Genetic Influences on Atrophy Patterns in Familial Alzheimer’s Disease: A Comparison of APP and PSEN1 Mutations


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Abstract. Mutations in the presenilin1 (PSEN1) and amyloid β-protein precursor (APP) genes account for the majority of cases of autosomal dominantly inherited Alzheimer’s disease (AD). We wished to assess and compare the patterns of cerebral loss produced by these two groups of mutations. Volumetric magnetic resonance imaging and neuropsychological assessments were performed in individuals with clinical AD carrying mutations in the APP (n = 10) and PSEN1 (n = 18) genes and in healthy controls (n = 18). Voxel-based morphometry (VBM), cortical thickness, and region of interest analyses were performed. Mini-Mental State Examination scores were similar in the two disease groups suggesting similar levels of disease severity. There was evidence that APP subjects have smaller hippocampal volume compared with PSEN1 subjects (p = 0.007), and weak evidence that they have larger whole-brain and grey matter volumes (both p = 0.07). Although there was no evidence of statistically significant differences between APP and PSEN1 in VBM or cortical thickness analyses, effect-maps were suggestive of APP subjects having more medial temporal lobe atrophy and conversely PSEN1 subjects showing more neocortical loss. Neuropsychological data were consistent with these regional differences and suggested greater memory deficits in the APP patients and greater impairment in non-memory domains in the PSEN1 group, although these differences were not statistically significant. We conclude that the mechanisms by which APP and PSEN1 mutations cause neuronal loss may differ which furthers our understanding of the neuropathology underlying AD and may inform future therapeutic strategies and trial designs.

Keywords: Alzheimer’s disease, APP, atrophy, autosomal dominant, magnetic resonance imaging, PSEN1

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INTRODUCTION

Alzheimer’s disease (AD) is the most common cause of dementia, affecting up to 20 million people worldwide [1]. Although AD is usually sporadic and of late onset, a minority of cases are autosomal dominantly inherited—familial AD (FAD). FAD can be caused by mutations in the amyloid beta (Aβ) precursor protein (APP) [2], presenilin 1 (PSEN1) [3], and PSEN2 genes [4] and by duplications of the APP gene [5]. Most recently, mutations in the sortilin-related receptor LR11 (SORL1) gene have been implicated in FAD [6]. Individuals with FAD tend to have a younger age at onset than those with the more typical sporadic AD (SAD) and the disease is inherited with virtually 100% penetrance. Studying subjects with such mutations has the potential to further our understanding of AD [7]; subjects can be diagnosed with greater certainty than those with SAD, comorbidity is rare, and at-risk individuals can be feasibly assessed many years prior to the onset of symptoms. These mutations have been incorporated into cellular models of AD, and transgenic mouse models have driven therapeutic developments. Recent initiatives have suggested a need to investigate treatments as early as possible in the disease process, ideally in healthy individuals who have a high risk of developing the disease, and FAD individuals form a promising cohort for such studies [7, 8]. Recently, the first ever therapeutic prevention trial in cognitively healthy individuals carrying familial AD mutations was announced (http://www.nia.nih.gov/newsroom/2012/05/obama-administrat-on-presents-national-plan-fight-alzheimers-disease). According to the amyloid cascade hypothesis, the effects of mutations in the APP and PSEN genes are thought to feed into a final common pathway driven by the accumulation of toxic misfolded amyloid-β (Aβ) entities, resulting in neurodegeneration and cognitive decline. Previous work has suggested that individuals with FAD may have relative preservation of naming, spelling, and visuoperceptual function compared with SAD subjects [9], conversely isolated slowly-progressive amnesia has been reported in an individual with a V717G mutation in the APP gene [10]. There is clearly heterogeneity in the clinical presentation of FAD [11] and it is likely that the location of the mutation within the gene can affect the phenotypic profile [12]. Additional neurological symptoms and signs appear to occur more commonly in FAD than SAD; myoclonus and seizures have been reported in both APP and PSEN families [5, 13, 14] and PSEN1 mutations producing spastic paraparesis [15] and cerebellar ataxia [16] have also been described. Extrapyramidal signs have been reported in both PSEN1 [17] and PSEN2 subjects [18].

