

## Research Report

# Synthesis, Characterization, and Biological Activity of Anthraquinone-Substituted Imidazolium Salts for the Treatment of Bladder Cancer

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### Abstract.

**BACKGROUND:** Bladder cancer is one of the most common types of cancer diagnosed each year, and more than half of patients have non-muscle invasive bladder cancer (NMIBC). The standard of care for patients with high-grade NMIBC is Bacillus Calmette-Guerin (BCG). Unfortunately, multiple BCG shortages have limited access to this treatment. Available alternatives using intravesical administration of chemotherapy have some efficacy, but lack prospective validation and long-term outcomes. Development of novel intravesical therapies may provide more active alternatives to BCG for patients with high-grade NMIBC.

**OBJECTIVE:** To develop an optimal imidazolium salt for the intravesical treatment of NMIBC and determine preliminary *in vitro* activity of anthraquinone-substituted imidazolium salts.

**METHODS:** The development of the anthraquinone-substituted imidazolium salts was undertaken in an attempt to increase the potency of this class of compounds by incorporating the quinone functional group observed in the chemotherapeutics doxorubicin, valrubicin, and mitomycin. All compounds were characterized by <sup>1</sup>H and <sup>13</sup>C NMR spectroscopy and infrared spectroscopy. Furthermore, these imidazolium salts were tested for *in vitro* cytotoxicity by the Developmental Therapeutics Program (DTP) on the NCI-60 human tumor cell line screening. Additional *in vitro* testing was performed against diverse bladder cancer cell lines (RT112, TCCSUP, J82, and UMUC13) using CellTiter-Glo® assays and colony-forming assays.

**RESULTS:** The NCI-60 cell line screening indicated that compound **7** had the highest activity and was concluded to be the optimal compound for further study. Using CellTiter-Glo® assays on bladder cancer cell lines, 50% growth inhibitory concentration (IC<sub>50</sub>) values were determined to range from 32–50 μM after an exposure of 1 h, for compound **7**. Further evaluation of the compound by colony-forming assays showed the complete inhibition of growth at 10 days post a 100 μM dose of compound **7** for 1 h.

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**CONCLUSIONS:** The most active lipophilic anthraquinone imidazolium salt, compound **7**, could be a viable treatment for non-muscle invasive bladder cancer as it exhibits a cell-killing effect at a 1 h time period and completely inhibits cancer regrowth in colony-forming assays.

Keywords: Anticancer, antitumor, anthraquinone, imidazolium salts, non-muscle invasive bladder cancer

## INTRODUCTION

Bladder cancer is estimated to affect over 80,000 new patients nationally in 2020, with more than 17,000 deaths anticipated [1]. Additionally, bladder cancer is the 4th and 8th most prevalent cancer in men and women, respectively, with the most common stage at the time of diagnosis being non-muscle invasive bladder cancer (NMIBC) [2]. Many therapies are used for NMIBC, but few are effective at elimination of the disease in some patients. The current standard of care for NMIBC is intravesical treatment with Bacillus Calmette-Guerin (BCG). Although BCG is an effective therapy, it is currently underproduced, which puts many patients at risk for cancer recurrence, progression, and subsequent radical cystectomy [3, 4]. Immunotherapy isn't the only option as intravesical chemotherapy can be used as a treatment for NMIBC. A problem with intravesical chemotherapy is that the treatment has been shown to lead to 62% recurrence in intermediate grade patients within 5 years, according to a study performed by the European Association of Urology [5, 6]. This may be circumvented with a treatment regimen of gemcitabine/docetaxel that has shown a reduction of recurrence, but lacks the evaluation of long term outcomes and prospective validation [7]. Furthermore, there are more difficult cases to treat with high-grade NMIBC such as carcinoma *in situ* (CIS), which have shown cancer progression when treated with intravesical chemotherapy [8]. We presume that one of the reasons for the reduced efficacy of intravesical chemotherapy is that the activity of the chemotherapeutic used is dependent on prolonged exposure in order to be incorporated into the cell in S-phase (during DNA replication) or mitosis, when most chemotherapies exert their toxicity. Prolonged exposure requires longer intravesical retention of the chemotherapy to achieve a therapeutic effect, which is not practical for most patients.

Due to the numerous BCG shortages and the reduced efficacy of intravesical chemotherapy compared to BCG, novel chemotherapeutics are needed to treat bladder cancer. Development of novel anticancer

materials is a necessity to combat bladder cancer and improve patient outcomes. We recently developed an imidazolium salt (TPP1) that selectively targets and is cytotoxic toward bladder cancer *in vivo* [9]. This compound has somewhat low potency, with 500–1000  $\mu\text{M}$  concentrations needed to treat select bladder cancer models, although this is still easily achievable in an intravesical setting (for reference, gemcitabine is often given at a concentration of 2.00 g in 50.0 mL, or 152 mM). To increase the potency of imidazolium salts for the treatment of bladder cancer, quinone moieties (which are key functional groups in the highly cytotoxic chemotherapeutics doxorubicin, valrubicin, and mitomycin C (Fig. 1)) can be incorporated. Therefore, we present the synthesis and characterization of novel imidazolium salts containing anthraquinone moieties and their *in vitro* activity against bladder cancer.

