

# Safety and efficacy of a novel iopromide-based paclitaxel-eluting balloon following bare metal stent implantation in rabbit aorta abdominalis

Zhengbin Zhu<sup>a,b,\*</sup>, Hui Han<sup>b,\*</sup>, Jinzhou Zhu<sup>a,b,\*</sup>, Jing Zhang<sup>c</sup>, Run Du<sup>a</sup>, Jingwei Ni<sup>a</sup>, Chen Ying<sup>a</sup>, Xuanqi An<sup>a</sup> and Ruiyan Zhang<sup>a,b,\*\*</sup>

<sup>a</sup> Department of Cardiology, Rui Jin Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai, China

<sup>b</sup> Institute of Cardiovascular Diseases, School of Medicine, Shanghai Jiao Tong University, Shanghai, China

<sup>c</sup> Shanghai Pharmaceuticals Holding Co. Ltd, Central Research Institute, Shanghai, China

Received 13 April 2015

Accepted 3 August 2015

## Abstract.

**BACKGROUND:** Drug-eluting balloons (DEB) may be promising technology for treating atherosclerotic arterial disease. In fact, several DEBs have been clinically available for the treatment of coronary in-stent restenosis (ISR), *de novo* coronary lesions, and peripheral artery disease.

**OBJECTIVE:** We sought to elucidate the mechanism of action and *in vivo* safety and efficacy of a novel iopromide-based paclitaxel-eluting balloon.

**METHODS:** *In vitro* cytotoxicity of a novel DEB on human umbilical vein endothelial cells (HUVECs) and *in vivo* pharmacokinetics of DEB in a rabbit aorta abdominalis were assessed. Then, bare metal stents (BMS) were implanted at both the proximal and distal sites of the rabbit aorta abdominalis. Stented vascular segments were immediately dilated with a bare balloon (control group) or the DEB (DEB group) randomly. Histological evaluation was performed in all treated segments at 28 days. Because paclitaxel is a tubulin-disrupting agent that binds preferentially to  $\beta$ -tubulin, we measured  $\beta$ -tubulin expression in aortal stent specimens via immunohistochemistry.

**RESULTS:** We observed that DEB was compatible and could reduce neointimal hyperplasia compared with the bare balloon. Meanwhile, immunohistochemistry revealed that  $\beta$ -tubulin expression in the DEB group increased compared with the control group, indirectly suggesting successful uptake of paclitaxel by vessel walls after DEB dilation.

**CONCLUSIONS:** The novel DEB is safe and has a favorable vascular healing response on neointimal hyperplasia.

Keywords: Neointimal hyperplasia, paclitaxel-eluting balloon, pharmacokinetics

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\*Zhengbin Zhu, Hui Han and Jinzhou Zhu contributed equally in this study.

\*\*Corresponding author: Ruiyan Zhang, Department of Cardiology, Rui Jin Hospital, School of Medicine, Shanghai Jiao Tong University, 197 Ruijin Road II, Shanghai, 200025, China. Tel.: +86 21 64370045; Fax: +86 21 64457177; E-mail: [rjzhangruiyan@aliyun.com](mailto:rjzhangruiyan@aliyun.com).

## 1. Introduction

Vascular stenosis and occlusion caused by atherosclerosis is a leading cause of death globally and these are initiated by coronary artery disease (CAD), stroke, and peripheral artery disease [1]. As a standard therapy, percutaneous transluminal angioplasty (PTA) has been upgraded from simple balloon angioplasty to bare metal stent (BMS) implantation, and on to drug-eluting stent (DES) treatment [2]. Early vessel recoil and subsequent neointimal hyperplasia caused by arterial remodeling still causes restenosis, however, and this limits the long-term clinical outcomes of balloon angioplasty [3]. These limitations were thought to have been largely eliminated with the introduction of BMS [4]. However, BMS stimulated greater neointimal and produced a new problem: in-stent restenosis (ISR) [5]. DES were then developed and were reported to reduce neointimal hyperplasia [6,7] and this decreased ISR to ~5–10% after treating coronary lesions [8]. Even so, restenosis rates are unacceptably high and the small but unpredictable risk of very late stent thrombosis may still occur due to hypersensitivity reactions to the stent polymer coating [9,10].