Although previous pathological studies have shown some differences between the sporadic and familial forms of the disease (reviewed in [19]), there are few studies directly comparing pathology in the different mutations. Gregory and colleagues demonstrated more grey matter reduction in the frontotemporal regions in individuals with PSEN1 than those with an APP mutation [20], but Shepherd et al. failed to show a difference between these genotypes in terms of neuronal loss [21]. Amyloid PET studies have reported different patterns of Pittsburgh compound B (PiB) retention in FAD compared with SAD, most notably elevated PiB retention in the striatum of both PSEN1 and APP mutation carriers [22, 23]. The numbers of subjects in these studies are unsurprisingly small, and it appears that there can be considerable variability in pathology, even within the same family [24]. Understanding the neuropathological changes occurring early in the course of AD is important as novel disease-modifying therapies being developed may well be more effective at this stage. Patients with FAD in whom a definitive early diagnosis can be established in vivo can therefore provide valuable insights. Using this approach, we have previously demonstrated changes prior to symptom onset in FAD [25–27]. Examining variation in the patterns of atrophy in the different familial mutations at a group level (APP versus PSEN1) may be a means of understanding the extent to which genotype influences the distribution of neuropathology. We therefore aimed to investigate differences between individuals with FAD caused by mutations in the APP and PSEN1 genes. Using an automated whole-brain image analysis technique, voxel-based morphometry (VBM) [28], we examined differences in grey and white matter throughout the brain. We measured cortical thickness with an independent technique and also performed specific manual volumetric measures of the hippocampus and the cingulate gyrus in APP and PSEN1 cases. Correlates of these volumetric differences in terms of the cognitive profiles of each mutation group were examined.

MATERIALS AND METHODS

Subjects

Twenty-eight FAD subjects with an autosomal dominant family history of AD were included; ten had a
Table 1
Demographics and neuropsychological assessments. Mean (SD)

<table>
<thead>
<tr>
<th></th>
<th>Controls</th>
<th>APP (SM, SF)</th>
<th>PSEN1 (1M, TF)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age/years</td>
<td>51.3 (12.8)</td>
<td>55.0* (6.8)</td>
<td>47.0* (7.8)</td>
</tr>
<tr>
<td>APOE status</td>
<td>3/3</td>
<td>5</td>
<td>11</td>
</tr>
<tr>
<td>MMSE/30</td>
<td>29.5* (8.8)</td>
<td>18.6* (4.5)</td>
<td>17.3* (6.0)</td>
</tr>
<tr>
<td>NART – verbal IQ</td>
<td>17.6 (16.4)</td>
<td>19.6 (14.7)</td>
<td></td>
</tr>
<tr>
<td>NART – performance IQ</td>
<td>25.6 (20.2)</td>
<td>27.5 (18.8)</td>
<td></td>
</tr>
<tr>
<td>RMT – words/grade</td>
<td>1.67 (1.12)</td>
<td>1.88 (1.26)</td>
<td></td>
</tr>
<tr>
<td>RMT – faces/grade</td>
<td>2.00 (1.30)</td>
<td>2.06 (1.48)</td>
<td></td>
</tr>
</tbody>
</table>

NART, National Adult Reading Test; RMT, Recognition Memory Test.

*PSEN1 < APP (p = 0.003).
†Controls > AD groups (p ≤ 0.01).

mutation in the APP gene and eighteen had a mutation in the PSEN1 gene. The majority of the mutations in this study have been reported elsewhere but three subjects with novel mutations were also included. The APP group comprised subjects with the following mutations: four V717I (of which three were from the same family), three V717G (all from the same family), one V717L, one A692G, and one with the novel T719N mutation. The PSEN1 group comprised subjects with the following mutations: six M139V (including three from one family and two from an unrelated family), two L235V (both from the same family), two intron 4 (from unrelated families), and one each of E280G, I143F, L250S, E184D, pL166del, and F237L, and the novel mutations Q222P and F283L. All novel sequence variants identified were absent from 100 healthy unrelated white control patients. The individuals with novel mutations all had a typical clinical presentation for FAD, with impairment of episodic memory progressing to global cognitive decline from their mid-late 40s. Further information regarding these novel mutation cases is detailed in a separate paper undergoing preparation for publication. APOE status was available for nine of the APP group and 16 of the PSEN1 group and details are provided in Table 1.

All patients in this study fulfilled criteria for probable AD at the time of their MRI assessment [29]. Eighteen cognitively normal controls were also included, in order to improve the estimation of any age and gender effects. Subjects gave written informed consent according to the Declaration of Helsinki. Consent was taken by a clinician experienced in the assessment of patients with cognitive impairment and all subjects were considered to have capacity to consent according to the UK Mental Capacity Act of 2005. Ethical approval was received from the local ethics committee. Subject demographics are shown in Table 1.