## MATERIALS AND METHODS

### General considerations

All reactions were performed aerobically unless otherwise stated. Solvents and chemical reagents were unmodified and purchased from VWR, Fisher Scientific, or Sigma Aldrich. The synthesis of starting material **1** was performed by a modified literature procedure [10, 11].  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were acquired on an Inova 400 MHz instrument ( $^{13}\text{C}$  NMR 100 MHz). All NMR samples were prepared using DMSO- $d_6$  and spectra were referenced to DMSO- $d_6$  ( $^1\text{H}$ : 2.500 ppm,  $^{13}\text{C}$ : 39.520 ppm). Infrared spectroscopy was performed on imidazolium salts **3**, **4**, **6**, and **7** using a Thermo Scientific iS5 FT-IR w/ iD7 attachment.

### X-ray crystallographic analysis

Crystals of the compounds were coated in Paratone oil, mounted on a CryoLoop and placed on a goniometer under a stream of nitrogen. Crystal structure data sets were collected on a Bruker Kappa APEX II Duo CCD system equipped with a Mo I  $\mu\text{S}$

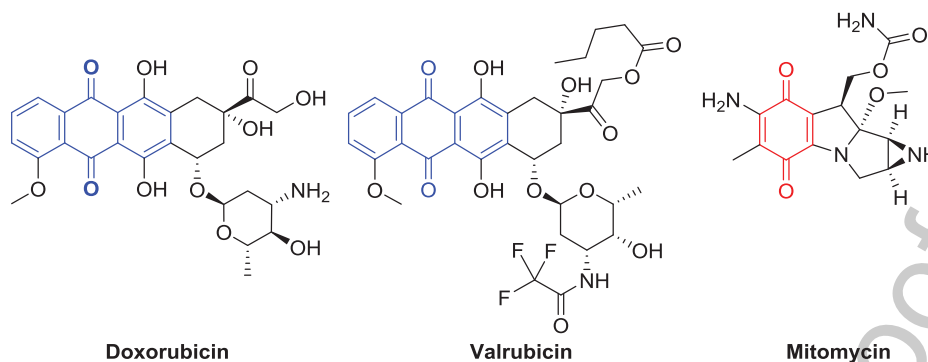


Fig. 1. Anthraquinone and quinone-containing chemotherapeutics, with anthraquinone (3-ring system) moieties highlighted in blue and quinone moieties (the basic functional group of a single conjugated cyclic dione) highlighted in red. Structures of doxorubicin, valrubicin, and mitomycin are shown.

115 source and a Cu I  $\mu$ S micro-focus source equipped  
116 with QUAZAR optics ( $\lambda = 1.54178 \text{ \AA}$ ). The unit cells  
117 were determined by using reflections from three dif-  
118 ferent orientations. Data sets were collected using  
119 APEX II software packages. All data sets were pro-  
120 cessed using the APEX II software suite [12, 13].  
121 The data sets were integrated using SAINT [14].  
122 An empirical absorption correction and other correc-  
123 tions were applied to the data sets using multi-scan  
124 SADABS [15]. Structure solution, refinement, and  
125 modelling were accomplished by using the Bruker  
126 SHELXTL package [16]. The structures were deter-  
127 mined by full-matrix least-squares refinement of  $F^2$   
128 and the selection of the appropriate atoms from  
129 the generated difference map. Hydrogen atom posi-  
130 tions were calculated and  $U_{\text{iso}}(\text{H})$  values were fixed  
131 according to a riding model.

#### 132 DTP NCI-60 cell line screening

133 The developmental therapeutics program (DTP)  
134 treatment protocol is summarized as follows: 60  
135 cell lines ranging from leukemia to breast cancer  
136 are treated with a  $10 \mu\text{M}$  concentration of the  
137 compound. The results are presented as growth  
138 percent. Continued growth is quantified from 0 to  
139 100 and lethality is scored from  $-100$  to 0. Further  
140 information on the experimental procedure for  
141 cell treatment can be viewed at NCI-DTP website  
142 ([https://dtp.cancer.gov/discovery\\_development/nci-60/methodology.htm](https://dtp.cancer.gov/discovery_development/nci-60/methodology.htm)).  
143

#### 144 CellTiter-Glo<sup>®</sup> assay

145 Bladder cancer cell lines (RT112, TCCSUP, J82,  
146 and UMUC13) were seeded at 10,000 cells/well

147 in black-walled 96 well plates and incubated for  
148 24 h. Cells were treated with compound 7 in their  
149 respective media (as recommended by ATCC) at con-  
150 centrations of 500 to  $7.8 \mu\text{M}$  dissolved in two-fold  
151 dilution increments of vehicle for 1 h, followed by  
152 replacement of treatment media with normal growth  
153 media. After 24 h, plates with cells were incubated  
154 at room temperature for 30 min, followed by the  
155 addition of  $20 \mu\text{L}$  of CellTiter-Glo<sup>®</sup> luminescent  
156 cell viability assay reagent (Promega) and mixing.  
157 After 2 min of incubation at room temperature, lumi-  
158 nescence for each plate was measured using IVIS.  
159 Relative survival for each treatment group was cal-  
160 culated using vehicle control. All treatments were  
161 performed in quadruplicate.

