	2	2	4	3	4	4		4	3.09
P003	4	3	1	2	4	3	2	2	2	4			2.7
P009	3	4	4	4	4	4	3	4	4	4	4	4	3.83
P010	4	2	4	4	4	2	3	2		4		3	3.2
P012	4	4	2	4	3	2	3	3		4	4		3.3
P019	3	2	2	3	1	4	3	3	2	4	2	4	2.75
P020	4	0	1	3	3	3	4	3	4	4	4	1	2.83
Mean per area	3.71	2.43	2.29	3.29	3.00	2.86	3.14	2.86	3.20	4.00	3.50	3.20	3.09

1: A = anterior area, 2: AL = anterior lateral area, 3: L = lateral area, 4: D = dorsal area, 5: DL = dorsal lateral area. Lung opacities were graded from 0 to 4. Lung aeration score was assessed by building the arithmetic mean of all examined areas. Results are shown for patients in supine position [a)] and patients in prone position [b)]. The mean of the grading for each examined area (mean per area) is given for patients examined in supine position and patients examined in prone position.

120 LAS was determined by calculating the arithmetic mean of all grades defined for each patient. In 13  
 121 out of 20 patients, LUS was performed in supine position, in 7 patients while being settled in prone  
 122 position. Notably, in 55% of patients at least one out of twelve areas of the lung, which have been  
 123 intended to be examined, could not be examined. This was mostly true for the left lateral segment when  
 124 patients were placed in lateral-decubitus position. Two patients placed in supine position had multiple  
 125 band-aids, which were necessary due to pressure ulcers after being placed in prone position. In these  
 126 two patients (Pat 007 and Pat 0016) only one area of the anterior left segment could be examined. For  
 127 this reason, these two patients were excluded from statistical analysis. Median LAS was 3.21 (Range  
 128 2.58 – 3.83); in patients in supine position median LAS was 3.42 (2.58 – 3.82), median LAS of patients

129 in prone position was 3.09 (2.58 – 3.83). Comparing the grading of the examined lung segments by  
130 arithmetic means showed highest levels in lateral lung segments. *T*-Test, comparing patients in supine  
131 and in prone position showed no significant difference ( $p=0.43$ ).

132 To analyse the correlation of clinical and laboratory parameters with the levels of LAS Pearson  
133 correlation was performed. LAS of each patient and ventilation parameters as well as laboratory  
134 findings, which were included in this comparison are shown in Table 3.

135 We choose CRP, LDH, IL-6 and ferritin for laboratory findings, because these parameters are associ-  
136 ated with bad prognosis of COVID-19 pneumonia [12, 13]. Median time of ventilation was 23.5 days.  
137 This study was performed as cross section study. Some patients were transferred to the university  
138 hospital of Regensburg to optimize ventilation or for ECMO treatment. These patients were already  
139 ventilated for a longer period of time in cooperating hospitals before. This implicates a wide range of  
140 days of ventilation at the timepoint when LUS was performed (1 – 52 day). 8 Patients were examined in  
141 the first 10 days of ventilation (mean 4 days) and 10 patients after more than 10 days (mean 20.5 days).  
142 5 patients were treated at ECMO when LUS was done. LAS was higher in the group of patients with  
143 ventilation time of more than 10 days (median LAS 2.9 vs 3.4;  $p=0.016$ ). Patients with or without need  
144 of ECMO-treatment had no difference in LAS ( $p=0.49$ ). Spearman rho correlation revealed moderate  
145 correlation of LAS with  $paO_2$  levels ( $r=0.336$ ;  $p=0.173$ ) and ferritin levels ( $r=0.467$ ;  $p=0.051$ ),  
146 however showing no significance.  $PaCO_2$  level was the only parameter showing a strong significant  
147 correlation to the LAS ( $r=0.632$ ;  $p=0.005$ ) in our cohort. Ventilation-time, Horovitz-index, PEEP,  
148  $SO_2$  and values of CRP, LDH and IL-6 showed no correlation with LAS (Table 4). Patients with a  
149 Horovitz-Index of  $< 150$  mmHg did not show any difference in LAS compared to  $> 150$  mmHg (*T*-Test,  
150  $p=0,495$ ).