MRI

All subjects underwent MR imaging on a 1.5 tesla Signa unit (General Electric, Milwaukee). T1-weighted volumes were acquired with 124 1.5 mm coronal partitions and a 256 × 256 matrix covering a 24 cm field of view. T2-weighted dual-echo images were acquired with 44 5 mm slices with 2.5 mm inter-slice spacing. All images were reviewed to identify artifacts or white matter hyperintensities or infarcts.

Voxel-based morphometry

VBM processing was performed using SPM5 (Statistical Parametric Mapping, Wellcome Trust Centre for Neuroimaging, London, UK, (http://www.fil.ion.ucl.ac.uk/spm/) running on Matlab 7 (release 14) platform (The Mathworks, MA, USA). Initial tissue classification was performed using the unified segmentation process with rigid alignment [30]. DARENTEL (diffeomorphic anatomic registration through exponentiated lie algebra) was then used to spatially normalize (with volume-preserving modulation) the initial tissue segmentations to an iteratively estimated study-specific template [31]. We used a smoothing kernel of 4 mm full width at half maximum (FWHM) with the aim of allowing accurate anatomical localization of fine-scale group differences.

Cortical thickness

Cortical thickness measurements were performed using FreeSurfer version 4.0.3 (https://surfer.nmr.mgh.harvard.edu/). This technique has been described previously [32]. In summary, FreeSurfer generates a white-matter segmentation, from which the grey/white matter and pial surfaces are generated; cortical thickness measurements are computed using the average of the shortest distance from one surface to the other. Cortical thickness measures were smoothed on the average surface by 20 mm FWHM.

Volumetric analysis

The in-house MIDAS package was used to measure whole-brain volume semi-automatically [33]; hippocampal and cingulate volumes were measured manually using previously described protocols [34, 35]; left and right sides were summed together for
hippocampal and cingulate volumes. Total intracranial volume (TIV) was measured according to the protocol of Whitwell et al. [36].

Neuropsychological assessment

All subjects underwent a neurological assessment including testing with the Mini-Mental State Examination (MMSE) [37]. In addition, FAD subjects were assessed on measures of verbal IQ and performance IQ, and estimated premorbid IQ (National Adult Reading Test (NART)) [38]. Two Dementia Index scores were calculated by subtracting the verbal IQ and performance IQ scores respectively from the NART IQ score. Assessment of visual and verbal memory was performed using the recognition memory tests (RMT) for words and faces [39]. Individuals completed the full, short [40] or easy [41] version of each test and all raw scores were converted to grades as follows: 1: <1st percentile; 2: 1st–5th percentile; 3: 6th–10th percentile; 4: 11th–25th percentile; 5: 26th–50th percentile and 6: >50th percentile. The neuropsychological test battery has been described in detail previously [13].

Statistical analysis

Regional volumetric data, VBM, and cortical thickness were all fitted with linear models comprising terms for group (Controls, APP, and PSEN1 as a three-level factor), gender, age, and TIV. Models were fitted which allowed the residual variance to differ between subject group. In addition to reporting (adjusted) means, we report the difference between the estimated (adjusted) means for APP and PSEN1 expressed as a percentage of the average of these two estimated means: $200 \times (\text{APP} - \text{PSEN1})/(\text{APP} + \text{PSEN1})$.

MRI

We found no evidence of significant artifacts or white matter lesions on visual inspection with the exception of one PSEN1 subject (F283L mutation) who had a small (clinically non-specific) localized lesion in the white matter of the superior frontal lobe. This lesion was not judged sufficient to exclude this individual.

VBM analysis

A direct comparison of the APP and PSEN1 groups did not reveal any significant differences in atrophy pattern in either grey or white matter at the FDR <5% level. However, effect-maps suggest that APP subjects had more atrophy in the medial temporal lobe regions and those with the PSEN1 mutation had more widespread cortical involvement (Fig. 1) and more widespread white matter atrophy (Fig. 2). Comparing each disease group with the controls (Fig. 3), we found evidence that the APP subjects have atrophy predominantly in the medial temporal lobes, with the white matter atrophy restricted to this region, whereas PSEN1 subjects showed widely distributed atrophy in both the grey and white matter.