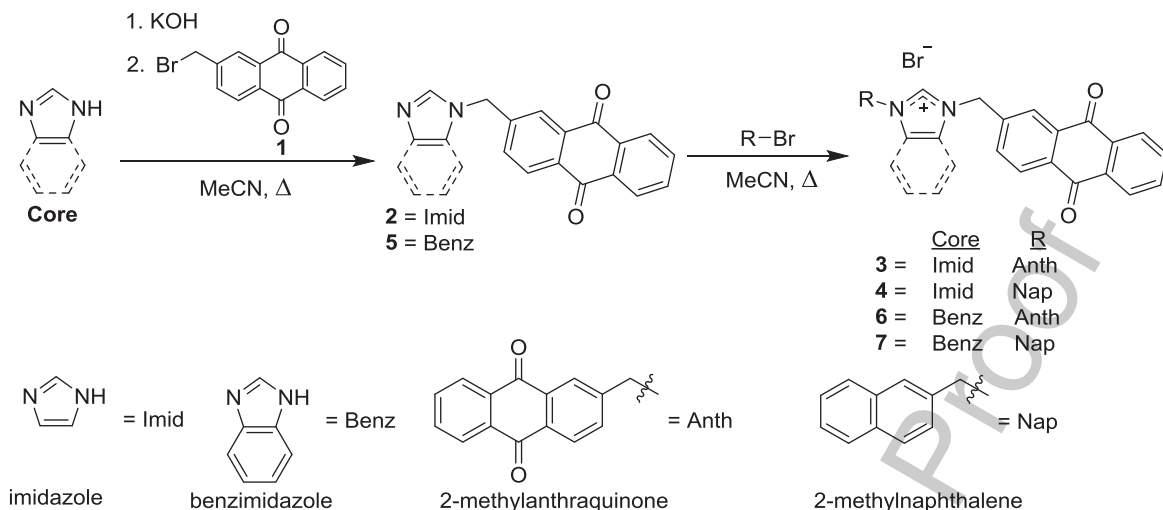
#### 162 Colony-forming assay

163 Cells were seeded at 500 cells/well in 6-well plates  
164 and 24 h later treated with  $25 \mu\text{M}$ ,  $50 \mu\text{M}$  or  $100 \mu\text{M}$   
165 of compound 7 or vehicle for 1 h. Cells were then  
166 allowed to grow in normal media for 10 days followed  
167 by staining with crystal violet (0.5% crystal violet in  
168 20% MeOH/water) and incubation at  $4^\circ\text{C}$  for 5 min.  
169 The wells were then washed several times with PBS  
170 and dried prior to imaging. All the experiments were  
171 performed in triplicate.

#### 172 Synthesis of 2-(bromomethyl)-9,10- 173 anthraquinone (1)

174 2-(Bromomethyl)-9,10-anthraquinone (compound  
175 1) was synthesized by a modified literature procedure  
176 [10, 11]. The radical initiator used was azobi-  
177 sisobutyronitrile (AIBN) instead of benzoyl peroxide  
178 (BPO). To obtain adequate purification for further  
179