#### 151 4. Discussion

152 Clinical outcome of patients with SARS-CoV-2 is often determined by pulmonary disease. Severe  
153 ARDS is reported in up to 15 to 31 % of patients with pneumonia [14, 15]. However, also beyond  
154 the massive aggression of the lung with severe pulmonary failure, as well as kidney and liver injuries,  
155 heart, brain, bowel and spleen damages with lymph nodes necrosis and even cutaneous manifestations  
156 have been observed [16]. Ultrasound diagnostic is a cost effective and world-wide accessible device  
157 to assess pneumonia [17, 18]. Especially in the COVID-19 pandemic it is proposed as a bed-side tool  
158 to minimize need for patient transport and therefore reduce exposure of medical staff and nosocomial  
159 transmission [19]. Typical findings have been described during the onset of the pandemic and include  
160  $\geq 3$  single B-lines, confluent / coalescent B-Lines and consolidations [20].

161 In regards of the technique used to assess lung opacities by LAS, Soldati et al. anticipated a “stan-  
162 dardization of the use of lung ultrasound for patients with COVID-19” in March 2020 and proposed  
163 to examine three areas on the back two lateral areas and two ventral areas [21]. The authors conceded  
164 that the posterior areas might be difficult to evaluate in supine position. However, as proposed by the  
165 German association for intensive care medicine (DGIIN), LUS is especially recommended in severely  
166 ill patients with ARDS [19]. In ventilated patients with ARDS it emerged in real life setting, that it is  
167 often only possible to assess either the ventral and lateral or the dorsal and lateral areas of the lung  
168 by point-of care bed side LUS, depending if the patient is placed in supine position or prone position.  
169 Therefore, we decided to examine either 4 ventral and 4 dorsal areas plus 2 lateral areas at each side  
170 of the thorax.

171 Before COVID-19 pandemic, the assessment of lung aeration by ultrasound has been used in patients  
172 with ARDS and a scoring system to quantify the grade of lung involvement in ARDS has been set up for  
173 these patients. Lung-collapse regions could be assessed by LUS in ARDS and influenced by alteration

Table 3  
Lung aeration score and clinical parameters of COVID-19 patients

Patient	Lung aeration score	Time of ventilation (days per patient)	ECMO	pCO2 (mmHg)	PaO2 (mmHG)	FiO2 (%)	Horovitz index	PEEP	SO2 (mmHg)	CRP (mg/l)	LDH (U/l)	IL-6 (pg/ml)	Ferritin (µg/l)
P001	2.75	20	no	38	76	45	170	10	96	126	232	665	733
P002	3.09	23	no	12	117	23	509	9	95	433	766	271	1986
P003	2.70	19	no	40	84	50	169	12	97	210	323	389	887
P004	2.58	12	no	30	65.9	98	150	8	89	151	396	179	686
P005	3.42	39	yes	53	86	55	123	12	97	102	339	385	726
P006	3.58	74	no	61	84	60	139	10	94	180	502	1822	615
P008	3.22	33	no	46	96	30	299	5	98	27	263	24	2745
P009	3.83	24	no	68	77	61	126	17	95	290	180	190	1245
P010	3.20	32	yes	40	64	35	184	13	93	202	656	278	2316
P011	3.00	25	yes	45	116	35	331	13	98	51	794	71	1918
P012	3.30	17	no	74	102	50	204	16	98	211	489	85	5666
P013	3.57	22	no	52	78	41	196	13	96	48	399	25	2367
P014	2.90	12	no	58	89	45	199	15	97	275	875	231	1593
P015	3.75	25	no	60	99	45	220	14	96	220	283	82	2560
P017	3.50	46	yes	45	88	35	253	10	98	201	444	2533	8621
P018	3.82	38	no	45	125	55	260	8	98	39	684	33	41174
P019	2.75	24	yes	35	104	40	299	14	98	283	325	347	1282
P020	2.83	16	no	48	77	60	129	14	91	129	354	129	799
Median <sup>1</sup>	3.21	23.5	yes = 5	46.8	88.7	45	197.5	12.7	96.5	180.5	375	210.5	1885
(Interquartile Range)	(2.58–3.82)	(10–74)		(12–74)	(64–125)	(23–98)	(123–509)	(5–17)	(87–98)	(27–433)	(180–875)	(24–2533)	(615–41174)