Cortical thickness measurements

A direct comparison of the APP and PSEN1 groups did not reveal any significant differences in cortical...
thinning at the FDR <5% level. However, effect-maps suggest that there may be more focal medial temporal lobe thinning in the APP subjects and greater involvement of cortical areas in the PSEN1 group (Fig. 4). Comparisons of each mutation group with controls are shown in Fig. 5, which is consistent with a differential pattern of cortical thinning in these two groups.

Volumetric analysis

There was weak evidence that after adjustment for age, gender, and TIV the APP group had larger (52 ml 95% CI (−3.6, 108), $p = 0.065$) whole-brain volumes and larger (44 ml 95% CI (1.4, 91), $p = 0.067$) grey matter volumes (Table 2). There was no evidence of a difference in mean white matter volume ($p = 0.15$) or cingulate volume ($p = 0.83$). There was evidence that APP patients had significantly smaller hippocampi than the PSEN1 group (6.99 ml 95% CI 0.29, 13.69), $p = 0.042$. Supplementary Figures 1 to 5 show scatterplots of volumetric measures against age by group (available online: http://www.j-alz.com/issues/35/vol35-1.html#supplementarydata04). When adjusted for whole-brain volume (in addition to age, gender, and TIV), there was strong evidence that hippocampal volume was smaller in APP than PSEN1 (9.51 ml 95% CI 2.85, 16.17), $p = 0.007$.

DISCUSSION

This study suggests that there may be differing patterns of regional atrophy between APP and PSEN1 FAD mutation groups. We found statistically significant evidence that APP patients have smaller hippocampi compared with PSEN1 subjects, despite being similar in terms of disease severity. This difference became larger when adjustment was made for whole-brain volume, suggesting that the hippocampal volume difference cannot be ascribed to greater global atrophy in the APP group. Furthermore, there was weak evidence that the APP group had less global atrophy, with higher whole-brain and grey matter volumes than PSEN1 subjects.

Although direct group comparisons did not reveal statistically significantly different grey matter atrophy patterns, VBM and cortical thickness effect-maps suggest the PSEN1 subjects have at least as much, if not more cortical involvement compared with the APP patients, and while non-significant, the trends for reduced whole-brain, grey and white matter volumes in the PSEN1 group compared with the APP subjects further support this suggestion. Likewise, the effect-maps are suggestive of more focal temporal lobe grey matter atrophy in the APP subjects then the PSEN1 patients. There is some support for this finding from postmortem studies which have shown greater grey matter loss in PSEN1 subjects compared with those with APP mutations [20].

In parallel with the grey matter changes, white matter atrophy appears relatively more localized in the APP group, whereas the PSEN1 subjects had extensive white matter involvement of occipital, parietal, and frontal lobes. This marked white matter involvement might explain the greater occurrence of atypical clinical features including spastic paraparesis in some patients with PSEN1 mutations. Since VBM results may be affected by regional signal intensity differences as well as by atrophy we conducted a post hoc SPM analysis (not shown here) and were unable to detect any significant local intensity differences in white matter. It is likely that other imaging modalities such as diffusion-weighted MRI and quantitative mapping of parameters such as magnetization transfer, which reflect more specific properties of white matter, will allow for more thorough investigation of gene-specific white matter pathology [45].

Amyloid β-protein precursor (AβPP) and presenilin (PSEN) have different biological functions [46]. AβPP is the substrate for the γ-secretase complex of which PSEN forms a part; differential abnormali-

ties in function may therefore exert different regional neuropathological effects. PSEN has roles beyond the cleavage of AβPP, for example in NOTCH processing [47]. In AβPP, the precise location of the mutation has critical implications for its functional effects: mutations pathogenic for AD are particularly clustered around the β- and γ-secretase cleavage sites that bracket the Aβ domain or within the Aβ domain around position 692/693 (http://modgen-www.uia.ac.be/ADMutations) [11, 48]. All but one of the APP subjects in our study had mutations just distal to the γ-secretase site at codon 717, which cause relative overproduction of Aβ peptides ending at residue 42/43. Irrespective of effects on other substrates, it is increasingly clear that different familial AD mutations may affect Aβ generation by different mechanisms which results in qualitative changes in the profiles of the Aβ peptides produced [49]. The proportion of Aβ fragments differ between the two mutation groups so the composition and kinetics of the toxic agent is also likely to differ [49]. Mechanistically, the apparent dif-


Fig. 1. Effect-map comparing grey matter volume in the APP and PSEN1 carriers. Regions showing reduced grey matter in the APP group are shown in blue and in PSEN1 in red. Differences between the two adjusted group-means are expressed as a percentage of their average and are overlaid on a mean study-specific template.
Fig. 2. Effect-map comparing white matter volume in the APP and PSEN1 carriers. Regions showing reduced white matter in the APP group are shown in blue and in PSEN1 in red. Differences between the two adjusted group-means are expressed as a percentage of their average and are overlaid on a mean study-specific template.
Fig. 3. Grey (A) and white (B) matter reductions compared with controls. SPM results for (top) APP and (bottom) PSEN1 mutation carriers are shown overlaid on a mean study-specific template and are corrected for multiple comparisons using the false discovery rate (FDR <5%).