179	reactions the product was collected from the hot reaction	( $I > 2\sigma(I)$ ). The final R1 values were 0.1276 (all data).	226
180	mixture by vacuum filtration and subsequently	The final $wR(F^2)$ values were 0.1331 (all data).	227
181	recrystallized with ethanol.		
182	<i>Synthesis of 2-((imidazol-1-yl)methyl)-9,10-</i>	<i>Synthesis of 1-((9,10-anthraquinone-2-yl)</i>	228
183	<i>anthraquinone (2)</i>	<i>methyl)-3-(naphthalen-2-ylmethyl)imidazolium</i>	229
		<i>bromide (4)</i>	230
184	Imidazole (0.308 g, 4.52 mmol) was stirred at	Compound <b>2</b> (0.283 g, 0.981 mmol) was refluxed	231
185	reflux in acetonitrile (10 mL) with potassium hydroxide	in acetonitrile (2 mL) with 2-(bromomethyl)naphthalene	232
186	(0.281 g, 5.01 mmol). After 15 minutes compound	(0.225 g, 1.02 mmol) for 15 h. A yellow precipitate	233
187	<b>1</b> (1.361 g, 4.52 mmol) was added and the reaction	formed and was collected via vacuum filtration	234
188	mixture was allowed to reflux for 24 h. The reaction	(0.164 g, 32.8%). $^1\text{H}$ NMR (400 MHz DMSO- $d_6$ )	235
189	was filtered hot to remove KBr and the filtrate	$\delta = 9.56$ (1H, s, NCHN) 8.25 (4H, m, Ar) 7.98 (9H,	236
190	volatiles were removed under reduced pressure. The	m, Ar) 7.56 (3H, m, Ar) 5.71 (2H, s, CH <sub>2</sub> ) 5.65 (2H,	237
191	resulting crude solid was stirred in hot water. The	s, CH <sub>2</sub> ). $^{13}\text{C}$ NMR (100 MHz DMSO- $d_6$ ): $\delta$ 182.0,	238
192	product was collected via vacuum filtration, washed	181.9, 141.2, 136.9, 134.63, 134.58, 134.1, 133.3,	239
193	with ethyl ether, and dried under vacuum. The solid	132.9, 132.84, 132.82, 132.7, 132.6, 132.0, 128.8,	240
194	was dissolved in dichloromethane and impurities	127.8, 127.7, 127.6, 127.5, 126.7, 126.6, 126.5,	241
195	were precipitated with ethyl ether. Upon filtration,	125.7, 123.2, 123.0, 52.3, 51.4. ATR-IR: 3140w (CH,	242
196	the filtrate volatiles were removed affording com-	sp <sup>2</sup> ), 3009w (CH, sp <sup>2</sup> ), 2968w (CH, sp <sup>3</sup> ), 1672s	243
197	compound <b>2</b> as a yellow solid (0.963 g, 73.9%). $^1\text{H}$ -NMR	(C=O), 1592 m (C=C) cm <sup>-1</sup> .	244
198	(400 MHz DMSO- $d_6$ ): $\delta$ 8.18 (m, 2H, Ar), 8.08 (d,		
199	1H, Ar), 8.00 (s, 1H, Ar), 7.91 (dd, 2H, Ar), 7.83 (s,	<i>Synthesis of 2-((benzimidazol-1-yl)methyl)-</i>	245
200	1H, Ar), 7.72 (m, 1H, Ar), 7.26 (s, 1H, Ar), 6.96 (s,	<i>9,10-anthraquinone (5)</i>	246
201	1H, Ar), 5.45 (s, 2H, CH <sub>2</sub> ).		
202	<i>Synthesis of 1,3-bis((9,10-anthraquinone-2-yl)</i>	Benzimidazole (0.506 g, 4.52 mmol) was stirred	247
203	<i>methyl)imidazolium bromide (3)</i>	at reflux in acetonitrile (10 mL) with potassium	248
204		hydroxide (0.273 g, 5.01 mmol). After 30 minutes,	249
205	Compound <b>2</b> (0.499 g, 1.73 mmol) was refluxed	<b>1</b> (1.271 g, 4.52 mmol) was added and the reaction	250
206	in acetonitrile (8 mL) with compound <b>1</b> (0.535 g,	mixture was allowed to reflux for 24 h. The reaction	251
207	1.78 mmol) for 15 h. A yellow precipitate formed and	mixture was filtered hot to remove KBr and the fil-	252
208	was collected via vacuum filtration. The crude prod-	trate volatiles were removed under reduced pressure.	253
209	uct was stirred in refluxing acetonitrile, collected via	The resulting crude solid was stirred in hot water.	254
210	vacuum filtration and washed with THF and ethyl	Compound <b>5</b> was collected via vacuum filtration and	255
211	ether to produce compound <b>3</b> (0.609 g, 59.7%). $^1\text{H}$ -	washed with ethyl ether (0.520 g, 73.9%). $^1\text{H}$ -NMR	256
212	NMR (400 MHz DMSO- $d_6$ ): $\delta$ 9.61 (s, 1H, NCHN),	(400 MHz; DMSO- $d_6$ ): $\delta$ 8.50 (s, 1H, Ar), 8.18 (m,	257
213	8.26 (d, 2H, Ar), 8.21 (m, 4H, Ar), 8.15 (m, 2H, Ar),	3H, Ar), 8.07 (s, 1H, Ar), 7.89 (m, 2H, Ar), 7.79 (d,	258
214	7.99 (s, 3H), 7.97 (m, 5H, Ar), 5.73 (s, 4H, CH <sub>2</sub> ).	1H, Ar), 7.70 (m, 1H, Ar), 7.54 (m, 1H, Ar), 7.22 (m,	259
215	$^{13}\text{C}$ NMR (100 MHz DMSO- $d_6$ ): $\delta$ 181.9, 141.1,	2H, Ar), 5.77 (s, 2H, CH <sub>2</sub> ).	260
216	137.2, 134.6, 134.5, 134.1, 133.3, 132.9, 132.8,	<i>Synthesis of 1,3-bis((9,10-anthraquinone-2-yl)</i>	261
217	127.6, 126.7, 126.5, 123.3, 51.5. ATR-IR: 3068w	<i>methyl)benzimidazolium bromide (6)</i>	262
218	(CH, sp <sup>2</sup> ), 3006w (CH, sp <sup>2</sup> ), 1674s (C=O), 1590m		
219	(C=C) cm <sup>-1</sup> .	Compound <b>5</b> (0.502, 1.73 mmol) was refluxed	263
220	Crystal data for <b>3</b> : C <sub>33</sub> H <sub>21</sub> BrN <sub>2</sub> O <sub>4</sub> , M = 589.43,	in acetonitrile (5 mL) with compound <b>1</b> (0.456 g,	264
221	monoclinic, $a = 11.5769(8)$ Å, $b = 11.9943(8)$ Å,	1.78 mmol) for 16 h. A tan precipitate formed and	265
222	$c = 36.529(2)$ Å, $\beta = 98.832(4)^\circ$ , $V = 5012.2(6)$ Å <sup>3</sup> ,	was collected via vacuum filtration. The crude prod-	266
223	$T = 100(2)$ K, space group C2/c, $Z = 8$ , 21708	uct was dissolved in DMF and precipitated with ethyl	267
224	reflections measured, 5086 independent reflections	ether to afford compound <b>6</b> (0.752 g, 68.1%) $^1\text{H}$ -	268
225	( $R_{\text{int}} = 0.0894$ ). The final R1 values were 0.0594	NMR (400 MHz DMSO- $d_6$ ): $\delta$ 9.61 (s, 1H, NCHN),	269
	( $I > 2\sigma(I)$ ). The final $wR(F^2)$ values were 0.1074	8.26 (d, 2H, Ar), 8.21 (m, 4H, Ar), 8.15 (m, 2H,	270
		Ar), 7.99 (s, 3H), 7.97–7.92 (m, 5H), 5.73 (s, 4H).	271



Scheme 1. Syntheses of compounds 2–7 by alkylation of imidazole or benzimidazole with 1 or 2-bromomethylnaphthalene.