1: Median and range of each parameter is described and number of patients treated with extracorporeal membrane oxygenation (ECMO).



Table 4  
Spearman rho correlation of lung aeration score with parameters of ventilation and laboratory findings

Factor	n	Mean (range)	r-value	p-value
Ventilation				
Ventilation (days)	18	23.5 (10 – 74)	0.122	0.629
paO <sub>2</sub> (mmHg)	18	88.7 (64.4 – 125)	*0.336	0.173
paCO <sub>2</sub> (mmHg)	18	46.75 (12 – 74.3)	**0.632	0.005
Horovitz index (paO <sub>2</sub> /FiO <sub>2</sub> )	18	197.5 (123 – 509)	0.106	0.674
Laboratory findings				
CRP (mg/l)	18	180 (27.2 – 433)	–0.135	0.594
LDH (U/l)	18	375 (180 – 875)	0.016	0.951
IL-6 (pg/ml)	18	210.5 (23–2533)	–0.299	0.36
Ferritin (µg/l)	18	1885 (615 – 41174)	*0.467	0.051

\*moderate correlation. \*\*strong correlation.

of PEEP and position of the patient [22, 23]. A scoring system was applied by Soummer et al. and Algieri et al., which graded the severity of lung disease into four ultrasound findings describing normal A lines, non-coalescent (single) B-lines, coalescent B-lines and consolidations [24, 25]. Six predefined areas of the lung were assessed by the grading system and a cumulative score showed significant correlation to the CT scan and to the response to prone position [26]. In COVID-19 patients severely broken pleural lines is an additionally specific finding as described by Soldati et al. and Denault et al. [21, 27]. Soldati et al proposed 3 grades of severity in lung ultrasound in COVID-19 patients. A-lines giving 0 points, indented pleural lines giving 1 point, severely broken pleural lines with consolidations giving 2 points and a white lung with or without greater consolidations giving 3 points. However, this scoring System did include 30 COVID-19 patients of which an unknown quantity of cases was non-ventilated patients and the clinical condition of the patients was not correlated. However, our aim was to quantify lung aeration in severely ill patients in the style of prescribed lung aeration scores in patients with ARDS. For this reason, we decided to choose a scoring system according to the one proposed by Algieri et al. and to complement this system by the COVID-19 specific finding of broken pleural lines. In aspect of these findings we rated our scoring system, which describes an increase of lung opacities and assesses COVID-19 specific peripheral alterations of the lung. Our grading by the proposed LAS, confirms a CT scan analysis from Shi et al. [28], showing, that lower, lateral subpleural areas are more severely affected by COVID-19 pneumonia. In our findings the highest level of 4 points was reached in the lateral left lobe in patients which were settled in prone position. In supine position lateral areas had highest LAS, these are the best accessible dorsal-near points in patients settled in this position. These findings underline the accuracy of the proposed LAS.

ARDS in COVID-19 pneumonia isn't fully understand yet, however there have been described some differences compared to patients with non-COVID associated ARDS [29]. Berlin criteria defined ARDS onset as maximum 7 days after a respiratory insult [30]. However, the onset of ARDS occurs 8 to 12 days after the initial diagnosis of a COVID-19-associated pneumonia [1, 31, 32]. This specific feature of COVID-19 related ARDS is consistent with findings of LAS in our cohort, which was significantly higher in patients who have been ventilated for more than 10 days.