Thus, novel strategies to ameliorate neointimal hyperplasia and stent thrombosis are needed. The goal of using PTA without leaving behind a permanent device, led to the development of drug eluting balloons (DEB) [11]. First experiments with DEBS included paclitaxel-coated balloons (PCB) in pigs [12,13], and soon DEB was seen as an alternative treatment to plain balloon angioplasty and stent implantation. Studies with promising long-term results with DEB for treating coronary ISR compared with uncoated balloons and paclitaxel-coated stents were published [14–16] and reports of DEB treatment of lesions in small coronary arteries followed [17,18].

We sought to assess the blood compatibility of a newly designed iopromide-based paclitaxel-eluting balloon *in vitro*, and evaluate the efficacy of the DEB by measuring vessel neointimal hyperplasia in a rabbit aorta abdominalis model.

## 2. Materials and methods

### 2.1. Device description

DEB catheters are single-use hydrophilic coated with polyvinylpyrrolidone (PVP). Balloons were  $3.0 \times 20$  mm coated with a paclitaxel-iopromide formulation ( $3 \mu\text{g}/\text{mm}^2$  of balloon surface) using a proprietary dipping process that deposited the formulation preferentially in the folds of the balloon.

### 2.2. Cytotoxicity testing

Human umbilical vein endothelial cells (HUVECs) were seeded at low density (30,000 cells per 12-well culture plates) and cultured in M200 supplemented with 10% fetal bovine serum (FBS) and growth factors for 24 h. Cells were then co-incubated with an iopromide-coated balloon without paclitaxel ( $n = 6$ ) for 2 days. HUVEC cultures were incubated with a bare balloon as controls ( $n = 6$ ). After the incubation period, 50  $\mu\text{l}$  of cell proliferation reagent, WST-1 (Roche Applied Science, Mannheim, Germany) was added to each well. After 4 h of WST-1 addition, absorbance at 450 nm was measured via an enzyme-linked immunosorbent assay (ELISA) plate reader (BioTek, Winooski, VT, USA).

### 2.3. *In vivo* pharmacokinetic study

All the experimental procedures were approved by the Animal Use and Care Committee of Shanghai Jiao Tong University School of Medicine. *In vivo* pharmacokinetics of DEB was measured in a normal rabbit aorta abdominalis, as normal vessels are typically used to study *in vivo* pharmacokinetics of new biomaterials [19,20]. Briefly, New Zealand white rabbits (SLACCAS, Shanghai, China) (2.5–3.0 kg) were sedated and anesthetized with ketamine (20 mg/kg, iv). The right common iliac artery was surgically exposed under sterile conditions and punctured with a 20 G puncture needle (Terumo, Tokyo, Japan). After advancement of a 0.014-in Runthrough™ guidewire (Terumo, Tokyo, Japan) into the aorta with the tip near the aortic arch under digital subtraction angiography (INNOVA 2100, GE Healthcare, Buckinghamshire, UK), an area of the aorta abdominalis ~2.5 mm in diameter as measured by quantitative coronary angiography (QCA) was chosen and the DEB was inflated in this area to standard 10 atm to 3.0 mm for 60 s with a balloon vessel ratio of 1.2:1. The arteriotomy and dermal layers were sutured after the catheter, wire, and sheath were removed. Animals were euthanized after 1 h, or 1, 7, and 28 days after the procedure ( $n = 3$  for each time point), and treated vessels (nearly 30 cm including the upper and lower ends of the dilated vessels) were harvested for paclitaxel measurement using a reverse phase high performance liquid chromatography (HPLC) assay. Results are expressed as the cumulative weight percent of paclitaxel released.