Figures suggest that regional atrophy differences in APP and PSEN1 subjects could reflect additional, non-AbetaPP related damage in PSEN1 mutation carriers. Alternatively, effects on AbetaPP metabolism of APP and PSEN1 mutations could be subtly different in vivo. It is noteworthy both that AbetaPP levels are highest in the entorhinal pathway [50] and that AbetaPP expression is upregulated by damage [51]. Under these circumstances, one could envisage that a mutant APP gene would initiate a more local and self-propagating hippocampal lesion in the early stages of disease, whereas the more widespread distribution of presenilin might predict a more global effect.

When we examined whether these regional atrophy patterns were reflected by differences in the neuropsychological profiles, we found greater memory impairment in APP, albeit not statistically significantly. This provides some clinical support for the finding of greater involvement of the medial temporal and limbic regions in these patients compared with the PSEN1 group. Conversely, compared with APP, PSEN1 patients showed greater impairment in the non-memory domains, although again non-significantly, which is also consistent with the imaging findings of greater cortical loss in PSEN1. The lack of significance between the groups may be due to a lack of power or
possibly because many patients performed at floor on these tests. Hence, though the neuropsychological data are supportive of the double dissociation suggested by the imaging results, more sensitive tests in larger cohorts are required for confirmation.

It is possible that differences between these groups are due to factors other than the specific mutation, e.g., age, disease severity, or other genetic modifiers. However, the MMSE scores suggest that the patient groups were similarly affected, and the apparent double dissociation between the most atrophied brain areas could not be explained by a simple severity effect. Previous work has shown regional specificity in the decline in brain volume with age [52], and it can be hypothesized that the neurotoxic effects of the amyloid cascade increase with age, perhaps particularly so in the hippocampus which may have greater age-related vulnerability to Aβ oligomers [53]. The older APP subjects may be exhibiting this effect, but may not be as far advanced as the PSEN1 group in terms of cortical loss. Frisoni and colleagues report more medial temporal lobe atrophy in late-onset AD compared with non-familial early onset AD [54], supporting the idea that there may be additional age-related atrophy in this region. However, whole-brain volumes also decline with increasing age [52, 55]. By including age as a covariate in the models for VBM and cortical thickness, we have tried to minimize the effects of this potential confound. Nonetheless larger studies are needed to confirm this.

Our study has demonstrated differences in the extent of atrophy and its distribution between APP and PSEN1 mutations, suggesting that studies comparing FAD and SAD groups may differ depending on the proportions of APP and PSEN1 subjects included. The finding of greater atrophy in FAD compared with SAD reported previously may largely reflect the effect of PSEN1, the most common cause of FAD [19]. For example, a recent study suggested that visual assessment of hippocampal atrophy was insensitive in the identification of presymptomatic and affected subjects with FAD [56]. However, this sample was dominated by PSEN1 mutation carriers who may have relative hippocampal sparing. Our data suggest that it may therefore be important for individual mutation groups to be considered separately when drawing conclusions about the atrophy pattern and presentation of FAD.

Phenotypic differences between APP and PSEN1 also have implications for generalization from animal models incorporating these mutations.
Fig. 5. Cortical thickness reductions for APP (A) and PSEN1 (B) compared with controls. Results are rendered on a mean study-specific cortical surface and are corrected for multiple comparisons using the false discovery rate (FDR < 5%).