CO<sub>2</sub> retention is associated with bad prognosis in patients with severe COVID-19 pneumonia and need of ventilation or extracorporeal membrane oxygenation [33]. A possible explanation would be, that in COVID-19 associated ARDS alveolar endothelial cells are more affected than endothelial cells, causing a higher shunting of CO<sub>2</sub> rich blood [34]. Furthermore, alveolar capillary microthrombi could

205 decrease CO<sub>2</sub> diffusion and increase dead air volume [35]. Interestingly in our study oxygenation  
206 index showed no correlation with LAS and patients with a Horovitz-Index of <150 mmHg did neither  
207 show any difference in LAS. Notably, CO<sub>2</sub> measured in blood gas analysis at the day when LUS was  
208 performed, was the only ventilation associated parameter which showed significant correlation with  
209 the level of the proposed LAS. These findings might be associated with the distinct pathogenesis of  
210 ARDS in patients with COVID-19 pneumonia.

211 Biochemical parameters associated with poor outcome in COVID-19 pneumonia [9, 10] did not  
212 show any significant correlation with the LAS. However, ferritin levels tended to correlate with LAS.  
213 This observation suggests that the onset of inflammatory processes in COVID-19 pneumonia is not  
214 necessarily associated with the grade of lung opacities.

215 Several limitations of the study should be noted. The number of patients included in this single-centre  
216 cross-section study is limited. Ultrasound diagnostic was performed at different stages and phases of  
217 patients with severe COVID-19 pneumonia. The bedside LUS examination for Covid-19 pneumonia  
218 needs time to perform, experienced examiners and high-resolution probes, especially to detect the early  
219 peripheral pulmonary changes. LUS is very useful for the follow up in severe cases. Cineloops were  
220 only taken, if the examiner found frames not precise. Therefore, a prospective multicentre study with  
221 a follow-up examination of the lung by ultrasound is needed, to verify the precision of the proposed  
222 LAS and the correlation to clinical parameters and laboratory markers.

## 223 5. Conclusion

224 A lung aeration score, defining 4 grades of lung alterations in specified areas of the lung by bed side-  
225 ultrasound could be a useful tool to monitor lung disease and function in patients with severe COVID-19  
226 pneumonia. The proposed score shows significant correlation with CO<sub>2</sub> retention in patients with severe  
227 ARDS. However, these findings were collected in a cross-section analysis and accuracy of the score  
228 must be confirmed in a longitudinal prospective study.

## 229 References

- 230 [1] Wang DW, Hu C, Hu B, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected  
231 pneumonia in Wuhan, China. *JAMA*. 2020;323(11):1061-9.
- 232 [2] Yang X, Yu Y, Xu J, et al. Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan,  
233 China: A single centered, retrospective, observational study. *Lancet Respir Med*. 2020;8(5):475-81.
- 234 [3] Küpper JH, Jung F, Krieger V, Hufert F. A comparison of \*COVID\*-19\* mortality rates between European and Asian  
235 States. *Clin Hemorheol Microcirc*. 2020;75(1):3-5.
- 236 [4] Shi H, Han X, Jiang N, Cao Y, Alwalid O, Gu J, et al. Radiological findings from 81 patients with COVID-19 pneumonia  
237 in Wuhan, China: a descriptive study. *Lancet Infect Dis*. 2020;20(4):425-34.
- 238 [5] Schaible J, Meiler S, Poschenrieder F, Scharf G, Maintz D, Pregler B, Stroszczyński C, Hamer OW. Radiology of COVID-  
239 19 Pneumonia - Pictorial Essay and Structured Reporting. *Rofo*. 2020;192(6):513-7. doi: 10.1055/a-1164-7001. Epub  
240 2020 May 27
- 241 [6] Jung EM, Stroszczyński C, Jung F. Contrast enhanced ultrasound (CEUS) to assess pleural pulmonal changes in severe  
242 Covid-19 infection: First results. *Clin. Hemorheol. Microcirc*. 2020;75(1):19-26. doi: 10.3233/CH-209005
- 243 [7] Jung EM, Stroszczyński C, Jung F. Contrast enhanced ultrasonography (CEUS) to detect abdominal microcirculatory  
244 disorders in severe cases of COVID-19 infection: First experience. *Clin Hemorheol Microcirc*. 2020;74(4):353-61. doi:  
245 10.3233/CH-209003
- 246 [8] Sultan RL, Sehgal MC. A review of early experience in lung ultrasound (LUS) in the diagnosis and management of  
247 COVID-19. *Ultrasound Med Biol*. 2020;25:S0301-5629(20)30221-0. doi: 10.1016/j.ultrasmedbio.2020.05.012
- 248 [9] Bello G, Balnco P. Lung Ultrasonography for Assessing Lung Aeration in Acute Respiratory Distress Syndrome. *J*  
249 *Ultrasound Med*. 2019;0278-4297. doi: 10.1002/jum.14671