### 2.4. Safety and efficacy study

Five Male New Zealand white rabbits were fed normal laboratory chow for seven days. Three days before the procedure and throughout the following period, all animals received 10 mg aspirin and 12.5 mg clopidogrel daily (po). Arterial access was achieved with the method described above and two BMSs were implanted in the aorta abdominalis, which would damage the normal endothelial cell layer of the vessel based on previous reports [21]. A control balloon and the DEB were then randomly dilated in each of the stented vessels to 10 atm for 120 s, deflated for 60 s, and re-dilated to 10 atm for another 60 s. Four weeks after the procedure animals were sacrificed with an overdose of anesthetic and the stented segments of the aorta abdominalis were harvested and immediately fixed in 10% formalin.

### 2.5. Histology analysis

The stented and balloon-dilated arterial segments, obtained after 28 days, were stained with hematoxylin and eosin (H&E) according to published methods [21]. Neointimal thickness is obvious after 4 weeks of stent implantation in rabbit model. Samples were dehydrated in a graded series of ethanol and embedded in methylmethacrylate plastic. After polymerization, the center section of each stent was cut on a rotary microtome (EXAKT, Norderstedt, Germany) into 15–20  $\mu\text{m}$  sections and stained with H&E. An experienced pathologist blinded to the treatment groups performed all histological analyses. Ordinal data including injury and inflammation scores, neointimal thickness and area, percent luminal stenosis, lumen area, and internal elastic membrane area were collected on each stent section according to previously reported methods [22].

### 2.6. Immunohistochemistry

Paclitaxel is a tubulin-disrupting agent that binds preferentially to  $\beta$ -tubulin, the basic structural unit of microtubules. Thus, paclitaxel can interfere with the normal microtubules degradation during cell division. For immunohistochemical analysis of  $\beta$ -tubulin, cross-sections of stented vessels were fixed in

10% formalin at room temperature for 1 day. Stents were then carefully removed from vessels under microscopy. Vessels were dehydrated through an upgraded ethanol series and embedded in paraffin blocks. Ultrathin sections (5 mm) were deparaffinized, rehydrated, and incubated for 5 min in 3% H<sub>2</sub>O<sub>2</sub> to inhibit endogenous peroxidase activity. Masking of non-specific binding sites was performed by incubation in 5% bovine serum albumin for 30 min. Subsequently, sections were incubated with mouse anti-rabbit  $\beta$ -tubulin monoclonal antibody (Abcam, Cambridge, MA, USA) diluted to a ratio of 1:1,000 overnight at 4°C, and then incubated with biotin-conjugated rat anti-mouse secondary antibody for 30 min at 37°C, and finally incubated in streptavidin-biotin complex (SABC) for 30 min at 37°C. Peroxidase activity was measured after incubation with diaminobenzidine tetrahydrochloride (DAB). After washing, sections were weakly counterstained with hematoxylin. Observations and photographic records were made using a phase contrast microscope.

### 2.7. Statistics

Results are expressed as means  $\pm$  standard deviation (SD). Statistically significant differences among different treatment groups at a single time point were determined by ANOVA, followed by two-tailed Student's *t*-tests. Statistical significance was assumed for *P* values  $<0.05$ .

## 3. Results

### 3.1. Cytotoxicity testing

The cytotoxicity of iopromide-coated balloon treatment was quantitatively measured via WST-1 assay. Cell viability was expressed as a fraction of viable cells and normalized to that of cells co-incubated with the bare balloon (control). Data showed that the cytotoxicity of iopromide-coated balloon was Grade 1 according to the test standard of United States Pharmacopeia (USP) XXII, as HUVEC viability in iopromide-coated balloon group was 86.4% compared with controls, suggesting reliable cytocompatibility for the DEB (Fig. 1).