272 <sup>13</sup>C NMR (100 MHz DMSO-d<sub>6</sub>): δ 182.0, 181.9,  
 273 143.7, 140.4, 134.6, 134.5, 134.0, 133.4, 132.9,  
 274 132.8, 131.1, 127.5, 127.1, 126.7, 126.5, 114.1, 49.6.  
 275 ATR-IR: 3133w (CH, sp<sup>2</sup>), 3071w (CH, sp<sup>2</sup>), 3015w  
 276 (CH, sp<sup>2</sup>), 2928w (CH, sp<sup>3</sup>), 1668s (C=O), 1603 m  
 277 (C=C) cm<sup>-1</sup>.

278 *Synthesis of 1-((9,10-anthraquinone-2-yl)*  
 279 *methyl)-3-(naphthalen-2-ylmethyl)*  
 280 *benzimidazolium bromide (7)*

281 Compound **5** (0.630, 1.86 mmol) was refluxed  
 282 in acetonitrile (5 mL) with 2-(bromomethyl)naphtha  
 283 lene (0.428 g, 1.94 mmol) for 16 h. A tan precipitate  
 284 formed and was collected via vacuum filtration. The  
 285 crude product was triturated with dichloromethane to  
 286 afford an off-white solid, compound **7** (0.643 g, 61.7  
 287 %) <sup>1</sup>H-NMR (400 MHz DMSO-d<sub>6</sub>): δ 10.24 (s, 1H,  
 288 NCHN), 8.37 (s, 1H, Ar), 8.23 (m, 3H, Ar), 8.15 (s,  
 289 1H), 8.03 (dd, 4H, Ar), 7.94 (m, 4H, Ar), 7.66 (d,  
 290 3H, Ar), 7.57 (m, 2H, Ar), 6.10 (s, 2H, CH<sub>2</sub>), 6.02 (s,  
 291 2H, CH<sub>2</sub>). <sup>13</sup>C NMR (100 MHz DMSO-d<sub>6</sub>): δ 182.5,  
 292 182.4, 143.8, 141.0, 135.06, 135.02, 134.4, 133.8,  
 293 133.3, 133.28, 133.24, 133.2, 133.1, 131.7, 131.6,  
 294 131.5, 129.2, 128.3, 128.2, 128.1, 127.9, 127.4,  
 295 127.3, 127.2, 127.1, 126.9, 126.2, 114.7, 114.4, 50.8,  
 296 50.0. ATR-IR: 3129w (CH, sp<sup>2</sup>), 3069w (CH, sp<sup>2</sup>),  
 297 3025w (CH, sp<sup>2</sup>), 2952w (CH, sp<sup>3</sup>), 1668s (C=O),  
 298 1591 m (C=C) cm<sup>-1</sup>.

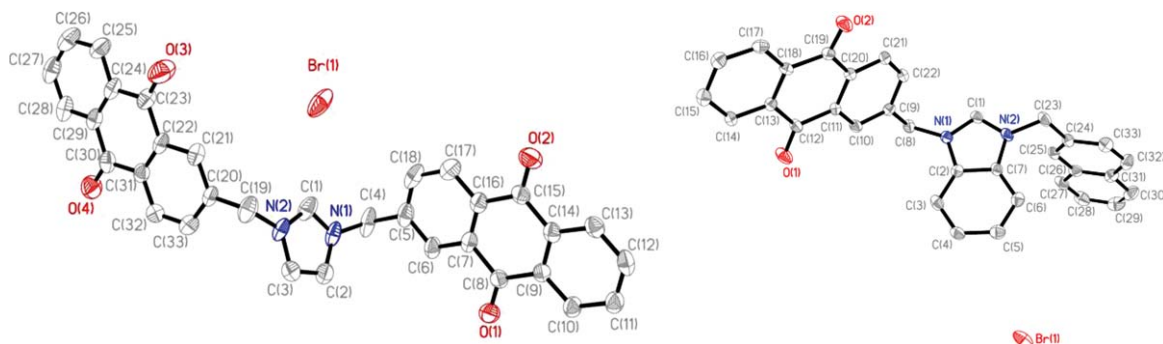
299 Crystal data for compound **7**: C<sub>33</sub>H<sub>23</sub>BrN<sub>2</sub>O<sub>2</sub>,  
 300 M = 559.44, Triclinic, *a* = 7.9808(4) Å, *b* = 11.3358  
 301 (5) Å, *c* = 15.2282(8) Å, α = 74.908(3)°, β = 86.015  
 302 (3)°, γ = 70.555(3)° V = 5012.2(6) Å<sup>3</sup>, T = 100(2)

K, space group P-1, Z = 2, 20211 reflections mea-  
 303 sured, 5073 independent reflections (R<sub>int</sub> = 0.0640).  
 304 The final R1 values were 0.0601 (I > 2σ(I)). The final  
 305 wR(F<sup>2</sup>) values were 0.1359 (I > 2σ(I)). The final R1  
 306 values were 0.0854 (all data). The final wR(F<sup>2</sup>) values  
 307 were 0.1538 (all data).  
 308