- 250 [10] Huang Y, Wang S, Liu Y, Zhang Y, Zheng C, Zheng Y, et al. A Preliminary Study on the Ultrasonic Manifestations  
251 of Peri pulmonary Lesions of Non-Critical Novel Coronavirus Pneumonia (COVID-19). SSRN Electron J 2020. doi:  
252 https://dx.doi.org/10.2139/ssrn.3544750
- 253 [11] Peng QY, Wang XT, Zhang LN. Findings of lung ultrasonography of novel corona virus pneumonia during the 2019–2020  
254 epidemic. *Intensive Care Med.* 2020;46(5):849-50.
- 255 [12] Velavan TP, Meyer GC. Mild Versus Severe COVID-19: Laboratory Markers. *Int J Infect Dis.* 2020;95:304-7.
- 256 [13] Henry MB, Santos de Oliveira MH, Benoit S, Plebani M, Lippi G. Hematologic, biochemical and immune biomarker  
257 abnormalities associated with severe illness and mortality in coronavirus disease 2019 (COVID-19): a meta-analysis.  
258 *Clin Chem Lab Med.* 2020;58(7):1021-8.
- 259 [14] Huang C, Wang Y, Li X, et al. Clinical features of patients with 2019 novel coronavirus in Wuhan, China. *Lancet.*  
260 2020;395(10223):497-506.
- 261 [15] Rodriguez-Morales AJ, Cardona-Ospina JA, Gutiérrez-Ocampo E, Villamizar-Peña R, Holguin-Riveraa Y, Escalera-  
262 Antezana JP, et al. Clinical, laboratory and imaging features of COVID-19: A Systematic Review and Meta-Analysis.  
263 *Travel Med Infect Dis.* 2020; 34:101623. doi: 10.1016/j.tmaid.2020.101623
- 264 [16] Martini R. The compelling arguments for the need of microvascular investigation in \*COVID\*-19\* critical patients.  
265 *Clin Hemorheol Microcirc.* 2020;75(1):27-34.
- 266 [17] Mayo PH, Copetti R, Feller-Kopman D, Mathis G, Maury E, Mongodi S, Mojoli F, Volpicelli G, Zanobetti M. Thoracic  
267 ultrasonography, A narrative review. *Intensive care medicine.* 2019;45:1200-11.
- 268 [18] Volpicelli G, Elbarbary M, Blaivas M, Lichtenstein DA, Mathis G, Kirkpatrick AW, Melniker L, et al. International  
269 evidence-based recommendations for point-of-care lung ultrasound. *Intensive Care Med.* 2019;45(9):1200-11.
- 270 [19] Kluge S, Janssens U, Welte T, Wber-Carstens S, Marcx G, Karaglannidis C. Empfehlungen zur intensivmedizinischen  
271 Therapie von Patienten mit COVID. *Dtsch Arztebl.* 2020;117(12): A-582 / B-503.
- 272 [20] Xing C, Li Q, Du H2 Kang W, Lian J, Yuan L. Lung ultrasound findings in patients with COVID-19 pneumonia. *Critical*  
273 *Care.* 2020;24:174. doi: 10.1186/s13054-020-02876-9
- 274 [21] Soldati G, Smargiassi A, Inchingolo R, Buonsenso D, Perrone T, Briganti DF, et al. Proposal for International Stan-  
275 dardization of the Use of Lung Ultrasound for Patients With COVID-19. *J Ultrasound Med.* 2020;39(7):1413-9.
- 276 [22] Stefanidis K, Dimopoulos S, Tripodaki ES, et al. Lung sonography and recruitment in patients with early acute respiratory  