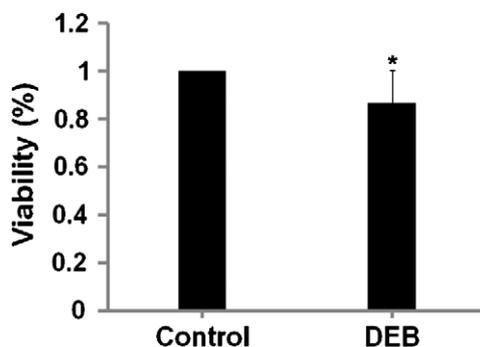


Fig. 1. WST-1 assay indicates human umbilical vein endothelial cell (HUVEC) viability after co-incubation with iopromide-coated balloon or bare balloon up to 48 h. Data are expressed as means  $\pm$  SD of six independent experiments (ratio on control 1). \**P*  $< 0.05$  vs. control group.

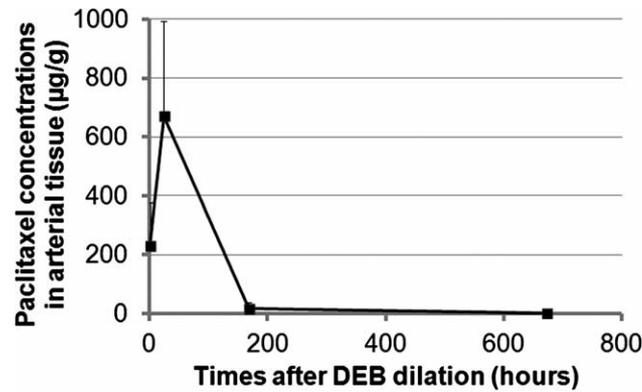


Fig. 2. Paclitaxel release kinetics of DEB *in vivo*.

### 3.2. Release pharmacokinetics of DEB *in vivo*

*In vivo* elution of paclitaxel from the DEB revealed that in the tissue, the cumulative diffused drug was  $230.98 \pm 147.42 \mu\text{g/g}$  at 1 h,  $670.53 \pm 323.31 \mu\text{g/g}$  at 1 day,  $15.88 \pm 21.23 \mu\text{g/g}$  at 7 days, and  $1.1 \pm 1.64 \mu\text{g/g}$  at 28 days (Fig. 2).

### 3.3. Histomorphometry and histology

All animals underwent successful implantation of BMS and balloon dilation (Fig. 3). At 28 days, there were significant differences in luminal area, neointimal thickness, neointimal area, and percent stenotic lumen among DEB and control groups, and injury and inflammation scores were similar in both groups (Table 1; Fig. 4).

### 3.4. Immunohistochemical detection of $\beta$ -tubulin

Immunohistochemically, brown nuclei were  $\beta$ -tubulin positive. Data show that  $\beta$ -tubulin was dramatically increased in the DEB group compared with the control group (arrowhead), suggesting that paclitaxel was involved in anti-restenosis effects in the DEB group (Fig. 5).

## 4. Discussion

DES cannot completely reduce restenosis after PTA and ameliorate the need for revascularization; thus, the risk of stent thrombosis remains [23,24]. Also, longer dual antiplatelet therapy of usually 12 months is needed compared to only one month for BMS [25]. Thus, BMS is preferred for 20–30% of patients [26]. DEB was therefore designed to integrate advantages of bare balloon treatment and DES. We found that a novel iopromide based paclitaxel-eluting balloon could better reduce neointimal hyperplasia in rabbit aorta abdominalis compared with bare balloon therapy.

The drug of choice for local delivery should be optimally lipophilic, highly absorbed or rapidly taken up by the intima to compensate for the short contact time, as well as a high retention rate, and sustained