## 309 RESULTS AND DISCUSSION

### 310 Synthesis and characterization

311 The syntheses of the anthraquinone-substituted  
 312 imidazolium salt compounds **3**, **4**, **6**, and **7** are  
 313 outlined in the general reactions of **Scheme 1**.  
 314 Compounds **3** and **4** were first synthesized by depro-  
 315 tonation of imidazole with potassium hydroxide  
 316 and then adding 2-(bromomethyl)anthracene-9,10-  
 317 dione. The purified mono-substituted imidazole  
 318 (compound **2**) was subsequently combined with  
 319 either 2-(bromomethyl)anthracene-9,10-dione (com-  
 320 pound **1**) or 2-(bromomethyl)naphthalene to form  
 321 compounds **3** and **4** respectively. The compounds  
 322 were first evaluated by <sup>1</sup>H NMR spectroscopy and the  
 323 characteristic C<sup>2</sup> proton resonances were observed at  
 324 9.61 ppm for **3** and 9.57 ppm for **4**. Additionally a  
 325 single resonance at 5.73 ppm was observed for the  
 326 methylene protons of the symmetric compound **3**.  
 327 In contrast the asymmetric imidazolium salt (com-  
 328 pound **4**) yielded the methylene resonances at 5.71  
 329 and 5.65 ppm. Further characterization by <sup>13</sup>C NMR  
 330 spectroscopy yielded the C<sup>2</sup> carbon resonance at  
 331 141 ppm for both compounds. The inequivalent car-

Fig. 2. Thermal ellipsoid plots of **4** and **7** with hydrogen atoms not shown for clarity. Thermal ellipsoids were drawn to 50% probability.Table 1  
One Dose NCI-60 Cell Line Screening for Compounds 3,4,6, and 7 (GI%)

Compound	CELL LINE	Leukemia										SR	NSCLC									
		Leukemia	CCR6-CEM	HL-60(TB)	K-562	MOLT-4	RPMI-8226	A549/ATCC	EKVX	HOP-62	HOP-92		NCI-H226	NCI-H23	NCI-H322M	NCI-H460	NCI-H522					
3		90.0	97.2	92.2	98.7	91.2	48.5	92.9	98.5	96.5	122.2	77.8	90.9	89.3	88.4	103.3						
4		87.0	100.2	98.6	97.0	88.3	48.1	91.2	109.2	94.4	117.2	88.8	92.9	87.7	92.9	107.9						
6		71.0	43.0	32.5	79.1	17.3	73.7	110.2	70.9	106.9	47.8	83.3	75.6	91.0	94.0	38.9						
7		22.1	-49.2	17.6	28.1	2.5	38.1	91.0	73.9	76.2	15.6	63.9	56.5	75.4	63.4	34.1						

Compound	CELL LINE	Breast						CNS	Prostate						
		MC7	MDA-MB231/ATCC	HS 578T	BT-549	T-47D	MD A-MB-468		SF-268	SF-295	SF-539	SNB-19	SNB-75	U251	PC-3
3		83.8	93.5	101.2	103.1	81.2	43.2	100.0	104.0	102.3	99.0	103.7	118.1	88.5	102.3
4		92.9	100.6	103.6	101.8	90.8	71.2	98.5	102.4	98.0	92.4	96.6	103.1	83.7	104.6
6		59.2	81.4	92.6	60.1	70.1	-32.2	72.5	94.9	84.1	79.5	62.8	85.8	36.8	114.6
7		39.9	26.2	62.2	47.0	41.2	-27.6	49.3	82.2	70.6	45.0	42.3	57.1	27.7	83.1

Compound	CELL LINE	Colon						Renal	Ovarian							
		COLO 205	HCC-2998	HCT-116	HCT-15	HT29	KM12		SW620	766-0	A498	ACHN	C AKI-1	RXF 393	SN12C	TK-10
3		97.7	100.3	92.2	99.9	89.3	80.4	85.0	64.7	140.2	108.3	97.3	104.8	102.6	136.8	83.9
4		104.0	99.2	89.0	97.8	91.9	88.1	92.2	99.4	107.7	104.1	95.3	110.5	101.8	130.4	84.0
6		58.6	79.1	84.3	98.1	73.1	77.8	84.2	98.8	98.3	108.4	96.5	N/A	74.4	107.3	82.5
7		24.4	46.2	41.2	89.8	51.5	25.5	64.4	61.2	77.9	92.4	40.5	62.6	47.8	67.8	72.8

Compound	CELL LINE	Renal						Ovarian	NCl/ADR-RES							
		766-0	A498	ACHN	C AKI-1	RXF 393	SN12C		TK-10	UO-31	IGROV1	OVCAR-3	OVCAR-4	OVCAR-5	OVCAR-8	SK OV-3
3		64.7	140.2	108.3	97.3	104.8	102.6	136.8	83.9	90.2	88.8	98.8	102.5	97.2	103.5	98.2
4		99.4	107.7	104.1	95.3	110.5	101.8	130.4	84.0	91.7	94.1	93.8	100.6	100.9	97.4	102.7
6		98.8	98.3	108.4	96.5	N/A	74.4	107.3	82.5	29.6	40.8	58.7	84.6	89.5	104.4	105.3
7		61.2	77.9	92.4	40.5	62.6	47.8	67.8	72.8	3.7	34.9	47.2	59.3	41.4	91.1	83.2