277 distress syndrome: a pilot study. *CriticalCare.* 2011;15:R185. doi: 10.1186/cc10338
- 278 [23] Rode B, Vucic ´ M, Siranovic M, et al. Positive end-expiratory pressure lung recruitment: comparison between lower  
279 inflection point and ultrasound assessment. *Wien Klin Wochenschr.* 2012;124:842-7.
- 280 [24] Soummer A, Perbet S, Brisson H, et al. Ultrasound assessment of lung aeration loss during a successful weaning trial  
281 predicts postextubation distress. *Crit CareMed.* 2012;40:2064-72.
- 282 [25] Algieri I, Mongodi S, Chiumello D, et al. CT scan and ultrasound comparative assessment of PEEP-induced lung  
283 aeration changes in ARDS. *Crit Care.* 2014;18(suppl 1):P285. doi:10.1186/cc13475
- 284 [26] Prat G, Guinard S, Bizien N, et al. Can lung ultrasonography predict prone positioning response in acute respiratory  
285 distress syndrome patients? *J Crit Care.* 2016;32:36-41.
- 286 [27] Denault AY, Delisle S, Cauty D, Royse A, Royse C, Serra XC et al. A proposed lung ultrasound and phenotypic algorithm  
287 for the care of COVID-19 patients with acute respiratory failure. *Can J Anesth/J Can Anesth.* 2020;21:1-12.
- 288 [28] Shi H, Han X, Jiang N, Cao Y, Alwalid O, Gu J. Radiological findings from 81 patients with COVID-19 pneumonia in  
289 Wuhan, China: a descriptive study. *Lancet Infect Dis.* 2020;20(4):425-34.
- 290 [29] Xu L, Ma X. Acute respiratory failure in COVID-19: is it “typical” ARDS? *Critical Care.* 2020;24:198. doi:  
291 10.1186/s13054-020-02911-9
- 292 [30] Ranieri VM, Rubenfeld GD and the Definition Task Force ARDS. Acute respiratory distress syndrome: the Berlin  
293 definition. *JAMA.* 2012;307:2526-33.
- 294 [31] Guan WJ, Ni ZY, Hu Y, et al. Clinical characteristics of coronavirus disease 2019 in China. *N Engl J Med.*  
295 2020;382(18):1708-20.
- 296 [32] Zhou F, Yu T, Du R, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan,  
297 China: a retrospective cohort study. *Lancet.* 2020;395(10299):1054-62.
- 298 [33] Yang X, Cai S, Luo Y, Zhu F, Hu M, Zhao Y, et al. Extracorporeal Membrane Oxygenation for Coronavirus Disease  
299 2019-Induced Acute Respiratory Distress Syndrome: A Multicenter Descriptive Study. *Crit Care Med.* 2020. doi:  
300 10.1097/CCM.0000000000004447
- 301 [34] Jung F, Kruger-Genge A, Franke RP, Hufert F, Kupper J-H. COVID-19 and the endothelium. *Clin Hemorheol Microcirc.*  
302 2020;75(1):7-11. doi: 10.3233/CH-209007
- 303 [35] Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, et al. Pulmonary Vascular Endothelialitis,  
304 Thrombosis, and Angiogenesis in Covid-19. *N Engl J med.* 2020; doi: 10.1056/NEJMoa2015432