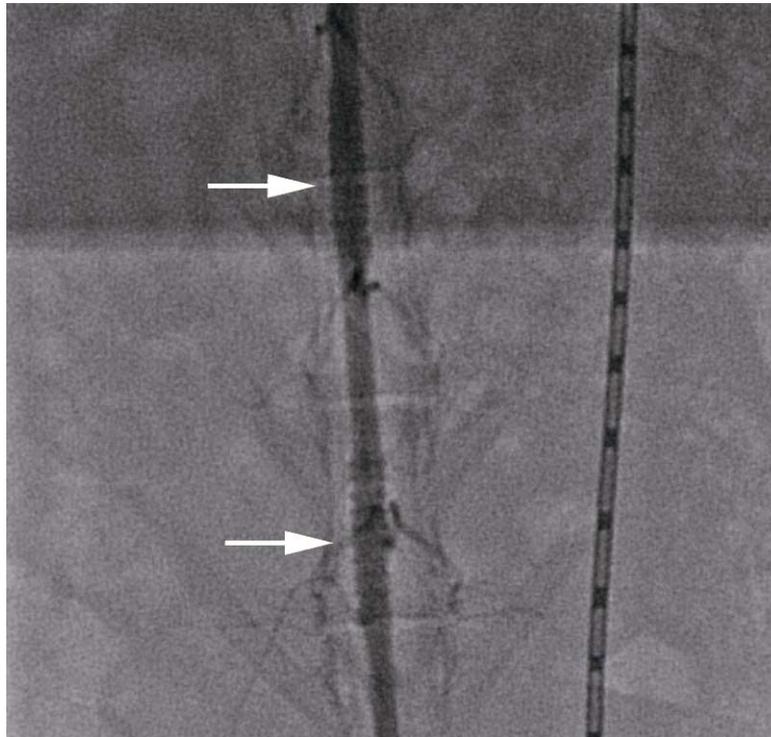


Fig. 3. Representative images of aorta abdominalis assessed by angiography immediately after BMS implantation (arrowhead: BMS implantation sites). (Colors are visible in the online version of the article; <http://dx.doi.org/10.3233/BME-151551>.)

Table 1  
Morphometric comparisons

|                            | DEB group                | Control group |
|----------------------------|--------------------------|---------------|
| 28 day follow-up, <i>n</i> | 5                        | 5             |
| Injury score               | 1.54 ± 0.12              | 1.52 ± 0.15   |
| Mean LA, mm <sup>2</sup>   | 8.14 ± 0.30 <sup>▲</sup> | 6.96 ± 0.49   |
| Mean IEM, mm <sup>2</sup>  | 8.63 ± 0.13              | 8.56 ± 0.23   |
| Mean NA, mm <sup>2</sup>   | 0.49 ± 0.18 <sup>▲</sup> | 1.60 ± 0.29   |
| Mean NT, mm                | 0.04 ± 0.01 <sup>▲</sup> | 0.09 ± 0.01   |
| Percent area stenosis, %   | 0.06 ± 0.02 <sup>▲</sup> | 0.19 ± 0.04   |
| Inflammation score         | 0.96 ± 0.16              | 0.97 ± 0.21   |

LA: average lumen area; IEM: average internal elastic membrane area; NA: average neointimal area; NT: average neointimal thickness; *n*: number of sections. <sup>▲</sup>Compared with control group, *P* < 0.05.

effects [27]. To date, paclitaxel has met these criteria. Contrast media agents (CMA) such as iopromide adhere to the coronary vessel wall for several seconds after injection, and these have been used safely and efficiently to deliver paclitaxel to the arterial wall [28,29].

Paclitaxel-coated balloon (PCB) technologies have now been clinically introduced as an alternative therapy to DES and plain balloon angioplasty (PBA) for treatment of coronary in-stent restenosis (ISR) [15,30,31], *de novo* peripheral artery disease [32,33], and even *de novo* coronary lesions [34]. Some