Compound	CELL LINE	Melanoma										Mean Growth
		LOXIMVI	MALME-3M	M14	MDA-MB-435	SK-MEL-2	SK-MEL-28	SK-MEL-5	UACC-257	UACC-62	Mean Growth	
3		97.5	100.9	96.0	101.7	109.9	112.0	104.0	103.6	109.5	97.0	
4		98.0	92.3	91.5	98.7	104.2	105.3	102.6	99.4	106.5	96.9	
6		94.9	66.5	N/A	84.8	89.1	94.5	68.3	56.1	61.1	75.0	
7		52.0	48.0	38.7	38.4	51.2	45.4	34.1	42.9	4.9	47.3	

332 bonyl resonances of the 9,10-anthraquinone moieties  
 333 were also observed for both compounds at 181.9 and  
 334 182.0 ppm.

335 The anthraquinone benzimidazolium salts, com-  
 336 pounds **6** and **7**, were synthesized by a similar route  
 337 to **3** and **4**, with formation of the mono-substituted  
 338 benzimidazole (**5**) first, then a subsequent alkylation.  
 339 Structure elucidation of both compounds was deter-  
 340 mined by <sup>1</sup>H NMR spectroscopy with the methylene

341 resonances at 6.11 ppm for the symmetric compound  
 342 **6** and 6.10 and 6.02 ppm for the asymmetric ben-  
 343 zimidazolium salt, **7**. Furthermore the C<sup>2</sup> proton  
 344 resonances were observed downfield at 10.26 and  
 345 10.24 ppm for **6** and **7** respectively. Similar to the  
 346 <sup>13</sup>C spectra of compounds **3** and **4**, both **6** and **7** had  
 347 C<sup>2</sup> carbon resonances at 143 ppm and the inequiva-  
 348 lent anthraquinone carbonyl resonances at 182.0 and  
 349 181.9 ppm.

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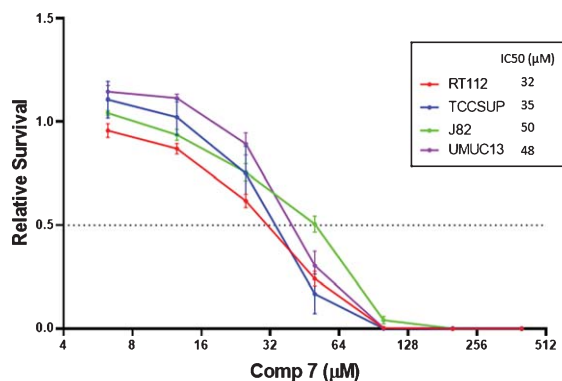


Fig. 3. IC<sub>50</sub> values determined by the CellTiter-Glo® assay for a 1 h exposure of select bladder cancer cell lines to compound 7.

To confirm the structures of the compounds discussed above, single crystal X-ray diffraction was used. Crystals suitable for analysis were obtained from vapor diffusion of ethyl ether into N,N-dimethylformamide solutions and only struc-

tures for compounds 4 and 7 were determined (Fig. 2).

#### *In vitro biological evaluation of anthraquinone compounds as anticancer agents*

The *in vitro* biological activity of compounds 3, 4, 6, and 7 was first evaluated by the Developmental Therapeutics Program (DTP) using the NCI-60 cell line screening (Table 1). At 10 µM the bisanthraquinone imidazolium salts, 3 and 6, had little inherent activity toward most of the 60 cell lines with a 48 h exposure.

The asymmetric compounds 4 and 7 had increased activity in comparison to compounds 3 and 6 against the NCI-60 cell lines, with compound 7 exhibiting the most cytotoxicity at an average percent growth inhibition (GI%) of 47.3% over all cell lines. This potency is believed to be directly caused by the addition of the naphthalene substituent and increased lipophilicity of the benzimidazole, as similar activity