A limitation of the study is that while there are several lines of imaging and neuropsychological evidence to support a double dissociation in regional disease burden between the groups, most direct comparisons of the disease groups did not reveal statistically significant differences. Studying individuals with genetically
confirmed AD allows definitive diagnosis during life; however, the numbers of subjects available for study are limited and despite the lack of co-morbidity a number of features may still lead to disease heterogeneity which reduces the power to detect group differences. First, different mutations within the same gene may exert different effects. Ideally, analysis of mutation subtypes would be performed; however insufficient data was available to make such comparisons meaningful. Second, other genetic factors may modify the disease, perhaps interacting with the different mutations. A recent report describes different clinical phenotypes in siblings carrying the same mutation type. Previous studies have highlighted volume differences prior to the onset of disease in order to identify the earliest focus of neurodegeneration in each mutation type. Previous studies have highlighted volumetric change in these subjects years before clinical onset [25, 34], and a recent study from the DIAN consortium used a variety of imaging techniques to estimate that there were alterations in cerebrospinal fluid markers, brain glucose metabolism, and amyloid deposition decades before the expected age of onset [62] in a large FAD cohort. We await further studies using similar techniques to explore the differential effects of the specific mutations within such FAD cohorts.

In conclusion, we suggest that FAD subjects with the APP and PSEN1 mutations at a similar disease stage may have differing patterns of regional atrophy. The APP subjects had significantly smaller hippocampal volumes than the PSEN1 group and a trend for larger whole-brain and grey matter volumes. This pattern of more localized medial temporal atrophy in AD subjects carrying the PSEN1 e4 allele compared with non-carriers [38, 59], while the APOE e2 allele may have a protective effect regarding time of disease onset. Possession of an e4 allele appears to hasten the onset of the disease in APP mutation carriers, but not so clearly in individuals with mutations in the PSEN1 gene [60]. Examination of other mutations, such as PSEN2 and modulating factors such as APOE status and newly identified risk factors such as clusterin and phosphatidylinositol binding clathrin assembly protein [61] may therefore provide further information about disease heterogeneity. Such analyses will require multi-center collaborations, such as the Dominantly Inherited Alzheimer Network (http://www.dian-info.org/) [7].

In this study, we aimed to match the mutation groups according to disease severity; we used the MMSE, a widely used clinical assessment tool for AD. Nevertheless, it is possible that the MMSE may not fully capture the severity of the disease when individuals have different phenotypic presentations. Other rating scales, such as the Clinical Dementia Rating, might provide additional information regarding disease severity but unfortunately were not available for this cohort. It would also be interesting to examine volumetric brain changes prior to the onset of disease in order to identify the earliest focus of neurodegeneration in each mutation type. Previous studies have highlighted volumetric change in these subjects years before clinical onset [25, 34], and a recent study from the DIAN consortium used a variety of imaging techniques to estimate that there were alterations in cerebrospinal fluid markers, brain glucose metabolism, and amyloid deposition decades before the expected age of onset [62] in a large FAD cohort. We await further studies using similar techniques to explore the differential effects of the specific mutations within such FAD cohorts.

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Table 2

<table>
<thead>
<tr>
<th></th>
<th>Control (n=18)</th>
<th>APP (n=10)</th>
<th>PSEN1 (n=10)</th>
<th>APP-PSEN1 *</th>
</tr>
</thead>
<tbody>
<tr>
<td>Brain</td>
<td>1204 (1180, 1247)</td>
<td>1110 (1066, 1154)</td>
<td>1085 (1025, 1092)</td>
<td>52 (−3.8, 108) p=0.065</td>
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<tr>
<td>GM</td>
<td>468 (436, 500)</td>
<td>507 (493, 522)</td>
<td>509 (515, 579)</td>
<td>44 (−2.9, 91) p=0.047</td>
</tr>
<tr>
<td>WH</td>
<td>480 (457, 503)</td>
<td>460 (442, 479)</td>
<td>438 * (414, 462)</td>
<td>22 (−8.8, 35) p=0.15</td>
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<tr>
<td>Cingulate</td>
<td>16.9 (15.3, 18.7)</td>
<td>13.1 (11.9, 14.4)</td>
<td>11.3 (11.8, 14.9)</td>
<td>−0.20 (−2.15, 1.7) p=0.83</td>
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<td>Hippocampus</td>
<td>5.7 (5.45, 6.0)</td>
<td>4.39 (3.87, 4.91)</td>
<td>5.09 * (4.4, 5.55)</td>
<td>−0.70 (−1.3, −0.1) p=0.042</td>
</tr>
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</table>

* Differences between the APP and PSEN1 group means as a percentage of their average.

** Only n=9 APP subjects had data on cingulate volume.

† Controls > AD groups (p<0.05).

‡ Controls > AD groups (p<0.01).

− Controls > AD groups (p<0.001).
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references