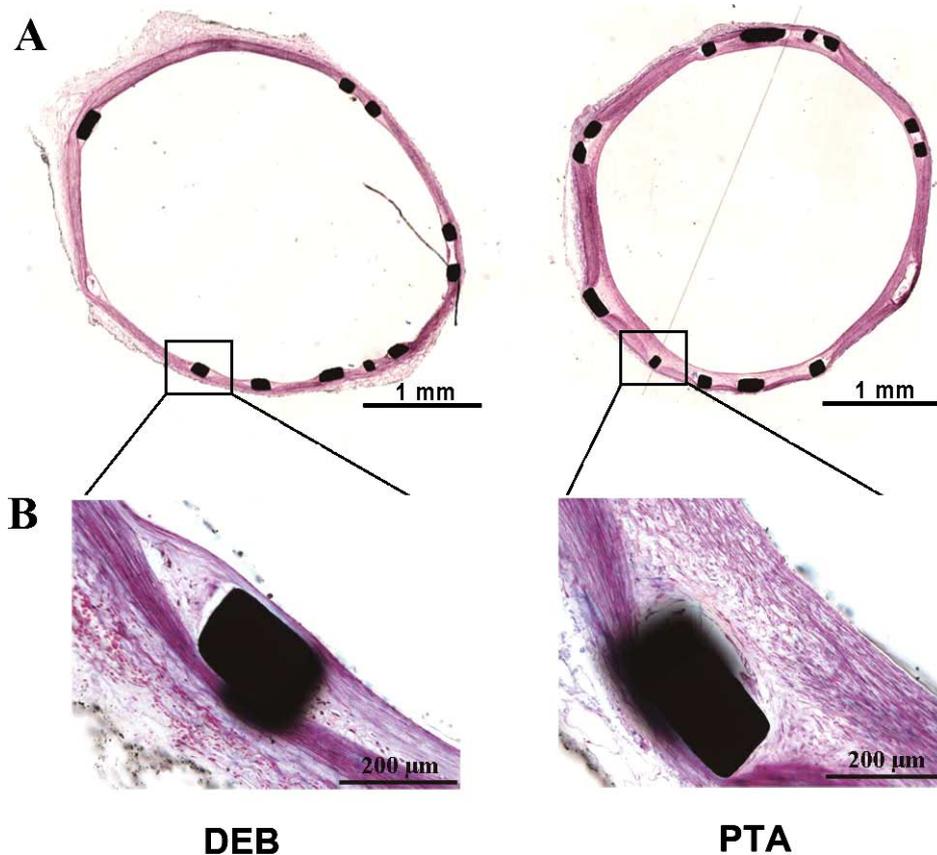


Fig. 4. Representative histological images at low-magnification (100x, upper image) and high power magnification (200x, lower image) 28 days after balloon treatment. DEB could inhibit neointimal hyperplasia in contrast with control. Sections shown are stained with hematoxylin and eosin (H&E). (Colors are visible in the online version of the article; <http://dx.doi.org/10.3233/BME-151551>.)

paclitaxel-coated balloons are also commercially available, including Paccocath™, SeQuent™, DIOR et al. [35].

It has been reported that an inflation time of 1 min is sufficient to allow diffusion of a high concentration of paclitaxel to the arterial wall [14,29,36]. In our hands, a 2-min dilation, with 1-min deflated, followed by another 1-min dilation dramatically increased paclitaxel content in the arterial tissue. Meanwhile, immunohistochemistry of paclitaxel-targeted  $\beta$ -tubulin revealed greater  $\beta$ -tubulin expression in the DEB group compared to controls.

Restenosis occurs because of vessel recoil and plaque prolapse after bare balloon angioplasty, and it can arise from neointimal hyperplasia due to smooth muscle cell migration and proliferation after stenting [37]. Moreover, stent thrombosis now is thought to chiefly occur due to a hypersensitivity reaction to the stent polymer coating [38]. Success in inhibiting neointimal proliferation has been achieved with paclitaxel which targets vascular smooth muscular cell proliferation. Although paclitaxel has been used in both DES and DEB, DES is limited by non-uniform drug delivery to the arterial and a high drug concentration at stent struts. This does not occur between the struts and at the margins [9]. Ideally, DEB should permit homogenous drug transfer to the vessel wall, avoid the use of polymers and stents and