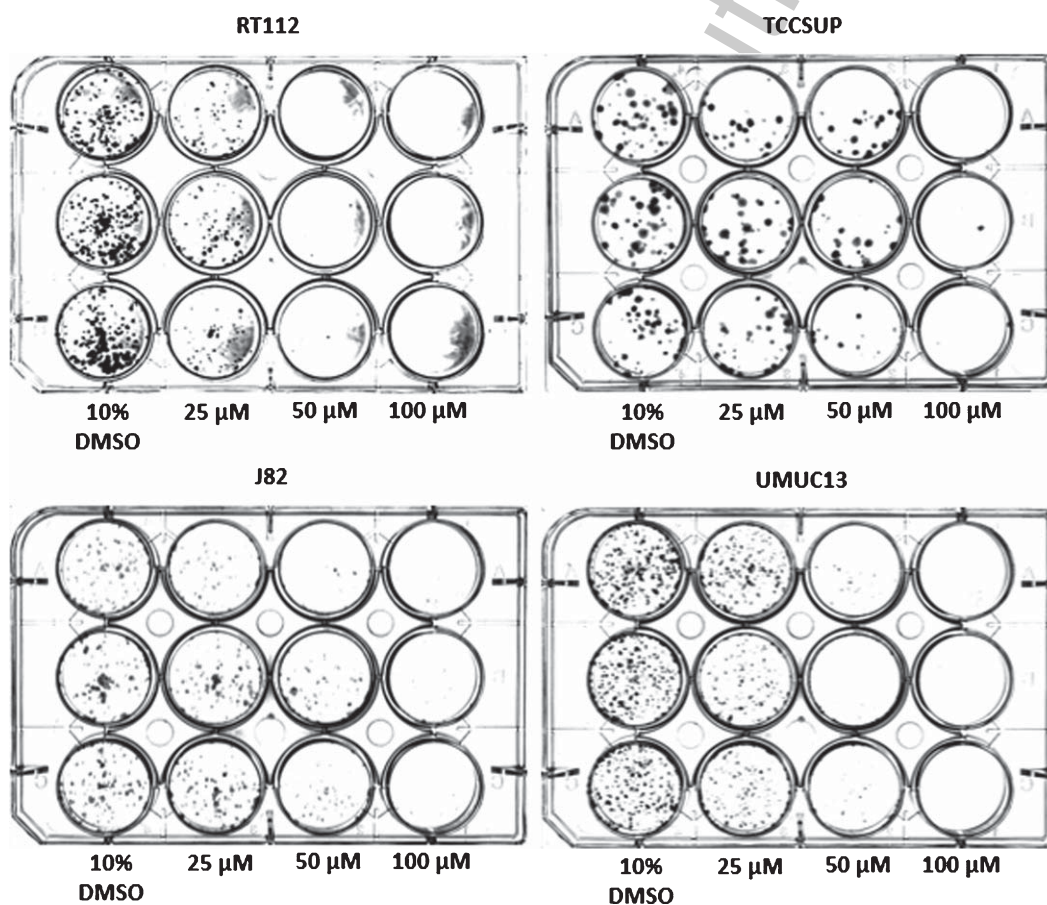


Fig. 4. Colony-forming assay of select bladder cancer cell lines 10 days after a 1 h treatment with compound 7.

374 has been previously reported by the Youngs group  
375 with respect to lung cancer [17].

376 With preliminary data indicating the increased  
377 cytotoxicity of compound **7** against the NCI-60  
378 cell line screening, further *in vitro* activity was  
379 evaluated against select bladder cancer cell lines.  
380 IC<sub>50</sub> values of 32, 35, 50, and 48 μM were deter-  
381 mined for a 1 h exposure of compound **7** to  
382 bladder cancer cell lines RT112, TCCSUP, J82, and  
383 UMUC13 respectively (Fig. 3). The shortened expo-  
384 sure time is necessary as intravesical treatments  
385 cannot be retained in patients for prolonged periods  
386 of time.

387 The long-term cytotoxicity of short-term expo-  
388 sure to compound **7** was evaluated next. Using the  
389 colony-forming assay, compound **7** was shown to  
390 inhibit cell growth against RT112 and UMUC13  
391 at 50 μM (1 h exposure) after a recovery period  
392 of 10 days. When increasing the concentration to  
393 100 μM, compound **7** inhibited cancer regrowth  
394 in all tested cell lines after 10 days recov-  
395 ery.

## 396 CONCLUSION

397 In this work, a series of anthraquinone-substituted  
398 imidazolium salts was synthesized. The bisan-  
399 thraquinone imidazolium and benzimidazolium salts,  
400 when tested by DTP on the NCI-60 cell line screen-  
401 ing, yielded little inherent activity against all cell lines  
402 at the 10 μM test concentration. Even though these  
403 compounds are lipophilic, the correlation between  
404 the naphthalene substituent and activity is appar-  
405 ent, as compounds **4** and **7** both exhibited activity  
406 against the cell lines in the screening. Moreover  
407 it was determined that replacement of the imi-  
408 dazole (**4**) with benzimidazole (**7**) resulted in a  
409 significant increase in cytotoxicity, believed to be  
410 due to the increase in lipophilicity. With prelimi-  
411 nary data from the NCI-60 showing compound **7**  
412 to be the most cytotoxic of the compounds, it was  
413 chosen for further study. Compound **7** exhibited  
414 significant *in vitro* activity against select bladder  
415 cancer cell lines after a 1 h exposure time. Fur-  
416 ther, compound **7** prevented bladder cancer regrowth  
417 in the colony-forming assay against all tested cell  
418 lines over a period of 10 days. With these results  
419 compound **7** exhibits a cell-killing effect in an  
420 acceptable time period against bladder cancer cell  
421 lines and may be a highly effective treatment for  
422 NMIBC.

## SUPPLEMENTARY INFORMATION 423

424 One dose NCI-60 human tumor cell line screening  
425 data for compounds **3**, **4**, **6**, and **7** as presented from  
426 DTP. NMR spectroscopy data for compounds **2–7**. IR  
427 spectroscopy data for compounds **3**, **4**, **6**, and **7**

428 Crystallographic Information File for **4** (CCDC #  
429 1988214) and **7** (CCDC # 1988213) can be found free  
430 of charge on the Cambridge Crystallographic Data  
431 Center website.

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## AUTHOR CONTRIBUTIONS 450

451 Conceptualization, M.S. and W.Y.; development  
452 and structural characterization, M.S.; biological  
453 investigation, D.W., U.S., and P.A.; writing-original  
454 draft preparation, M.S.; writing-review and editing,  
455 M.S., D.W., U.S., P.A., and W.Y. All authors have read  
456 and agreed to the published version of the manuscript.

## CONFLICTS OF INTEREST 457

458 The authors declare no conflict of interest.

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