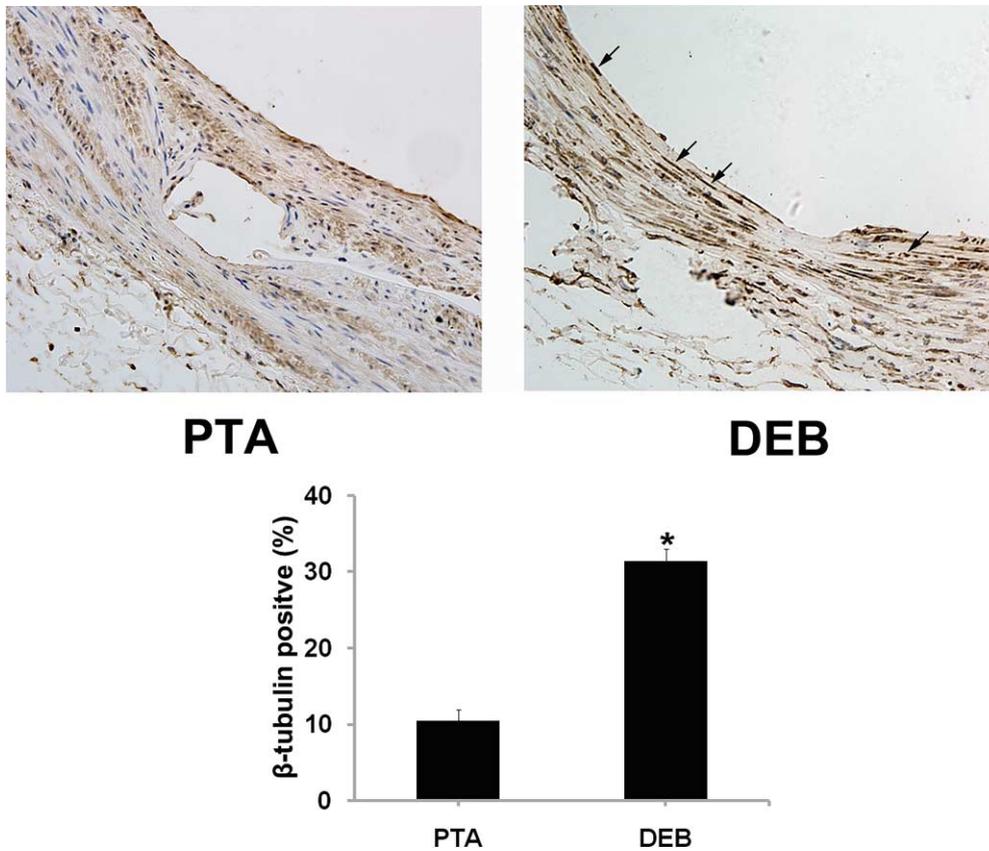


Fig. 5.  $\beta$ -tubulin expression in stented vascular vessels was measured via immunohistochemistry. Quantitative measurement of immunohistochemical images as a ratio of  $\beta$ -tubulin-positive cells indicated that  $\beta$ -tubulin expression was induced in the DEB group (arrowhead). All values are given as means  $\pm$  SD. \* significantly different compared to the control group ( $P < 0.05$ ). (Colors are visible in the online version of the article; <http://dx.doi.org/10.3233/BME-151551>.)

thus obviate the need for antiplatelet therapy. Also, DEB may be used for high-risk re-stenotic lesions such as small vessel-, bifurcation-, or in-stent re-stenotic lesions.

DEB angioplasty may be a promising treatment option for ISR [31]. A recently published meta-analysis indicated that DEB was superior to bare balloon angioplasty and comparable to DES for treating DES restenosis [39]. However, DEB does not appear to be superior to current standard therapies (BMS or DES) for treating *de novo* coronary lesions, but may be better than BMS alone [34]. Additional and ongoing trials may help to conclude this matter [40].

## 5. Conclusions

Our results revealed a comparable efficacy profile and more favorable vascular healing response of this novel DEB. This at least partially indicate the further study of the novel DEB in treating high-risk re-stenotic lesions and pre-treatment of lesions preferred to receive BMS thus making complementary advantages.

## Acknowledgements

This work was supported by the Science and Technology Commission of Shanghai Municipality (grant 10441901102 and 12441903200), and was partly supported by the National Natural Science Foundation of China (81300178 and 81370401).

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